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(54) Title: PEPTIDES AND ANTI-SENSE PEPTIDES WITH BROAD NEOPLASTIC SPECIFICITY

## (57) Abstract

An analogue peptide comprises the variable regions of the light or heavy chains of an antibody of a first species selectively binding to a tumor antigen, such as a carcinoma antigen, has 1 to 46 amino acids of the framework regions per chain substituted with amino acids such as those present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 variable region CDRs per chain and optionally flanking regions thereof of 1 to 10 or more amino acids, alone or with an N-terminal fragment of 1 to 10 or more amino acids, combinations or mixtures thereof. The polypeptide may also comprise an effector agent and/or be glycosylated, and is presented as a composition with a carrier. The analogue peptides are used in diagnostic kits for neoplasms such as carcinomas and methods for *in vivo* imaging and treating a primary or metastasized neoplasm such as a carcinoma, and *in vitro* diagnosing a neoplasm, *ex vivo* purging neoplastic cells from a biological fluid. RNAs and DNAs encode the analogue peptide, and a hybrid vector carrying the nucleotides and transfected cells express the peptides and a method produces the analogue peptide. An anti-idiotype polypeptide comprises polyclonal antibodies raised against an antineoplastic antibody or the analogue peptide of this invention, monoclonal antibodies thereof, Fab, Fab', (Fab')<sub>2</sub>, CDR, variable region, or analogues or fragments thereof, combinations thereof with an oligopeptide comprising a TRP trimer, tandem repeats thereof, or combination or mixtures thereof. An anti-idiotype hybrid polypeptide with an effector agent and the anti-idiotype polypeptide, an anti-neoplastic vaccine, an anti-neoplastic vaccination kit, a method of vaccinating against neoplasms such as carcinomas and a method of lowering the serum concentration of a circulating antibody or polypeptide are provided.

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PEPTIDES AND ANTI-SENSE PEPTIDES WITH BROAD NEOPLASTIC  
SPECIFICITY

BACKGROUND OF THE INVENTION

Field of the invention

This invention relates to the *in vitro* and *in vivo* diagnosis, immunization, and therapy of neoplastic tumors, particularly carcinomas, by means of specifically targeted analogue peptides comprising amino acid sequences encompassing the complementarity determining regions (CDRs) of a different species, and analogues of the variable (F<sub>v</sub>) region of anti-carcinoma antibodies, among others. The carcinoma specific peptides for use in one species, e.g., humans, are provided as a single amino acid chain having the specificity of F<sub>v</sub> regions obtained in different species, e.g., murine antibody F<sub>v</sub> regions of the light or heavy chains, or as paired chains. These peptides are provided either by themselves or bound to other molecules such as synthetic polymers or oligopeptides resulting in sequences of mixed species, and more particularly analogues of human/non-human chimeric antibodies or other polymeric constructs. The analogue peptides comprise sequences derived from the variable regions of heterologous antibodies specific for, e.g., human carcinoma antigens that elicit a lesser immunological response in humans than the whole heterologous antibodies. The anti-idiotype polypeptides and their analogues are suitable for immunizing humans or other animals against carcinoma. Polynucleotide segments encoding the analogue peptide and anti-idiotype polypeptides, and hybrid vectors and transfected host cells carrying the segments are useful for preparing the peptides disclosed herein.

Description of the Background

Carcinomas result from the carcinogenic transformation of cells of different epithelia. Two of the most damaging characteristics of carcinomas are their uncontrolled growth and their ability to create metastases in distant sites of the host, particularly a human host. It is usually these distant metastases that cause serious consequences to the host, since frequently the primary carcinoma may be, in most cases, removed by surgery. The treatment of metastatic carcinomas, that are seldom removable, depends on irradiation therapy and systemic therapies of different natures. The systemic therapies currently include, but not fully comprise, chemotherapy, radiation, hormone therapy, different immunity-boosting medicines and procedures, hyperthermia and systemic monoclonal antibody treatment. The latter can be labeled with radioactive elements, immunotoxins and chemotherapeutic drugs.

Radioactively labeled monoclonal antibodies were initially used with success in lymphomas and leukemia, and recently in some carcinomas. The concept underlying the use of labeled antibodies is that the labeled antibody will specifically seek and bind to the carcinoma and, the radioactive element, through

its decay, will irradiate the tumor *in situ*. Since radioactive rays travel some distance in tumors it is not necessary that every carcinoma cell bind the labeled antibody. The specificity of the monoclonal antibodies will permit a selective treatment of the tumor while avoiding the irradiation of innocent by-stander 5 normal tissues, that could be dose limiting. Chemotherapy produces serious toxic effects on normal tissues, making the chemotherapy of carcinomas less than desirable, and the use of radiolabeled monoclonal antibodies a valid alternative.

Non-human antibodies raised against human epitopes have been used for 10 the diagnosis and therapy of carcinomas as is known in the art. Also known are the methods for preparing both polyclonal and monoclonal antibodies. Examples of the latter are BrE-2, BrE-3 and KC-4 (e.g., US patent Nos. 5,077,220; 5,075,219 and 4,708,930..

The KC-4 murine monoclonal antibody is specific to a unique antigenic 15 determinant, the "antigen", and selectively binds strongly to neoplastic carcinoma cells and not to normal human tissue (U.S. Patent No. 4,708,930 to Coulter). The antigen appears in two forms in carcinoma cells, only the smaller of these forms being expressed in the cell membrane. The larger form appears only in the cytoplasm and has an approximate 490 Kdalton molecular weight 20 (range of 480,000-510,000). The second form occurs at a higher density of expression, is found both in the cytoplasm and the membrane of carcinoma cells and has an approximate 438 Kdalton molecular weight (range of 390,000-450,000) as determined by gel electrophoresis with marker proteins of known molecular weights. Labeled KC-4 was applied to the diagnosis and medical 25 treatment of various carcinomas, particularly adenocarcinoma and squamous cell carcinoma regardless of the human organ site of origin.

The BrE-3 antibody (Peterson et al., *Hybridoma* 9:221 (1990); US patent No. 5,075,219) was shown to bind to the tandem repeat of the polypeptide core 30 of human breast epithelial mucin. When the mucin is deglycosylated, the presence of more tandem repeat epitopes is exposed and the binding of the antibody increases. Thus, antibodies such as BrE-3 bind preferentially to neoplastic carcinoma tumors because these express an unglycosylated form of the breast epithelial mucin that is not expressed in normal epithelial tissue. This preferential binding combined with an observed low concentration of epitope for 35 these antibodies in the circulation of carcinoma patients, such as breast cancer patients, makes antibodies having specificity for a mucin epitope highly effective for carcinoma radioimmunotherapy. A <sup>90</sup>Y-BrE-3 radioimmunoconjugate proved highly effective against human breast carcinomas transplanted into nude mice. Human clinical studies showed the <sup>90</sup>Y-BrE-3 radioimmunoconjugate to

considerably reduce the size of breast tumor metastases without any immediate toxic side effects. Moreover, an  $^{111}\text{In}$ -BrE-3 radioimmunoconjugate was successfully used for imaging 15 breast cancer patients, providing excellent tumor targeting in 13 out of 15 of the patients. Out of all the breast tumor metastases occurring in another study, 86% were detected by  $^{111}\text{In}$ -BrE-3. Unfortunately, 2 to 3 weeks after treatment, the patients developed a strong human anti-murine antibody (HAMA) response that prevented further administration of the radioimmunoconjugate. The HAMA response, which is observed for numerous murine monoclonal antibodies, precludes any long-term administration of murine antibodies to human patients. Similarly, other heterologous antibodies, when administered to humans, elicited similar antibody responses. The anti-heterologous human response is, thus, a substantial limiting factor hindering the successful use of heterologous monoclonal antibodies as therapeutic agents, which could, otherwise, specifically annihilate breast carcinomas, causing little or no damage to normal tissue and having no other toxic effects.

Chimeric antibodies are direct fusions between variable domains of one species and constant domains of another. Murine/human chimeric antibodies prepared from other types of B cells binding to other types of antigenic determinants have been shown to be less immunogenic in humans than whole murine antibodies. These proved to be less immunogenic but still in some cases an immune response is mounted to the rodent variable region framework region (FR). A further reduction of the "foreign" nature of the chimeric antibodies was achieved by grafting only the CDRs from a rodent monoclonal into a human supporting framework prior to its subsequent fusion with an appropriate constant domain, (European Patent Application, Publication No. 239,400 to Winter; Riechmann, et al., *Nature* 332:323-327 (1988)). However, the procedures employed to accomplish CDR-grafting often result in imperfectly "humanized" antibodies. That is to say, the resultant antibody loses avidity (usually 2-3 fold, at best).

The ligand binding characteristics of an antibody combining site are determined primarily by the structure and relative disposition of the CDRs, although some neighboring residues also have been found to be involved in antigen binding (Davies, et al., *Ann. Rev. Biochem.* 59:439-473 (1990)).

The technologies of molecular biology have further expanded the utility of many antibodies by allowing for the creation of class switched molecules whose functionality has been improved by the acquisition or loss of complement fixation. The size of the bioactive molecule may also be reduced so as to increase the tissue target availability of the antibody by either changing the class

from an IgM to an IgG, or by removing most of the heavy and light chain constant regions to form an F<sub>v</sub> antibody. Common to all of these potentially therapeutic forms of antibody are the required complementary determining regions (CDRs), which guide the molecule to its ligand, and the framework residues (FRs) which support the CDRs and dictate their disposition relative to one another. The crystallographic analysis of numerous antibody structures revealed that the antigen combining site is composed almost entirely of the CDR residues arranged in a limited number of loop motifs. The necessity of the CDRs to form these structures, combined with the appreciated hypervariability of their primary sequence, leads to a great diversity in the antigen combining site, but one which has a finite number of possibilities. Thus, its hypermutability and the limited primary sequence repertoire for each CDR would suggest that the CDRs derived for a given antigen from one species of animal would be the same derived from another species. Hence, they should be poorly immunogenic, if at all, when presented to a recipient organism.

Accordingly, there is still need for a product of high affinity and/or specificity for carcinoma antigens suitable for the detection and therapy of carcinomas which elicits a lesser antibody response than whole non-human antibodies or chimeric antibodies containing, for instance the entire non-human variable region.

#### SUMMARY OF THE INVENTION

This invention relates to an analogue peptide and its glycosylated derivative which specifically and selectively bind to the human mammary fat globule (HMFG) antigen and to an antigen found on the surface or in the cytoplasm of tumor cells such as carcinoma cells or that is released by the cells, the analogue peptide consisting essentially of at least one variable region of the light or heavy chains of an antibody of a first species having affinity and specificity for an antigen found on the surface or the cytoplasm of a carcinoma cell or released by the cell, wherein preferably about 1 to at least 46 amino acids in the FR are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain or 1 to 3 CDRs plus flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids, combinations thereof or combinations thereof with other variable regions or analogues thereof, wherein each analogue peptide is operatively linked to at least one other peptide or analogue thereof, or mixtures thereof.

Also provided herein are a fusion protein and a hybrid polymer comprising the analogue of the invention, the corresponding DNAs encoding them, hybrid vector thereof, transfected host thereof, and RNA.

This invention also encompasses a method of producing an analogue 5 peptide or hybrid analogue peptide by recombinant technology, in vitro methods of diagnosing and immunohistochemistry of tissue slices, an ex vivo method of purging neoplastic cells, and in vivo methods for imaging and therapy of neoplasias such as carcinomas.

Also disclosed herein are anti-idiotype polypeptides comprising polyclonal 10 antibodies raised against the analogue peptide of this invention, analogues thereof, monoclonal antibodies thereof, fragments thereof selected from the group consisting of Fab, Fab', (Fab')<sub>2</sub>, CDRs, variable regions and analogues thereof as described above, combinations thereof operatively linked to one another, an anti-carcinoma vaccine, a vaccination kit, a method of vaccinating 15 against neoplasias including carcinomas, and a method of lowering the serum concentration of circulating anti-tumor antibody with the anti-idiotype polypeptide of this invention.

Other objects, advantages and features of the present invention will become apparent to those skilled in the art from the following discussion.

## 20

DESCRIPTION OF THE PREFERRED EMBODIMENTS

This invention arose from a desire by the inventors to improve on antibody technology suitable for use in diagnostic, prognostic, vaccine and therapeutic applications in the field on neoplasias and cancer. The present invention will be described for application to humans. However, it is also 25 suitable for use in other species. The monoclonal antibodies obtained up to the present time have been prepared by fusing immortalized cell lines with B-cells of non-human origin such as murine, rat, rabbit, goat, and the like. Many of these hybridomas can produce large quantities of monoclonal antibodies that have desirable binding properties such as high affinity and/or specificity and selectivity 30 for carcinoma antigens of a species, e.g., for human carcinoma antigens. However, in general, antibodies from other species, e.g., non-human antibodies, may only be administered once to a subject of a predetermined species, e.g., humans, due to the detrimental effects they produce. This is true for most heterologous antibodies being administered to mammalian animals. For example, 35 the repeated administration of murine antibodies to a human subject elicits a strong human anti-murine antibody (HAMA) response, which precludes their further utilization as therapeutic agents in humans. These non-human antibodies initiate an immediate adverse reaction in many human patients and are, thus,

rendered ineffective for further administration as therapeutic agents. In some cases, non-human-human chimeric antibodies and non-human CDR "grafted"-human antibodies may have lower affinity and/or specificity for their antigens than the corresponding non-human antibody. On the other hand, human 5 monoclonal hybridoma cell lines have not been very stable and have, therefore, not been suitable for the large scale, repeated production of monoclonal antibodies.

The present invention is applicable to the manufacture of analogue peptides having a high number of amino acids in the respective positions found 10 in the endogenous antibodies, and where at least the CDRs are of another xenogeneic species.

The present inventors, thus, have undertaken the preparation of antitumor non-human CDRs and non-human variable regions of antibodies, having affinity and specificity for an antigen found on the surface or the 15 cytoplasm of a human neoplastic cell such as a carcinoma cell or that is released by the cell, wherein about 1 to 46 amino acids in the framework region (FR) are substituted per chain with amino acids present in equivalent positions in other human antibodies, or fragments thereof comprising 1 to 3 CDRs per chain or 1 to 3 CDRs per chain plus flanking regions thereof, each of about 1 to 10 or more 20 amino acids, alone or plus an N-terminal fragment of about 1 to 10 or more amino acids, to lower or even circumvent the anti-xenogeneic response. To preserve substantial binding specificity in the molecules intended for use in humans, the present invention utilizes CDRs and/or analogues of varying lengths of the variable regions of light and/or heavy chains of antibodies of other species 25 such as murine, rat, rabbit, goat, equine, primate, bovine, and guinea pig antibodies, among others. Heterologous human CDRs and/or analogues of their variable regions may be utilized are intended for use in other species.

The present inventors have found, surprisingly, that these analogue antibody fragments substantially preserve the binding, specificity and selectively 30 characteristics of the whole non-human antibody while eliciting a lesser detrimental immunological response. However, the simple preservation of the binding region of an antibody does not by itself ensure that the binding characteristics of the antibody will be maintained. Antibodies are glycopolypeptides that are folded into specific conformations. When the 35 glycoside portion of the molecule or portions of the amino acid sequence are perturbed or excised, the folding pattern of the molecule may be perturbed. Thus, any deletion or modification of the sequence of an antibody must be made taking into consideration that its folding-dependent properties may be diminished or even obliterated if the folding is substantially affected, even though the amino

acid sequences involved in the binding of the antigen are preserved.

The present inventors selected the following strategy for the preparation and manufacture of the analogue and hybrid peptides of this invention. The cDNAs that encode the variable chains of an antibody may be obtained by 5 isolation of mRNA from a hybridoma cell and reverse transcription of the mRNA, amplification of the cDNA by polymerase chain reaction (PCR) and insertion of the DNA into a vector for optional sequencing, and for restriction enzyme cutting. The cDNAs encoding the CDRs or variable chain ( $F_v$ ) region fragments of the light ( $V_L$ ) and heavy ( $V_H$ ) chains of an antibody having affinity and 10 specificity for a carcinoma cell antigen may be reverse transcribed from isolated mRNA. The cDNAs encoding the CDRs may then be ligated to other segments encoding neighboring sequences, and the variable region cDNAs may then be modified with predesigned primers used to PCR amplify them or synthesized de novo, cloned into a vector optionally carrying DNA sequences encoding, e.g., 15 a constant region(s), optionally sequenced, and then transfected into a host cell for expression of the analogue gene product. The binding specificity characteristics of the analogue peptides may then be determined and compared to those of the whole antibodies.

X-ray crystallographic studies demonstrate that the framework 20 structures of the  $F_v$  of different antibodies assume a canonical structure regardless of the species of origin, amino acid sequence, or ligand specificity. This is generally taken as evidence that the ligand-binding characteristics of an antibody combining site are determined primarily by the structure and relative disposition of the CDRs, although some neighboring framework residues may 25 also be involved in antigen-binding. Thus, if the fine specificity of an antibody is to be preserved, its CDR structures, and probably some of the neighboring residues, their interaction with each other and with the rest of the variable domains, must also be maintained. These crystallographic studies point to the possible need for retaining most, if not all, of the many interior and inter-domain 30 contact residues since the structural effects of replacing only a few of them cannot be predicted.

While at first the necessity of keeping these amino acids might seem to defeat the goal of decreasing immunogenicity by "humanization", the actual 35 number of amino acids that must be retained is usually small because of the striking similarity between human and murine variable regions. Moreover, many, if not most, of the retained amino acids possess side chains that are not exposed on the surface of the molecule and, therefore, may not contribute to its antigenicity. Clearly, it is most of the exposed amino acids that are good candidates for substitution since it is these amino acids that are exposed to the

immunological environment of a mammal and may form epitopes of increased immunogenicity.

The challenge in humanizing the variable regions of a non-human antibody, e.g., a murine antibody, thus begins with the identification of the 5 "important" xenogeneic amino acids. "Important" amino acids are those, for example, that are involved in antigen binding, contact the CDRs and the opposite chains, and have buried side-chains. Ideally, these residues might be identified from a well characterized three-dimensional structure. However, when direct structural data are not available, it is, fortunately, still possible to predict the 10 location of these important amino acids by analyzing other related antibody structures, especially those whose variable light and heavy regions belong to the same class. The classes of variable regions can be determined from their amino acid sequence.

A method by which these important amino acids are identified has been 15 described for the case of the amino acids with buried side chains by Padlan, E.A. (Padlan, E.A., "A Possible Procedure for Reducing the Immunogenicity of Antibody Variable Domains While Preserving Their Ligand-Binding Properties", Molecular Immunology, 28:489-494 (1991)). In the present case, various antibody variable region structures were compared using a computer program 20 that determines the solvent accessibility of the framework residues as well as their contacts with the opposite domain as described by Padlan, E.A. (1991), supra. Surprisingly, a close examination of the fractional solvent accessibility reveals a very close similarity in the exposure patterns of the  $V_H$  and the  $V_L$  domains. Put in simple terms, regardless of the particular antibody in question, 25 and of its amino acid sequence, the buried residues occupy similar relative positions in most antibodies.

A similar analysis can be done by computer modeling, to determine which amino acids contact the CDRs and which contact the opposite domain. At this point, the Fab structures that are currently in the Protein Data bank 30 (Bernstein, F.C., et al., J. Mol. Biol. 112:535-542 (1977)) may be examined to determine which FRs are probably important in maintaining the structure of the combining site. Thus, after a close inspection of many high resolution three-dimensional structures of variable regions, the positions of all important framework amino acids, that is, those that contact the CDRs, the opposite 35 domain, and those whose side chains are inwardly pointed, may be tabulated. Keeping these amino acids, as well as those from the CDRs, and finally those FR amino acids that may be involved in ligand binding, should insure to a great extent the preservation of affinity. The precise identification of FR amino acids that are involved in ligand-binding cannot be generalized since it varies for

different antibodies. Nevertheless, conservative decisions can be made to preserve the amino acids located in FR that have a high probability of contacting the antigen. These regions are located immediately adjacent to the CDRs and at the N-terminus of both chains, because the surfaces of these regions are contiguous with the CDR surfaces.

Surprisingly, it is possible to keep all of these important amino acids in a heterologous humanized antibody and still increase dramatically the similarity with a human consensus sequence. That is, the final number of amino acids with murine identities differing from human identities that are kept is typically small.

10 This is usually possible because human frameworks that are similar to the murine frameworks, especially at the positions of the important amino acids, can be found. This is because many of the important amino acids have the same identities in both murine and human antibodies.

All the amino acids that are determined to be not important by the method described above may be completely replaced by their corresponding human counterparts. The surface of the finally humanized antibody should look very much like that of a human antibody except for the antigen binding surfaces. The original shape of those binding surfaces, however, is maintained by leaving the internal composition of the antibody intact, preserving inter-domain contacts and by keeping very few key amino acids that contact the CDRs.

15 a) Choosing the Best Human Framework to Use in the "Humanization" of an Antibody When Its Structure Is Known  
At the present time, there are 11 Fab structures for which the atomic coordinates are known and have been placed in the Protein Data Bank as shown in Table 1 below, 2 from human and 9 from murine antibodies.

Table 1: Fab Structures for Which Coordinates are in the Protein Data Bank

	ANTIBODY	RESOLUTION (Å)	R-VALUE	PDB CODE
BOHAN:	NEWK KOL	2.0 1.9	0.16 0.189	3FAB 2FB4
MURINE:	McPC603 JS39 HyHEL-5 HyHEL-10 R19-9 4-4-20 36-71 B1312 D1.3	2.7 1.95 2.54 3.0 2.8 2.7 1.85 2.8 2.5	0.225 0.194 0.245 0.24 0.30 0.215 0.248 0.197 0.184	1MCP 2FBJ 2HFL 3HEM 1F19 4FAB 6FAB 1IGF 1FOL

The contacts between side chains in the variable domains of the 11 Fab's have been collected and are presented in Tables 2 to 4 below. The framework (FR) amino acids (aa's) in the  $V_L$  domains that contact CDRs are listed in Table 2 below.

5 **Table 2:**  $V_L$  Framework Residues That Contact CDR Residues in Fab's of Known Three-Dimensional Structure

POSITION	ANTIBODY										
	JS39 McPC603 MyHEL-10 MyHEL-5 R19.9 4-4-20 36-71 B1312 D1.3 NEWH KOL										
1	GLU(2)	ASP(5)	ASP(10)	ASP(3)	ASP(8)	ASP(4)		ASP(11)			
2	ILE(11)	ILE(15)	ILE(17)	ILE(13)	ILE(5)	VAL(9)	ILE(20)	VAL(9)	ILE(10)	SER(3)	
3		VAL(3)	VAL(2)	VAL(3)	GLN(2)	VAL(2)	GLN(2)	LEU(5)		VAL(2)	
4	LEU(7)	MET(6)	LEU(6)	LEU(10)	MET(9)	MET(13)	MET(7)	MET(6)	LEU(4)	LEU(6)	
5					THR(1)	THR(2)			THR(4)	THR(1)	
7		THR(1)						SER(6)			
22										CYS(1)	
23	CYS(1)	CYS(1)	CYS(2)	CYS(2)	CYS(1)	CYS(1)	CYS(1)		TRP(4)	TRP(1)	TRP(2)
35	TRP(3)	TRP(2)	TRP(4)		TRP(2)		TRP(6)	TRP(4)	TRP(4)	TYR(14)	TYR(13)
36	TYR(12)	TYR(16)	TYR(8)	TYR(10)	TYR(22)	TYR(13)	TYR(15)	TYR(8)	TYR(14)	TYR(13)	TYR(11)
45					LYS(12)	LYS(5)					
46	PRO(3)	LEU(6)	LEU(4)	ARG(15)	LEU(5)	VAL(14)	LEO(5)	LEU(10)	LEU(6)	LEU(2)	LEU(6)
48	ILE(1)	ILE(1)	ILE(1)				ILE(3)	ILE(2)	VAL(1)		ILE(1)
49	TYR(28)	TYR(29)	LYS(13)	TYR(12)	TYR(40)	TYR(22)	TYR(22)	TYR(16)	TYR(25)		TYR(25)
58	VAL(3)	VAL(3)	ILE(1)	VAL(6)	VAL(6)	VAL(5)	VAL(4)	VAL(5)	VAL(1)	VAL(6)	
60	ASP(1)					ASP(2)	ASP(4)			ASP(2)	
62					PHE(1)	PHE(1)	PHE(1)				
66								LYS(2)	LYS(11)		
67	SER(3)							SER(1)			
69		THR(3)	THR(3)			THR(5)	THR(1)	THR(4)	THR(1)	SER(1)	
70	ASP(2)					ASP(1)	ASP(6)			SER(2)	
11	TYR(14)	PHE(23)	PHE(17)	TYR(17)	TYR(24)	PHE(17)	TYR(17)	PHE(19)	TYR(16)	ALA(3)	ALA(4)
88	CYS(1)		CYS(2)		CYS(1)	CYS(1)	CYS(1)	CYS(1)	CYS(2)		CYS(1)
98	PHE(8)	PHE(8)	PHE(10)	PHE(5)	PHE(8)	PHE(4)	PHE(8)	PHE(14)	PHE(3)	PHE(7)	

Those FR in the  $V_H$  domains that contact CDRs are listed in Table 3  
10 below.

Table 3:  $V_H$  Framework Residues That Contact CDR Residues in Fab's of Known Three-Dimensional Structure

POSITION	ANTIBODY										
	JS39 McPC603 MyHEL-10 MyHEL-5 R19.9 4-4-20 36-71 B1312 D1.3 NEWH KOL										
1	VAL(11)	VAL(3)	VAL(6)		VAL(1)		VAL(7)	VAL(3)	VAL(12)		VAL(9)
2	LEU(7)	LEU(5)			LEU(2)	LEU(1)	LEU(1)	LEU(1)	LEU(1)		LEU(1)
21	THR(2)	VAL(6)				ALA(1)					
22	PHE(3)	PHE(2)			TYR(14)	TYR(13)	PHE(24)	TYR(4)	PHE(4)	PHE(3)	
23					TYR(23)	TYR(20)	TYR(21)	TYR(24)	TYR(1)	TYR(1)	TYR(2)
26	ASP(3)	THR(3)			THR(6)	THR(6)	THR(2)	THR(3)	SEK(1)	ILE(2)	
27	PHE(4)	PHE(4)			PHE(20)	PHE(7)	PHE(13)	PHE(6)	PHE(3)	LEU(1)	TYR(4)
30					THR(6)	SEK(7)			ASP(6)		
34							TAP(2)				
37	VAL(3)				VAL(1)			VAL(1)	VAL(2)	VAL(1)	VAL(3)
38	ARG(11)	ARG(2)	ARG(4)		LYS(2)	LYS(1)	ARG(4)	LYS(2)	ARG(1)		ARG(3)
40					ARG(11)						
44	GLU(3)	GLU(4)	GLU(11)	GLU(2)	GLU(3)	GLU(4)	GLU(5)	GLU(4)	GLU(1)		GLU(1)
45	TRP(21)	TRP(29)	TYR(20)	TRP(21)	TRP(23)	TRP(24)	TRP(25)	TRP(13)	TRP(22)	TRP(15)	
46	ILE(11)	ILE(4)	ILE(2)	ILE(13)	VAL(11)	ILE(9)	VAL(3)	LEU(1)	ILE(2)	VAL(11)	ILE(2)
47					ALA(2)		ALA(2)		ALA(2)		ALA(2)
48					ALA(2)		ALA(3)		ALA(2)		ALA(2)
66					ARG(11)		ARG(5)		ARG(2)		ARG(11)
67	PHE(4)	PHE(1)	ILE(9)	ALA(1)			PHE(11)	THR(5)	PHE(12)	LEU(6)	VAL(2)
68					ILE(11)		THR(11)				TYR(2)
69	VAL(6)	VAL(6)	PHE(12)	LEU(5)	ILE(20)	LEU(6)	ILE(11)	ILE(8)	MET(1)	ILE(9)	
70	ILE(11)	ARG(14)	ARG(21)	ALA(3)	VAL(4)	ARG(6)	VAL(6)	ASG(3)	LYS(4)	ARG(9)	
71	ASP(3)	THR(3)				ASP(3)					
72	LEU(11)	LEU(7)	TYR(9)	ALA(1)	ALA(1)	VAL(2)	ALA(1)	LEU(6)	VAL(4)	PHE(5)	LEU(5)
73						LEU(1)					
74								HGT(1)	LEU(1)		
75											
76											
77											
78											
79											
80											
81											
82											
83											
84											
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100											

the FR amino acids, that contact the opposite domain and which presumably are the ones mainly responsible for the quaternary structure of the F<sub>v</sub> domains are listed in Table 4 below.

5 **Table 4:** Framework Residues That Contact Framework Residues in the Opposite Domain in Fabs of Known Three-Dimensional Structure

IN V <sub>L</sub> :		ANTIBODY									
POSITION	J539 McPC603 MyHEL-10 MyHEL-5 R19.9	4-4-20	36-71	B13Z2	D1.3	NEWM	KOL				
36	TYR(3)	TYR(1)	TYR(3)	TYR(5)	TYR(1)	TYR(7)	TYR(1)	TYR(7)	TYR(5)		
38	GLN(10)	GLN(9)	GLN(9)	GLN(5)	GLN(5)	GLN(6)	GLN(12)	GLN(6)	GLN(7)	GLN(10)	
43	SER(9)	PRO(1)	SER(8)	SER(5)	THR(3)	SER(3)	SER(2)	ALA(5)	ALA(11)		
46	PRO(10)	PRO(14)	PRO(6)	PRO(11)	PRO(7)	ILE(20)	PRO(16)	PRO(7)	PRO(13)		
48	PRO(3)										
55		HET(2)		THR(3)			VAL(1)		ASP(12)		
67	TYR(6)	TYR(4)	PHE(6)	TYR(2)		PHE(5)	TYR(10)	TYR(8)	TYR(6)	TYR(6)	
98	PHE(11)	PHE(8)	PHE(7)	PHE(12)	PHE(12)	PHE(8)	PHE(13)	PHE(12)	PHE(10)	PHE(15)	
100	ALA(2)										

  

IN V <sub>H</sub> :		ANTIBODY									
POSITION	J539 McPC603 MyHEL-10 MyHEL-5 R19.9	4-4-20	36-71	B13Z2	D1.3	NEWM	KOL				
31	VAL(4)	ILE(2)	VAL(1)	VAL(4)	VAL(2)	VAL(1)	VAL(2)	VAL(4)	VAL(1)	VAL(4)	
38	GLN(10)	GLN(4)	LYS(8)	GLN(3)	GLN(5)	GLN(3)	GLN(6)	GLN(10)	GLN(6)	GLN(4)	GLN(7)
43		ASN(4)									
44		ARG(2)									
45	LEU(13)	LEU(12)	LEU(8)	LEU(14)		LEU(8)	LEU(11)	LEU(13)	LEU(14)	LEU(11)	LEU(16)
47	TRP(1)		TYR(2)		TRP(2)				TRP(3)	TRP(2)	
51	TYR(6)	TYR(4)	TYR(3)	TYR(8)	PHE(3)	TYR(2)	PHE(4)	TYR(3)	TYR(5)	TYR(3)	
103	TRP(11)	TRP(13)	TRP(16)	TRP(11)	TRP(4)	TRP(18)	TRP(24)	TRP(22)	TRP(19)	TRP(8)	TRP(15)
105	GLN(5)										

The buried, inward-pointing FR amino acids in the V<sub>L</sub> domains, i.e., those which are located in the domain interior, are listed in Table 5 below.

10 **Table 5:** Inward-Pointing, Buried Framework Residues in the V<sub>L</sub> of Fabs of Known Three-Dimensional Structure

POSITION		ANTIBODY									
4	J539 McPC603 MyHEL-10 MyHEL-5 R19.9	4-4-20	36-71	B13Z2	D1.3	NEWM	KOL				
2	ILE	ILE	ILE	ILE	ILE	VAL	ILE	VAL	ILE		
4	LEU	NEU	LEU	NEU	NEU	NEU	NEU	NEU	NEU	LEU	LEU
6	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	
11	TYR	LEU	LEU	TYR	LEU	LEU	ILE	LEU	ILE	VAL	ALA
13	ALA	ALA	VAL	ALA	ALA	VAL	ALA	VAL	ALA		
15	VAL	VAL	VAL	VAL	VAL	VAL	ALA	VAL	VAL	VAL	
21	ILE	NEU	ILE	NEU	ILE	ILE	ILE	ILE	ILE	ILE	
23	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	
35	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	
37	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	GLN	
47	TRP	LEU	LEU	TRP	LEU	LEU	LEU	LEU	LEU	LEU	
49	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	
55	VAL	VAL	ILE	VAL	VAL	VAL	VAL	VAL	VAL	VAL	
61	ARG	ARG	ARG	ARG	ARG	ARG	ARG	ARG	ARG	ARG	
63	PHE	PHE	PHE	PHE	PHE	PHE	PHE	PHE	PHE	PHE	
71	TYR	PHE	PHE	TYR	TYR	TYR	TYR	TYR	TYR	TYR	
73	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	
75	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	
78	NEU	NEU	NEU	NEU	NEU	NEU	NEU	NEU	NEU	NEU	
82	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	
83											
84	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	
86	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	
88	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	
101	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	
104	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	
106	LEU	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	

Those in the V<sub>H</sub> domain are listed in Table 6 below.

Table Inward-Pointing, Buried Framework Residues in the  $V_h$  of Fab's of Known Three-Dimensional Structure

POSITION	JS39	HCFC603	HyHEL-10	HyHEL-5	R13.9	4-4-20	36-71	81332	01.3	NEWH	KOL	ANTIBODY
2	VAL	VAL	VAL	VAL	LEU	LEU	LEU	VAL	VAL	VAL	VAL	VAL
4	LEU	LEU	LEU	VAL	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU
6	GLU	GLU	GLU	GLN	GLU	GLU	GLU	GLU	GLU	GLN	GLN	GLN
9				PRO								
12	VAL	VAL	VAL	MET	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL
14	LEU	LEU	LEU	VAL	MET	VAL	LEU	LEU	LEU	LEU	LEU	LEU
16	LEU	LEU	LEU	ILE	ILE	MET	ILE	ILE	ILE	ILE	ILE	ILE
18							CYS	CYS	CYS	CYS	CYS	CYS
20	LEU	LEU	LEU	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE	ILE
22	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS
24	ALA	THR	VAL	ALA	ALA	ALA	ALA	ALA	ALA	VAL	VAL	SER
26	PHE	PHE	ASP	TIR	TIR	PHE	TIR	PHE	PHE	THR	PHE	PHE
28	PHE	PHE	ILE	PHE	PHE	PHE	PHE	PHE	PHE	PHE	PHE	PHE
30	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP
32	ARG	ARG	ARG	LYS	LYS	ARG	LYS	ARG	ARG	ARG	ARG	ARG
34						SER						
36												
38	ILE	GLU	GLU	GLU	ILE	VAL	ILE	VIS	LEU	ILE	VAL	VAL
40												
42	ALA	ILE	ILE	ILE	ILE	ALA	ALA	ALA	ALA	ALA	ALA	ALA
44												
46	LYS	ARG	ARG	LYS	ILE	VAL	ILE	VAL	ARG	ARG	ARG	ARG
48	PHE	ILE	ALA	VAL	PHE	PHE	PHE	PHE	LEU	VAL	PHE	PHE
50	ILE	VAL	ILE	PHE	ILE	LEU	ILE	ILE	ILE	MET	ILE	ILE
52	ARG	ARG	ALA	VAL	ARG	VAL	ARG	VAL	ARG	VAL	ARG	ARG
54												
56	LEU	TYR	ALA	ALA	VAL	ALA	LEU	VAL	PHE	LEU	LEU	LEU
58	LEU	LEU	ILE	MET	ILE	MET	ILE	ILE	LEU	LEU	LEU	LEU
60	MET	ILE	LEU	LEU	ILE	MET	ILE	MET	MET	LEU	MET	MET
62												
64	VAL	VAL	VAL	VAL	VAL	ILE	ILE	ILE	ILE	VAL	LEU	LEU
66	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP
68												
70	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR
72	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS
74	ARG	ASN	MIS	ARG	ARG	ARG	ARG	ARG	ARG	ARG	ARG	ARG
76	THR	THR	THR	THR	THR	THR	THR	THR	THR	SER	THR	THR
78	VAL	VAL	VAL	LEU	VAL	LEU	LEU	LEU	VAL	VAL	VAL	VAL
80	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL
82	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL

5 From the above, it may be seen that

(1) There are many FR amino acids that either contact the CDRs or the opposite domain, or are found in the domain interior.

(2) These FR amino acids, which could influence the structure of the combining site, and thus the antigen-binding characteristics of an antibody, are different from antibody to antibody.

It is obvious from these results that no one structure can serve as the perfect and sole basis of all "animalization", or in the present example "humanization", protocols. In fact, to "humanize" the 9 murine antibodies shown in Table 1 above by CDR-grafting with a view to preserving their ligand-binding properties, the FR amino acids listed in Table 2 to 6 above would have to be retained.

A search through the tables of immunoglobulin sequences (Kabat et al., "Sequences of Proteins of Immunological Interest", 5th Ed. US Dept. of Health and Human Service, NIH Publication No.91-3242 (1991)), shows that human variable domain sequences are known that already have most of the FR amino acids that need to be preserved as shown in Table 7 below.

**Table 7: Human Antibodies that are Most Similar in Sequence to Murine Antibodies of Known Three-Dimensional Structure**

ANTIBODY DOMAIN			MOST SIMILAR HUMAN SEQUENCE
HYHEL-10	VH		58P2'CL (77/112)
	VH FRAMEWORK		1SP1'CL, MLI'CL (62/87)
	VH IMPT		58P2'CL, AB26'CL, C6B2'CL (28/38)
	VL		IARC/BL41'CL (73/107)
	VL FRAMEWORK		IARC/BL41'CL (59/80)
	VL IMPT		IARC/BL41'CL (30/37)
HYHEL-5	VH		ND'CL (74/116)
	VH FRAMEWORK		783c'CL, X17115'CL (63/87)
	VH IMPT		21/28'CL, S1P1'CL, 783c'CL, 8E10'CL, AND, KAS, NEI'CL, X17115'CL (25/37)
	VL		HF2-1/17'CL, KAS (65/105)
	VL FRAMEWORK		HF2-1/17'CL (57/80)
	VL IMPT		BI, DEN, HF2-1/17'CL, KUE, REI, WALKER'CL, WIL(-) (27/36)
R19.9	VH		21/28'CL (73/119)
	VH FRAMEWORK		21/28'CL, S1P1'CL, AND, LS2'CL, NEI'CL (60/87)
	VH IMPT		21/28'CL, 8E10'CL, LS2'CL (28/38)
	VL		WALKER'CL (78/107)
	VL FRAMEWORK		RZ (62/80)
	VL IMPT		REI, WALKER'CL (33/36)
4-4-20	VH		30P1'CL (77/116)
	VH FRAMEWORK		2P1'CL, 3D6'CL (65/87)
	VH IMPT		4B4'CL, H26'CL (36/41)
	VL		RPM1-6410'CL (91/112)
	VL FRAMEWORK		GM-607-'CL (68/80)
	VL IMPT		CUM, FR, NIM (33/36)
J539	VH		30P1'CL, Vh38C1.10'CL (81/118)
	VH FRAMEWORK		18/2'CL, 30P1'CL, M43 (71/87)
	VH IMPT		38P1'CL, S6P1'CL, M72, M74 (36/40)
	VL		PA (62/105)
	VL FRAMEWORK		LEN, WEA (53/80)
	VL IMPT		BI, DEN, KUE, REI, WALKER'CL, WIL(-) (26/35)
HCPC603	VH		H72 (81/120)
	VH FRAMEWORK		4G12'CL, Ab18'CL, M72 (70/87)
	VH IMPT		56P1'CL, M72, M74, RF-SJ2'CL (36/42)
	VL		FK-001'CL, LEN (91/113)
	VL FRAMEWORK		LEN (70/80)
	VL IMPT		LEN (39/42)
36-71	VH		21/28'CL (74/119)
	VH FRAMEWORK		21/28'CL, S1P1'CL, 783c'CL, AND'CL, NEI'CL, X17115'CL, (61/87)
	VH IMPT		21/28'CL, 8E10'CL (28/38)
	VL		AG (76/105)
	VL FRAMEWORK		RZ (63/80)
	VL IMPT		REI, RZ, WALKER'CL (34/37)

Table Human Antibodies that are Most Similar in Sequence to Murine Antibodies of Known Three-Dimensional Structure (Cont'd)

B1312	VH	S6P1'CL (83/119)
	VH FRAMEWORK	4B4'CL, 4G12'CL, M26'CL, H72, RF-SJ2'CL, VH38C1.10'CL (68/87)
	VH IMPT	S6P1'CL, H72, H74, RF-SJ2'CL (37/39)
	VL	REH1-6410'CL (86/1121)
D1.3	VH FRAMEWORK	GH-607-'CL (69/80)
	VL IMPT	CUM, NIM (36/39)
	VH	C682'CL (72/116)
	VH FRAMEWORK	C682'CL (62/87)
D1.3	VH IMPT	H60'CL (32/37)
	VL	BR (75/107)
	VL FRAMEWORK	HE2-1/17'CL (64/80)
	VL IMPT	3D6'CL, BI, DEN, EU, KUE, PA, REI, WALKER'CL, WIL(-) (3)

5 These human sequences are not necessarily those which are most similar to the murine antibodies, overall or in the framework regions only, but rather, those that possess the largest number of important amino acids in common, the latter sequences being included in Table 7 above.

10 The number of murine amino acids that still need to be retained in order to have all the important FR amino acids in the "humanized" or analogue versions of the murine antibodies, as shown in Table 7 above, ranges from 21 (for HyHEL-5:12 in  $V_H$  and 9 in  $V_L$ ) to 5 (for B1312:2 in  $V_H$  and 3 in  $V_L$ ). These are not very many amino acids, considering that the resulting "humanized" or analogue molecules will probably retain most or all their ligand-binding characteristics. It is possible that there exist other human sequences that are even more similar to these murine domains that are not included in the compilation of Kabat, et al. (1991), supra. When more sequences become available these may also be incorporated to improve the pool of basic data available.

b) Choosing the best human framework to use in the "humanization" of an antibody when its structure is not known

In the absence of a three-dimensional structure, the identification of the 5 FR amino acids that are crucial to maintain the combining site structure is not easily done. Nevertheless, some proposals may be made from the data shown in Tables 2 to 6 above that have been collected in Tables 8 and 9 below for the  $V_L$  and  $V_H$  domains.

10 **Table 8:** Framework Residues in  $V_L$  That Probably  
Need to Be Preserved in Order to Reproduce the  
Ligand Properties of the Original Antibody

		CDR1	CDR2	CDR3
J539	EL.L.Q....T.A....V.I.C	asas-----avsaib	WYQQ....SP.PMIV	
HcPC603	GLVHTO....L.V....V.H.C	ksqillnsqgnqknrie	WYQQ....SP.LLIV	
HvHEL-10	DLV.L.Q....L.V....V.L.C	asas-----signnli	WYQQ....SP.LLIV	
HvHEL-5	DLV.L.Q....M.A....V.H.C	asas-----avnymy	WYQQ....SP.RWIV	
R19.9	.I.QMTO....L.A....V.I.C	rasq-----disayin	WYQQ....T.KLLVY	
4-4-20	DVVMHTO....L.V....A.I.C	rasq-----avhsqgnyir	WYQQ....P.KVLIV	
36-71	DIOM.O....L.A....V.I.C	rasq-----dinflin	WYQQ....I.LLIV	
B1312	.V.LH.QF....L.V....A.ISC	rasq-----tillsdgqyle	WYQQ....SP.LLIV	
DL.3	DI.M.Q....L.A....V.I.C	rasq-----nihnyla	WYQQ....SP.LLIV	

**Table 9:** Framework Residues in  $V_H$  That Probably  
Need to Be Preserved in Order to Reproduce the  
Ligand Properties of the Original Antibody

		CDR1	CDR2	CDR3
J539	V.L.G....V....L.G.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
HcPC603	VL.L.G....V....L.G.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
HvHEL-10	V.L.L.C....V....L.G.C.V..,res:	asas-----avsaib	WYQQ....SP.PMIV	
HvHEL-5	...L.G....V....V.H.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
R19.9	V.L.L.C....V....V.H.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
4-4-20	...L.G....V....V.H.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
36-71	DLV.L.Q....V.H.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
B1312	...L.G....V....V.H.C.A..,res:	asas-----avsaib	WYQQ....SP.PMIV	
DL.3	...L.G....V....V.H.C.V..,res:	asas-----avsaib	WYQQ....SP.PMIV	

15 From Tables 8 and 9 above, it may be seen that many of the important FR amino acids flank the CDRs. Among these flanking positions are most of the

FR amino acids that are involved in the contact with the opposite domain as shown in Table 4 above, and many of those which are in contact with the CDRs as shown in Tables 2 and 3 above. Moreover, almost all of the FR amino acids that have been observed to participate in the binding to antigen (Amit, A.G., et al., *Science* 233:747-753 (1986); Sheriff, et al., *P.N.A.S. (USA)* 82:1104-1107 (1987); Padlan, E.A., et al., *P.N.A.S. (USA)* 86:5938-5942 (1989); Tulip, et al., *Cold Spring Harbor Symp. Quant. Biol.* 54:257-263 (1989); Bentley, et al., *Nature (London)* 348: 254-257 (1990)), are in these flanking regions. Thus, during "animalization" or "humanization" or formation of the analogue peptides, not just the CDRs are retained, but also some of the residues immediately adjacent to the CDRs. This provides a better chance of retaining more of the ligand-binding properties of the original antibody. The likelihood of retaining the antigen binding properties of the original antibody is even greater if the first few amino acids in the NH<sub>2</sub>-termini of both chains are also retained, since some of them are found to be in contact with CDRs as shown in Tables 2 and 3 above. Further, Tables 8 and 9 above also show many other framework positions that are deemed structurally important in all the cases examined here. The xenogeneic residues at those positions should probably be retained as well.

Alternatively, it may be possible to reduce immunogenicity, while preserving antigen-binding properties, by simply replacing those exposed residues in the framework regions which differ from those usually found in human antibodies (Padlan, E.A. (1991), *supra*). This would "humanize" the surface of the xenogeneic antibody while retaining the interior and contacting residues which influence its antigen-binding characteristics. The judicious replacement of exterior residues should have little, or no, effect on the interior of the domains, or on the interdomain contacts. For example, the solvent accessibility patterns of the F<sub>v</sub>s of J539, a murine IgA (x) and of KOL, a human IgG1 (y) have been found to be very similar (Padlan, E.A. (1991), *supra*).

At present, more than 35 different Fab structures have been elucidated by X-ray diffraction analysis, although atomic coordinates for only 11 are currently in the Protein Data Bank as shown in Table 1 above. Most of the available structures have been analyzed to only medium resolution, some having been refined to only a limited extent. Eventually, atomic coordinates for more and better-refined structures will become available, so that the "important" FRs will be more easily assessed. This will improve the theoretical predictive record of the present method for determining the best mode for the analogue peptides.

As already indicated above, the specificity of an antibody depends on the CDR structures and sometimes, on some neighboring residues as well. These structures, in turn, depend on contacts with framework amino acids and on the

interaction of the  $V_L$  and  $V_H$  domains. Thus, to ensure the retention of binding affinity, not only the CDR residues must be preserved, but also those FRs that contact either the CDR's or the pposite domain, as well as all buried residues, which give shape to the variable domains.

5 This design of the humanized versions of murine antibodies is reached in stages as follows.

- 1- Choice of a xenogeneic model of known structure.
- 2- Choice of the target species FR.
- 3- Identification of xenogeneic/target species differences.
- 10 4- Identification of important xenogeneic amino acids.

**(1) Choice of a xenogeneic model of known structure**

The  $V_H$  and  $V_L$  domains of an antibody of desired specificity are classified according to Kabat et al.(1991), *supra*. Then, an antibody of the same species may be chosen, whose structure has been determined, and whose 15 variable regions belong to the same classes and subclasses. Modeling the xenogeneic antibody in question to such structure ensures maximal chance for success. This, however, is not absolutely necessary since the relative positions of the important amino acids do not vary considerably even in variable regions of different classes. Thus, with less than a perfect match this method may still 20 be applied to design the analogues of this invention. Once the xenogeneic model is chosen, it may be applied to identify the locations of important residues in the xenogeneic antibody to be animalized (humanized). Tables 2, 3, 4, 5, 6, 8 and 9 indicate the positions of the important amino acids in several antibodies whose structures have been determined to a high resolution level.

25 **(2) Choice of the target species FR**

The target species framework should, ideally, be a consensus framework. That is, one that has a maximum number of amino acids in common with all 30 human frameworks of the same class. This is important, because, the goal of humanization is to avoid an immunological response against the engineered analogue peptide.

The target species framework that is chosen is that which shares the greatest number of important amino acids with the original xenogeneic antibody. Thus, in choosing the target species (human) FR, the similarity between the important amino acids is more important than the overall similarity.

35 In practice, the sequences of the xenogeneic variable chains are aligned with the consensus sequences from all variable region classes of the target species and the number of differences in the amino acids that must be retained from the xenogeneic species are scored. The human consensus sequence(s) that

score(s) the lowest number of differences is (are) then chosen. These are the best analogue peptide candidates. Others with low numbers that are higher than the above may also be suitable, and are placed in a reserve pool, and so forth.

If there are too many differences in the chosen framework (e.g., more than 16),

5 then the same alignment procedure using all tabulated human sequences may be repeated in order to find a specific human framework whose similarity with the xenogeneic sequence is maximized at the positions of the important amino acids.

Thus, most preferably, the target species FR should be a consensus sequence.

Next preferable would be a framework of a common target species (human)

10 antibody, and finally, the framework of any target species (human) antibody.

**(3) Identification of xenogeneic target species differences**

The xenogeneic sequences are then aligned with the target species sequences and the positions of all amino acids that differ in the murine and in the human frameworks are tabulated. Such a table contains the maximum 15 number of amino acids that can be changed toward the full "animalization" ("humanization") of the xenogeneic antibody (see, Tables 31 and 32 below). If all those changes were to be made, a so-called CDR-grafted antibody would be obtained. That is, only the original CDRs would be retained from the murine antibody. In some cases, possibly, such CDR-grafted antibody may maintain the 20 original binding affinity. In most instances, however, the affinity of a CDR-grafted antibody would be considerably less than that of the original xenogeneic antibody. In order to maximize the chances for conserving the original affinity, the identities of all important amino acids must be preserved.

**(4) Identification of important xenogeneic amino acids**

25 If the outlined approach to animalization (humanizing) an antibody is followed strictly, the amino acids that are correspondingly important in the model xenogeneic antibody chosen in step 1 are retained. In a more preferred approach, however, the amino acids that have been shown to occupy important positions in other antibodies of the same species or of the target species may also be 30 retained and are therefore taken out from the group of candidates to be mutated. This preferred approach may be particularly appropriate when there is a chance that the amino acids in question could make contacts with the CDRs or with the opposite chains. Once the important xenogeneic amino acids are identified, the DNA sequence may be mutagenized to change all other amino acids, which for 35 the most part occupy exposed positions.

The present method is exemplified for a murine antibody humanized with the intent of diminishing or avoiding a HAMA response upon its administration

to humans. Murine and human antibodies, whose three-dimensional structures have been deduced to a high degree of resolution, were utilized as guidance in the choice of the amino acids to be substituted in order to humanize the particular murine antibody utilized. The method, however, may be applied more generally to transform antibodies from one species into a less immunogenic form to be administered to a second species, provided that adequate three-dimensional models are available for antibodies from those species. Information on other murine antibodies from a Data Bank was used in the exemplary disclosure provided below to modify the BrE-3 and anti-KC-4 murine-human chimeric antibodies with human amino acids. Similarly, antibodies of other species besides murine may also be utilized, their CDRs and other amino acids preserved and those amino acids not considered "important" replaced with human amino acids. Similarly, the above approach may be applied to the preparation of "animalized" antibodies for any animal species. This may be attained by substituting amino acids of the antibody target species into an antibody of another species in accordance with this invention.

Various peptide structures, such as CDRs, and analogue antibodies, Fab, Fab', (Fab')<sub>2</sub>, and variable fragments having a desired specificity, may be constructed and optionally bridged via a linker. In addition, one or more of the peptides may be attached to one or more effector agent(s) or bridged via a linker. Multiple antibody, variable regions, Fab, Fab', (Fab')<sub>2</sub>, CDRs and the like, and combinations thereof, may also be constructed and bridged via linkers or attached to one or more effector agents such as are described below.

The cDNAs encoding the analogue variable regions of an antibody of a desired specificity may be cloned into a vector, optionally containing sequences encoding constant regions or fragments thereof, enzymes, neuropeptides, other peptide transmitters, toxins, hormones, operative conjugation regions, cytokines, lymphokines and the like, optionally under the same promoter. Although this is the cloning strategy utilized in the exemplary disclosure of this invention, other methods known in the art may also be utilized such as co-expression and the like. In the exemplary disclosure provided herein, the anti-BrE-3 and anti-KC-4 murine-human chimeric antibodies specifically binding to human mammary mucin and carcinoma cells was constructed by joining the DNAs of the anti-BrE-3 or anti-KC-4 murine variable domain to a human constant domain (an effector agent) cloned into a hybrid vector, and the product expressed by transfecting the vector into myeloma cells. The variable regions of the chimeric antibody were modified at the DNA level to obtain an analogue or "humanized" chimeric polypeptide. The modifications to the variable regions of the peptides may either be conducted by PCR amplification with primers that are custom tailored to

Table 10: Alternative Amino Acids for V<sub>l</sub> Chain (Cont.)

BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR2				
35	W	W		
36	F	F	YL(F)	IVHN
37	L	L	Q	WLVKRTHDE
38	Q	Q		
39	K	K	R	IFMCATSDN WIPMCA
40	S	P	P(S)	FLKARTGO
41G				IYFPMCT
42	Q	Q		WYVCD
43	S	S	P	WYFKMHDNEQ
44	P	P		WYKCRGHDQ
45	K	Q	EQR(K)	YPCGHD
46	L	L	RV	KCD
47	L	L	V	WILMTSN
48	I	I		FPLVMTS
49	Y	Y	S	PLVMA
FR3				
57	G	G		WVTSGDNEQ
58	V	V		IYFLVMAEQ
59	P	P	S	
60	D	D	N	
61	R	R	T	WFMCR
62	F	F		
63	S	S	T	IYPLKARSG
64	G	G	D	
65	S	S		
66	G	G		YCHDQ
67	S	S	A	VKMCRHQ
68	E	G	GD(E)	VMCARSQ
69	T	T		WPMCE
70	D	D		WIFPMCR
71	F	F		WKMTEQ
72	T	T		WLMCGHNEQ
73	L	L		
74	K	K	NLRE	WLMCHD
75	I	I	L	
76	S	S	IT	LVMC
77	R	R	S	WYFLKCHQ
78	V	V	ALI	WYFKCRHNE
79	E	E	KGQ	WVKM
80	A	A	P	WLKM
81	E	E		ILVKMAGDNE
82	D	D		
83	L	V	MV(L)	WCRH
84	G	G		LVARTSG
85	V	I	IM(V)	
86	Y	Y		WFKCQ
87	F	Y	YL(F)	
88	C	C		IMSHE

Table 11: Alternative Amino Acids for  $V_{\kappa}$  Chain (Cont'd.)

VL	BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
<b>FR2</b>					
36	W	W			
37	V	V			
38	R	R			
39	Q	Q			
40	S	A	VA(S)		
41	P	P	TS	PLVARTSHNEQ	
42	E	G	G(E)		
43	K	K			
44	G	G	SR		
45	L	L			
46	E	E	Q		
47	W	W			
48	V	V	STG		
49	A	A			
<b>FR3</b>					
66	R	R			
67	F	F			
68	T	T	IS		
69	I	I			
70	S	S	L		
71	R	R			
72	D	D	N	VKRTSGHDE	
73	D	N	N(D)		
74	S	S	A	FPLVTGDN	
75	K	K	EN		
76	S	N	NTRK(S)		
77	R	T	TNIVSM(R)		
78	V	L	LA(V)		
79	Y	Y	FH		
80	L	L			
<b>WYLKCARGHEQ</b>					
81	Q	Q	E		
82	M	M			
82a	I	M	SDN(I)		
82b	S	S	IR		
82c	L	L		PLVMAGE	
83	R	R	EKT		
84	A	A	SPVTI		
85	E	E	D	WIYFPLMCRG	
86	D	D			
87	T	T	M		
88	G	A	A(G)		
89	L	V	ITVM(L)		
90	Y	Y	HF		
91	Y	Y			
92	C	C			
93	T	A	ASHAV(T)		
94	G	R	R(G)	TSDQLAW	
<b>FR4</b>					
103	W	W	A		
104	G	G	YH		
105	Q	Q	THR		
106	G	G			
107	T	T	AQ		
108	L	L	STGM		

suitable for imaging tumors *in vivo*. A CDR attached to an effector peptide may be synthesized chemically or recombinantly encoded in a DNA segment. Such small molecules have great tumor penetration and extremely rapid clearing properties when compared to larger antibody fragments. In some cases, it is 5 more convenient to produce these small molecules by chemical synthesis, as is known in the art, rather than by fermentation. In many cases, these small peptides are completely non-immunogenic and an immune response, such as the HAMA response, is altogether avoided. Also preferred are 2 and 3 CDR units per chain operatively linked to one another by 1 to 10 or more amino acids and up 10 to the entire inter-CDR segment length as positioned in the variable regions.

Heavy and light chain analogue variable regions may be obtained individually or in  $V_H/V_L$  pairs, or attached to an effector peptide such as a constant region(s) or portions thereof, a drug, an enzyme, a toxin, a whole antibody, or any other molecule or radioisotope. The fragments of the analogue 15 variable regions may be synthesized chemically as is known in the art or from the DNA segments encoding the non-human variable regions. This may be attained by PCR amplification of the DNA with primers synthesized to contain the desired mutation(s) as is known in the art. Similarly, the fragments encoding analogue 20 variable regions may be synthesized chemically or obtained by established cloning methods of restriction digestion, ligation, mutagenesis, and the like, as is known in the art.

There are advantages to using the different molecular variants of the analogue peptide depending on the specific applications for which they are intended, some of which are listed below.

- 25        a)    Smaller molecules penetrate target tissues more efficiently and are cleared from the body much more rapidly than larger molecules.
- b)    Single chain molecules can be manipulated and synthesized more efficiently than multiple chain molecules.
- 30        c)    Many of these variants can be synthesized efficiently and inexpensively in bacteria, including the non-glycosylated analogues.
- d)    Bi-functional or multifunctional molecules may carry polypeptide effectors, such as enzymes, toxins, radioisotopes, drugs, and other molecules, to a target tissue.

The following list encompasses exemplary analogue peptides of the 35 invention engineered with molecules derived from antibodies or antibody fragments. These analogue peptides, among others, are suitable for the practice of this invention. A more extensive list of polypeptide constructs may be found in O'Kennedy, R., and Roben, P. (O'Kennedy, R., and Roben, P., "Antibody Engineering: an Overview", Essays Biochem. (England) 26:59-75 (1991)).

40        The analogue peptides and hybrid peptides of this invention encompass CDRs and/or analogue variable regions, monoclonal antibodies, antibody

fragments such as Fab, Fab', (Fab')<sub>2</sub>, and fragments thereof, CDRs, constant regions, single or multiple-domain and catalytic fragments, bi-functional or multifunctional combinations thereof, enzymes, peptide hormones, molecules such as drugs and linkers, transmitters, and toxins, among others. These are 5 suitable for imaging, therapy, and diagnostics.

#### Single-Chain Antigen-Binding Polypeptides

A method for constructing single chain antigen-binding polypeptides has been described by Bird et al. (Bird, R.E., et al., *Science* 242:243-246 (1988); 10 Bird, R.E., et al., *Science* 244:409 (1989)). Single Chain F<sub>v</sub> (scF<sub>v</sub> or sF<sub>v</sub>) are single chain analogue peptides containing both V<sub>L</sub> and V<sub>H</sub> with a linker such as 15 a peptide connecting the two chains (V<sub>L</sub>-linker-V<sub>H</sub>). The engineering may be done at the DNA level, in which case knowledge of the sequence is required. These analogue peptides have the conformational stability, folding, and ligand-binding affinity of single-chain variable region immunoglobulin fragments and may be 20 expressed in *E. coli*. (Pantoliano, M.V., et al., *Biochem. (US)* 30:10117-25 (1991)). The peptide linker binding the two chains may be of variable length, for example, about 2 to 50 amino acid residues, and more preferably about 12 to 25 residues, and may be expressed in *E. coli*. (Pantoliano, M.V., et al. (1991), *supra*). An analogue peptide such as an scF<sub>v</sub> may be expressed and prepared 25 from *E. coli* and used for tumor targeting. The clearance profiles for scF<sub>v</sub> in some situations fragments are advantageous relative to those of normal antibodies, Fab, Fab' or (Fab')<sub>2</sub> fragments. (Colcher, D., et al., *J. Natl. Cancer Inst.* 82:1191-7 (1990)). Another type of analogue peptide comprises a V<sub>H</sub>-linker-V<sub>L</sub> and may have about 230 to 260 amino acids. A synthetic gene using *E. coli* 30 codons may be used for expression in *E. coli*. A leader peptide of about 20 amino acids, such that of Trp LE may be used to direct protein secretion into the periplasmic space or medium. If this leader peptide is not naturally cleaved, the sF<sub>v</sub> analogue peptide may be obtained by acid cleavage of the unique asp-pro peptide bond placed between the leader peptide and the sF<sub>v</sub>-encoding region (Houston, J.S., et al., "Protein Engineering of Antibody Binding Sites: Recovery of Specific Activity in an Anti-Digoxin Single-Chain F<sub>v</sub> Analogue Produced in *E. coli*.", *PNAS (USA)* 85 (16):5879-83 (1988)). The construction, binding properties, metabolism and tumor targeting of the single-chain F<sub>v</sub> analogue peptides derived from monoclonal antibodies may be conducted as previously 35 described (Milenic, D.E., et al., *Cancer Res. (US)* 51 (23 pt1):6363-71 (1991); Yokota, et al., "Rapid Tumor Penetration of a single-chain F<sub>v</sub> and Comparison with Other Immunoglobulin Forms", *Cancer Res. (US)* 52(12):3402-8 (1992)). This type of analogue peptide provides extremely rapid tumor penetration and

even distribution throughout tumor mass compared to IgG or Ig fragments Fab and F(ab')<sub>2</sub>.

**Bifunctional scF<sub>v</sub>-Fxn or Fxn-scF<sub>v</sub>**

An example of this type of analogue peptide is a V<sub>L</sub>-linker-V<sub>H</sub> with an effector peptide such as a hormone, enzyme, transmitter, and the like. These hybrid analogue peptides may be prepared as described by McCarney et. al. (McCarney, J.E. et al., "Biosynthetic Antibody Binding Sites: Development of a Single-Chain F<sub>v</sub> Model Based on Antidinitrophenol IgA Myeloma MOPC 315", J. Protein Chem. (US) 10 (6):669-83 (1991)). A bi-functional hybrid analogue peptide containing an F<sub>c</sub>-binding fragment B of staph protein A amino terminal to a single-chain analogue F<sub>v</sub> region of the present specificity is also encompassed and may be prepared as previously described. (Tai, M.S., et al., Biochem. 29 (35):8024-30 (1990)). In this example of a hybrid analogue peptide of this invention is a Staph. A fragment B (anti F<sub>c</sub>) - scF<sub>v</sub> polypeptide. The order is backward of normal cases. This FB-scF<sub>v</sub> may be encoded in a single synthetic gene and expressed as peptide in E. coli. This analogue peptide is a good example of a useful multifunctional targetable single-chain polypeptide. A hybrid analogue peptide also comprising antibodies to a human carcinoma receptor and angiogenin is also part of this invention. Angiogenin is a human homologue of pancreatic RNase. This is an (Fab')<sub>2</sub>-like antibody-enzyme effector. Another hybrid analogue peptide comprising a V<sub>H</sub>-CH1 heavy chain-RNase may be expressed in a cell that secretes a chimeric light chain of the same antibody. A secreted antibody of similar structure was shown to cause the inhibition of growth and of protein synthesis of K562 cells that express the human transferrin receptor (Rybak, S.M., et al., "Humanization of Immunotoxins", PNAS 89:3165-3169 (1992)).

**Bi-specific Antibodies**

A monoclonal antibody or antibody fragment may be incorporated into a bi-specific analogue peptide as described, for example, by Greenman et al. (Greenman, J., et al., Mol. Immunol. (England) 28 (11):1243-54 (1991)). In this example, a bi-specific F(ab')<sub>2</sub> comprising two (Fab'-(thioether-link)-Fab') was constructed. Bi-specific antibodies may also be obtained when two whole antibodies are attached. Another way to obtain bi-specific antibodies is by mixing chains from different antibodies or fragments thereof. In this manner the "left" branch of the bi-specific antibody has one function while the "right" branch has another.

**Phage Display Libraries**

The analogue peptides in accordance with this invention may be screened with a filamentous phage system. This system may also be used for expressing any genes of antibodies or fragments thereof as well as for screening for 5 mutagenized antibody variants as described by Marks et al. (Marks, J.D., et al., "Molecular Evolution of Proteins on Filamentous Phage. Mimicking the Strategy of the Immune System", *J. Mol. Biol.* (England) 267 (23):1607-10 (1992)). A library of  $V_H$  and  $V_L$  genes or analogue thereof may be cloned and displayed on the surface of a phage. Antibody fragments binding specifically to several 10 antigens may be isolated as reported by Marks (Marks, J.D., "By-Passing Immunization. Human Antibodies from V-gene Libraries Displayed on Phage", *J. Mol. Biol.* (England) 222 (3):581-97 (1991)).

**Covalent Oligosaccharide Modifications**

The present analogue peptides alone or as hybrid peptides comprising 15 antibodies and fragments thereof may be, e.g., covalently modified utilizing oxidized oligosaccharide moieties. The hybrid analogue peptides may be modified at the oligosaccharide residue with either a peptide labeled with a radioisotope such as  $^{125}\text{I}$  or with a chelate such as a diethylenetriaminepentaacetic acid chelate with  $^{111}\text{In}$ . The use of 20 oligosaccharides provides a more efficient localization to a target than that obtained with antibodies radiolabeled either at the amino acid chain lysines or tyrosines (Rodwell, J.D. et al., "Site-Specific Covalent Modification of Monoclonal Antibodies: In Vitro and In Vivo Evaluations", *PNAS (USA)* 83:2632-6 (1986)).

25 Of the analogue peptides of this invention, preferred are those having the sequences ID Nos. 67 through 73, 75 through 81, 95, 96, and analogues thereof wherein about 1 to 42 amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the species for which the analogue is intended, such as human, or fragments thereof 30 comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 amino acids, or up to the complete N-terminal region, or combinations thereof. Examples of possible substitute amino acids in all positions, including the most important positions are shown in Tables 10 and 11 above, and are 35 indicated in the columns titled BrE-3 and KC-4, "Most Preferred Analogue and Preferred Analogue" amino acids substituents. Others are also suitable as may be deduced by the method described herein. These amino acid sequences may be bound by a peptide or non-peptide linker such as is known in the art.

Examples of peptide linkers are polylysines, leucine zippers, EGKSSGSGSEJKVD, and (GGGGS)x3, and non-peptide polymers, among others. Effector agents such as peptides and non-peptides may also be attached to the analogue peptides of the invention. These include non-peptide polymers, monomers, atoms, etc.,

5 which are discussed below.

Another preferred embodiment comprises a bi-functional analogue peptide having a pair of light and heavy chains of the same specificity attached to one another by a linker, such as those provided above. In another preferred embodiment, a bi-functional analogue peptide comprises one set of light and 10 heavy chains comprising at least one xenogeneic CDR or variable region, e.g., the amino acid sequence ID No. 11 or 13 of Tables 15 and 16, sequence ID No. 95 or 96 from Tables 55 and 56 below, or the respective CDRs alone or separated by 1 to 10 amino acids and up to the entire amino acid segments of their flanking sequences, and optionally with 1 to 10 amino acids of each N- 15 terminal region, sequence ID No. 68, 70 and/or 72, alone or with flanking sequences of varying length, or 76, 78 and/or 80, alone or with flanking sequences of varying length, with the modifications shown above, wherein about 1 to 42 amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the target species, or 20 fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more amino acids, and one set of light and heavy chains comprising at least one xenogeneic CDR or analogue variable region, e.g., amino acid sequence ID No. 11, 13, 95, or 96 having a different set of substitute amino 25 acids, wherein about 1 to 46 or more amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the target species, or fragments comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more amino acids, or 30 fragments or combination thereof. Multi-functional hybrid analogue peptides may comprise several identical units or combinations of the above bi-functional analogue peptides of the same or different specificities or xenogeneic species. Preferred analogue peptides are those comprising murine CDRs and other regions substituted with human amino acids.

35 In another aspect, this invention provides a hybrid analogue polymer that comprises at least one anti-tumor analogue peptide and at least one effector agent operatively linked to the peptide, combinations thereof and mixtures thereof. The effector agent utilized in this invention comprises peptide polymers other than the constant region of an antibody of the same species as the CDRs,

HuKC4V3, ATCC No. HB 11456 deposited on September 23, 1993, all of which deposited under the Budapest Treaty as an example of a best mode of this invention).

The hybrid analogue polymer may comprise two heavy and two light chains, each light and heavy chain comprising at least one CDR or analogue variable region polypeptide or fragments thereof of one species and the constant region and the substitute amino acids of an antibody of a different species such as human, at least one other CDR, analogue variable region, chimeric Fab, Fab' or (Fab')<sub>2</sub>, fragments thereof, combinations thereof, and mixtures thereof. Still more preferred is a hybrid analogue peptide comprising at least two "humanized" murine-human or chimeric antibody fragments thereof, Fab, Fab' or (Fab')<sub>2</sub> fragments thereof operatively linked to one another. The peptide fragments may be covalently attached to one another as is known in the art (Marchis-Mouren G., et al., "HT 29, a Model Cell Line: Stimulation by the Vasoactive Intestinal Peptide (VIP); VIP Receptor Structure and Metabolism", Bioch. 70 (5):663-71 (1988)), or they may be synthesized by methods known in the art (Allen, G., et al., "Production of Epidermal Growth Factor in Escherichia Coli from a Synthetic Gene", J. Cell Sci. Suppl. 3:29-38 (1985)).

The hybrid analogue polymer of the invention described above may have two heavy and two light analogue chains operatively linked to one another, where each pair of heavy and light chains, has specificity for a different epitope. One example of this analogue peptide is a pair of "humanized" variable region heavy and light chains of a BrE-3 analogue peptide and a pair of "humanized" variable region light and heavy chains of a KC-4 analogue peptide that are covalently attached to one another by a peptide or non-peptide polymer or a disulfide bridge, or non-covalently by means of a leucine zipper or two helical structures, and the like. Non-peptide polymers may be covalently attached to peptides by methods known in the art (Duronio, V., et al., "Two Polypeptides Identified by Interleukin 3 Cross-Linking Represent Distinct Components of the Interleukin 3 Receptor", Exp. Hematol. 20 (4):505-11 (1992)). In another embodiment, the invention provides a hybrid analogue peptide comprising at least one CDR or analogue variable region of the heavy chain of an antibody of a first species or fragments thereof, operatively linked to a first effector agent, and at least one CDR or analogue variable region of the light chain of an antibody of a second species or fragments thereof operatively linked to a second effector agent and combinations thereof, wherein each pair of light and heavy chains has a predetermined specificity, combinations thereof, and mixtures thereof. In another preferred embodiment of the hybrid analogue peptide, at least one CDR or analogue variable region of the heavy chain of a murine antibody or fragments

thereof and at least one CDR or variable region of the light chain of a murine antibody or fragments thereof are linked to one another by a non-peptide polymer such as an isoprene polymer or monomer. In still another preferred embodiment, the hybrid analogue peptide of the invention is one wherein at least one pair of 5 light and heavy chains comprising at least one murine CDR or analogue variable region or fragment thereof is linked to at least one other pair of light and heavy chains comprising at least one murine CDR or analogue variable region or fragment thereof. In another embodiment the two or more F<sub>v</sub> regions are covalently attached to one another by a peptide or non-peptide polymer or a 10 disulfide bridge, or non-covalently by means of a leucine zipper or two helical structures, and the like. In a most preferred embodiment, the analogue peptides and hybrid polypeptides of the invention have affinity and specificity for an epitope located in the most hydrophilic region of a 20 amino acid tandem repeat that makes up a large part of the polypeptide core of mammary mucin, to 15 hexamer fragments of the sequence APDTRPAPG or trimer TRP fragments shown to afford the strong binding of all five different monoclonal antibodies raised against the human mammary fat globule (Mc1, Mc5, BrE-1, BrE-2 and BrE-3). The monoclonal antibodies were shown to bind to different but overlapping polypeptide epitopes but to have different tissue and tumor specificities, to 20 quantitatively differ in their binding to tumor cells such as breast carcinoma cell lines when observed by flow cytometry and have different competition patterns for binding to the native antigen on breast carcinoma cells. Also preferred amongst antibodies utilized for the preparation of the present analogue peptide and hybrid polypeptide are those that exhibit strong binding to the hexamer 25 peptides described above or to fragments comprising a TRP trimer to tandem repeats thereof. In one most preferred embodiment, the analogue peptide comprises the humanized mutein antibody expressed by the hybridoma cell line having the ATCC Accession No. HB 11200 (BrE-3 HZ). This cell was deposited with the ATCC under the Budapest Treaty on November 13, 1992. In another 30 most preferred embodiment, the analogue peptide comprises the humanized mutein antibody expressed by the hybridoma cell line having the ATCC Accession No. HB 11455 (HuKC-4V2), deposited under the Budapest Treaty on September 23, 1993. These were deposited as the best mode of the invention known to the inventors.

35 The anti-tumor analogue peptide and/or hybrid analogue polymer of the invention are also provided as an anti-tumor composition along with a carrier or diluent, preferably a pharmaceutically-acceptable carrier or diluent. The anti-tumor analogue peptide and the hybrid polymer provided herein may be present in the composition in an amount of about 0.001 to 99.99 wt%, more preferably

about 0.01 to 20 wt%, and still more preferably about 1 to 5 wt%. However, other amounts are also suitable. Carriers generally, and pharmaceutically-acceptable carriers in particular are known in the art and need not be further described herein. The carrier may be provided in a separate sterile container or 5 in admixture with the polypeptide. Typically, saline, aqueous alcoholic solutions, albumin-saline solutions, and propylene glycol solutions are suitable. However, others may also be utilized. When utilized for therapeutic purposes the proteic material must be of a purity suitable for human administration, and the composition may contain other ingredients as is known in the art. Examples of 10 these are other anti-neoplastic drugs such as adriamycin and mitomycin, cytoxan, PALA and/or methotrexate, among others. However, other therapeutic drugs, carriers or diluents, immunological adjuvants and the like may be also be added. When the composition described above is utilized for in vivo imaging, it may comprise about 0.001 to 99.9 wt% analogue peptide, and more preferably 15 about 0.01 to 25 wt% analogue peptide. Typically, when the composition is utilized for therapeutic purposes it may contain about 0.001 to 99.9 wt% analogue peptide, and more preferably about 0.01 to 30 wt% analogue peptide. When utilized for the ex vivo purging of neoplastic cells from bodily fluids such 20 as spinal fluid, the composition may comprise about 0.0001 to 50wt%, and preferably about 0.01 to 20wt% analogue peptide. When applied to the in vitro diagnosis of tumors such as carcinomas the composition of the invention may comprise about 0.001 to 35 wt% analogue peptide, and more preferably about 0.01 to 10 wt% analogue peptide. Other amounts, however, are also suitable.

Such products find one utility in the treatment of tumors such as 25 carcinomas, such as breast, lung, ovary, endometrial, pancreas, prostate and colon cancers, among others. The "humanized", "half humanized" and "partially humanized" analogue peptides may be used for the in vivo treatment or diagnosis of humans. The "animalized", "half animalized" and "partially animalized" analogue peptides of the invention may be utilized for the treatment of non- 30 human species such as were described above insofar as the amino acids of the species are substituted for those of the xenogeneic amino acids present at a specific location, and any constant region present in the analogue. The present analogue peptides are particularly suitable for repeated administration to a subject and for long term therapy, such as is the case of metastases and/or the 35 reoccurrence of tumors. Of all analogues described and encompassed herein, the ones most suitable for in vivo applications are those that exhibit low or no binding to serum antigens and to normal cells. Suitable for in vitro or ex vivo uses are those that exhibit good binding to tumor cell antigens such as the carcinoma cell antigen and weak or no binding to normal cells. Even though a

patient may have in circulation an interfering amount of a molecule that can bind the analogue peptide, the peptide may still be administered after removal of such serum molecule either by ex-vivo procedures or by administration of flush doses of the analogue peptide or fragments thereof.

5 A kit for the diagnosis of tumors such as carcinomas may comprise a composition comprising the anti-tumor analogue peptide of the invention, a solid support, anti-tumor antibody (positive control), immunoglobulins of a different species selectively binding the constant regions of the antibody, protein G or protein A, and instructions for its use. This diagnostic kit may be utilized by  
10 covalently attaching the antigen or the analogue peptide of the invention or a fusion protein thereof to the solid support by means of a linker as is known in the art. In a particularly preferred embodiment, the support is coated with a polypeptide such as methylated albumin as described in US Patent No. 4,572,901, the relevant text of which is incorporated herein by reference. When  
15 a biological sample is added to a well, the analogue peptide of the invention will bind any tumor antigen, such as a carcinoma antigen, present in the biological sample. If a competitive assay is utilized, to the solid supported antigen or hybrid peptide thereof are added a known amount of the analogue peptide and the sample. Thereafter,  $\gamma$ -globulin, protein G or protein A in labeled form may  
20 be added for detection. Anti-tumor antibodies of a first species may be obtained by challenging a subject of another species with tumor cells such as carcinoma cells, the human milk fat globule mucin and the like, as is known in the art (Peterson, J.A., et al., *Hybridoma* 9:221 (1990); US Patent No. 4,708,930). Monoclonal antibodies may be prepared as described by Kohler and Milstein  
25 (Kohler, G. and Milstein, C., "Continuous Culture of Fused Cell Secreting Antibody of Predefined Specificity", *Nature* 256:495-497 (1975)). Suitable for use in this invention are antibodies such as IgG, IgM, IgE, IgA, and IgD. Protein A, protein G and  $\gamma$ -globulin may be obtained commercially.

A diagnostic kit for detecting tumors such as carcinomas, and more  
30 particularly human carcinomas is provided herein that comprises an anti-tumor composition comprising a hybrid analogue peptide and an effector agent comprising an enzyme, a radioisotope, a fluorescent label and/or a peptide comprising the constant region of an antibody of the species for which use it is intended, or fragments thereof capable of binding anti-constant region  
35 immunoglobulins, protein G or A, anti-tumor antibody, anti-constant region immunoglobulins, protein G or protein A, a solid support having operatively linked thereto an antigen which specifically binds to the anti-tumor hybrid analogue peptide of the invention and the antibody, and instructions for its use. When the effector agent comprises a peptide, such as the constant region of an antibody

of the target species, the solid support may have operatively linked thereto an antibody which specifically binds to a portion of a fusion protein other than the antigen of the invention. This permits the binding of the anti-tumor analogue peptide to the antigen molecule now attached to the solid support. Any complex 5 formed between the hybrid analogue peptide of the invention and the supported tumor antigen will, thus, remain attached to the solid substrate. A competitive assay may then be conducted by addition to the solid supported antigen of a known amount of the hybrid antigen peptide and the sample. The amount of antigen present in the sample may be obtained from a dilution curve by addition 10 of anti-constant region immunoglobulins, protein G, protein A or other antibody binding molecules, e.g., labeled, to bind the hybrid analogue peptide that is now attached to the support. This kit may be used in a competitive assay where the supported antigen molecule competes with antigen in the sample for a known amount of the analogue peptide of the invention. The assay was described by 15 Ceriani, R.L., et al. (Ceriani, R.L., et al., Anal. Biochem. 201:178-184 (1992)), the relevant text thereof being incorporated herein by reference.

A tumor such as a carcinoma may be imaged *in vivo* and/or diagnosed by administering to a subject suspected of carrying a carcinoma the anti-tumor analogue peptide of the invention in radiolabeled form, in an amount effective to 20 reach the tumor cells and bind thereto, and detecting any localized binding of the labeled analogue peptide to the tumor. Typically, the analogue peptide of the invention may be administered in an amount of about 0.001 to 5000 mg/kg weight per treatment, more preferably about 0.01 to 5000  $\mu$ g/kg weight per treatment, and more preferably about 0.1 to 500  $\mu$ g/kg weight per treatment. 25 However, other amounts may also be utilized. Radiolabels that may be utilized are  $^{111}\text{In}$ ,  $^{125}\text{I}$ ,  $^{99\text{m}}\text{Tc}$ , and  $^{131}\text{I}$ , among others. These radioisotopes may be detected with a PET scanner, NMR imaging, and radioactivity counting apparatus that are in wide use by the medical community.

The presence of a tumor such as a carcinoma may also be diagnosed in 30 vitro by contacting a biological sample with the anti-tumor analogue peptide or hybrid polypeptide of the invention to form an anti-tumor analogue peptide-antigen complex with any tumor antigen present in the sample, and detecting any complex formed. The biological sample is typically obtained from a subject such as a human suspected of being afflicted with the tumor. Suitable biological 35 samples are serum, blood, sputum, feces, lymph fluid, spinal fluid, lung secretions, and urine, among others. Clearly, any source of fluid, tissue and the like may be prepared for use in this method as is known in the art.

Neither the hybrid half chimeric/half humanized BrE-3 analogue peptide nor the chimeric BrE-3 polypeptide of the invention or the murine BrE-3 antibody

show substantially strong binding to normal tissue. The hybrid BrE-3 analogue peptide shows a binding pattern similar to the chimeric BrE-3 polypeptide and the murine BrE-3 antibody. The murine BrE-3 antibody was shown to bind with specificity to various tumors such as carcinomas of the breast, lung, ovary, 5 bladder, and the endometrium, mesothelioma, colon, kidney, liver, pancreas, salivary glands, sarcomas and thyroid, among others. Weak binding was only shown to normal breast tissue, lung tissue, distal convoluted tubes of the kidney, acini of the pancreas and stomach mucosa (Peterson, J.A., et al. (1990), *supra*). The KC-4 hybrid murine peptide has tissue specificity similar to that of the 10 murine KC-4 antibody. The KC-4 monoclonal antibody was shown to bind specifically and strongly to solid tumor tissue in the lung, colon, kidney, breast, stomach, prostate, pancreas, lymph node duct and lymphoma, and non-specifically and weakly to normal breast, kidney, and stomach tissue. KC-4 also showed some weak binding to normal tissue including spinal cord, uterus, 15 thyroid, tongue, prostate, spleen, adrenal, lung, gall bladder, heart, lymph nodes, colon, liver, brain, testes, thymus, and placenta (U.S. Patent No. 4,708,930). In one preferred embodiment of the in vitro diagnostic method, the anti-carcinoma analogue peptide added to the biological sample comprises a labeled hybrid analogue peptide. Suitable labeling materials were described above. This 20 method may be practiced, with the solid support containing kit described above, as a competitive assay as disclosed by Ceriani, R.L., et al. (Ceriani, R.L., et al. (1992), *supra*).

The present analogue peptides are also applicable to the purging of neoplastic cells, such as carcinoma cells, from biological samples, be it fluid or 25 tissue samples. The purging of neoplastic cells from a fluid sample is part of the invention and may be practiced by contacting a biological fluid suspected of comprising neoplastic cells with the analogue peptide of the invention, which is capable of selectively binding to an antigen of the neoplastic cells and allowing the peptide to bind to the antigen, and separating the analogue peptide-cell 30 complex from the remainder of the fluid.

This method may be utilized for purging unwanted cells *ex vivo* by extracting a biological sample from a patient, eliminating the neoplastic cells therefrom by separation of the analogue peptide-cell complexes or by further addition of an effector such as complement or a toxin or a radioactive label that 35 can act upon the cell and then replenishing the purged sample to the patient. This is typically suitable for use with spinal taps where spinal fluid is rid of neoplastic cells such as carcinoma cells prior to reinjection. Other fluids may also be treated in this manner.

The present analogue peptides may also be applied to the histochemical

assessment of the presence of neoplastic cells such as carcinoma cells in a tissue obtained from a subject suspected of being afflicted by a carcinoma by methods that are standard in the art, like the preparation of tissue slices and fixation on a solid substrate to permit the application of the peptide and then the 5 assessment of any binding to neoplastic cells in the sample as indicated by the formation of complexes between the analogue peptide and antigens on or in the cells.

The growth or the size of a primary or metastasized tumor or neoplasia such as a carcinoma may be inhibited or reduced by administering to a subject 10 in a need of the treatment an effective amount of the anti-tumor hybrid analogue peptide of the invention. Typically, the hybrid analogue peptide may be administered in an amount of about 0.001 to 2000  $\mu\text{g}/\text{kg}$  body weight per dose, and more preferably about 0.01 to 500  $\text{mg}/\text{kg}$  body weight per dose. Repeated doses may be administered as prescribed by the treating physician. However, 15 other amounts are also suitable. Generally, the administration of the hybrid analogue peptide is conducted by infusion so that the amount of radiolabel, toxin or other effector agent present that may produce a detrimental effect may be kept under control by varying the rate of administration. Typically, the infusion of one dose may last a few hours. However, also contemplated herein is the 20 constant infusion of a dose for therapeutic purposes that will permit the maintenance of a constant level of the hybrid polypeptide in serum. The infusion of the hybrid analogue peptide of the invention may be conducted as follows. Intravenous (I.V.) tubing may be pretreated, e.g., with 0.9 % NaCl and 5% 25 human serum albumin and placed for intravenous administration. The prescribed dose of the analogue peptide may be infused as follows. Unlabeled analogue peptide may be infused initially. 30 minutes after completion of the unlabeled antibody infusion,  $^{111}\text{In}$ -labeled and  $^{90}\text{Y}$  labeled antibody may be co-infused. The I.V. infusion may comprise a total volume of 250 ml of 0.9 % NaCl and 5 % 30 human serum albumin and be infused over a period of about 2 hours depending on any rate-dependent side effects observed. Vital signs should be taken every, e.g., 15 minutes during the infusion and every one hour post infusion until stable. A thorough cardiopulmonary physical examination may be done prior to, and at the conclusion, of the infusion. Medications including acetaminophen, diphenhydramine, epinephrine, and corticosteroids may be kept at hand for 35 treatment of allergic reactions should they occur. The administration of the hybrid analogue peptide of the invention may be repeated as seen desirable by a practitioner. Typically, once a first dose has been administered and imaging indicates that there could be a reduction in the size of the tumor, whether primary or metastasized, repeated treatments may be administered every about

1 to 100, and more preferably about 2 to 60 days. These repeated treatments may be continued for a period of up to about 2 years, and in some circumstances even for longer periods of time or until complete disappearance of the tumor(s).  
5 The administration of the hybrid analogue peptides of this invention is typically more useful for therapeutic purposes when a primary tumor has, for example, been excised. Thus, it is primarily, for mopping up after surgical intervention or in cases of cancerous metastases that the present method is of most use.

A pure, isolated analogue polydeoxyribonucleotide that comprises an analogue oligodeoxyribonucleotide encoding the analogue peptide or hybrid 10 peptide of this invention, including all redundant sequences may be applied to the preparation of the peptide. In one preferred embodiment, the analogue polydeoxyribonucleotide of the invention comprises a DNA sequence selected from the group consisting of DNA sequence ID No. 64 of Table 45, or DNA sequence ID No. 65 of Table 46, or DNA segments encoding the CDR fragments 15 sequence ID No. 68, 70 and/or 72, or 76, 78 and/or 80 flanked by 1 to 10 or more amino acids, and the N-terminal fragment of 1 to 10 or more amino acids of human origin. The above DNA sequences may be cloned for expression under the same promoter. Similarly preferred are also the DNAs having the sequence ID No: 93 and/or 94 of Tables 52 and 53 and the segments encoding their CDR 20 fragments alone or separated by DNA segments encoding 1 to 10 or more flanking amino acids and/or terminated by DNA segments encoding 1 to 10 or more amino acids of the N-terminus.

Also provided herein is a hybrid vector that comprises a vector having the analogue polydeoxyribonucleotide of this invention operatively linked thereto.  
25 Typically, vectors capable of replication both in eukaryotic and prokaryotic cells are suitable. When the preparation of a glycosylated analogue polypeptide is desired the vector should be suitable for transfection of eukaryotic host cells. In one preferred embodiment, the hybrid vector comprises the analogue polydeoxyribonucleotide and a polydeoxyribonucleotide comprising an 30 oligodeoxyribonucleotide encoding an effector peptide, the effector peptide-encoding polydeoxyribonucleotide being operatively linked to the vector. As already indicated, the various DNA sequences may be cloned for expression under the same promoter. In addition, the polydeoxyribonucleotide encoding the effector polypeptide may also be cloned for expression under the same promoter.  
35 This invention also encompasses a host cell that has been transfected with the hybrid vector described above. Suitable hosts are prokaryotic and eukaryotic hosts such as bacteria, yeast, and mammalian cells such as insect cells and non-producing hybridoma cells, among others. Suitable vectors and/or plasmids for the transfection of each one of these types of hosts are known in

the art and need not be further described herein. Also known in the art are methods for cloning DNA sequences into each one of these types of vectors and for transfecting the different types of host cells. Particularly preferred is the cell line having the ATCC Accession No. HB 11200 (BrE-3 HZ).

5 Polyribonucleotides may be obtained by transcription of the polydeoxyribonucleotides described above as is known in the art. Provided herein are analogue polyribonucleotides comprising analogue oligoribonucleotides encoding at least one CDR or an analogue variable region or fragments thereof, combinations thereof, and combinations thereof with an effector peptide may be  
10 prepared by cloning the desired DNA segments and then transcribing the thus obtained hybrid polydeoxyribonucleotide into the corresponding RNA sequences.

The analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a neoplastic cell, or is released by the cell, may be produced by a method that comprises cloning the analogue  
15 polydeoxyribonucleotide of the invention into a vector to form a hybrid vector, transfecting a host cell with the hybrid vector and allowing the expression of the anti-tumor analogue peptide, and isolating the anti-tumor polypeptide or mixtures thereof. The DNA segment encoding the analogue polypeptide may be obtained by chemical synthesis or by site-specific modification of the sequence encoding  
20 the variable region of the xenogeneic species by PCR amplification with specifically designed primers as is known in the art. The fragment DNAs may also be prepared by PCR with primers that introduce a stop codon at a desired position as is known in the art. Preferably, the cloning and transfection steps are conducted by cloning polydeoxyribonucleotides encoding the analogue peptides  
25 selected from the group comprising at least one CDR or analogue variable region of the heavy or light chains of the xenogeneic species, antibodies thereof, or fragments thereof. The method may further comprise allowing the expressed analogue peptides to interact with one another to form double chain analogue peptides, one or both analogue peptide chain comprising at least one xenogeneic  
30 CDR or variable region of the light or heavy chain of the antibody or fragment thereof modified as described above. Still part of this invention is a method of producing a hybrid analogue peptide comprising an effector peptide and an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a tumor cell such as a carcinoma cell or that is released by the cell,  
35 the method comprising transfecting a host cell with the hybrid vector of this invention carrying a DNA sequence encoding the hybrid analogue peptide, allowing the expression of the hybrid analogue peptide, and isolating the hybrid analogue peptide or mixtures thereof. The techniques for obtaining mRNA, conducting reverse transcription and PCR amplification of DNA, chemical

synthesis of primers, cloning DNA sequences into a vector, transfecting a host cell, and purifying polypeptides from a culture medium are known in the art and need not be further described herein.

Another aspect of this invention relates to an anti-idiotype peptide that

5 comprises polyclonal antibodies raised against anti-tumor antibodies, the analogue peptide of the invention, monoclonal antibodies thereof, fragments thereof selected from the group consisting of CDRs, Fab, Fab', (Fab')<sub>2</sub>, and variable region fragments and fragments thereof, analogues thereof selected from the group consisting of Fab, Fab', (Fab')<sub>2</sub>, and variable regions thereof,

10 wherein about 1 to at least 46 amino acids in the FRs are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in human antibodies, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more

15 amino acids. The technique for obtaining anti-idiotype polypeptides is known in the art and need not be further described herein (Nisonoff, A. and Lamoyi, "Implication of the Presence of an Internal Image of an Antigen in Anti-Idiotype Antibodies: Possible Applications to Vaccine Production", Clin. Immunol. Immunopathol. 21:397-406 (1981)). Moreover, the technique for producing

20 hybridomas secreting monoclonal antibodies of a certain specificity is also known in the art (Kohler, G. and Milstein, C. (1975), *supra*). Techniques for obtaining different antibody fragments were described above or are known in the art and need not be further described herein (Wilbanks, T., et al., "Localization of Mammary Tumors In Vivo with <sup>131</sup>I-Labeled Fab Fragments of Antibodies Against

25 Murine Mammary Epithelial (MME) Antigens", Cancer 48:1768-1775 (1981)). The techniques for modifying peptides to obtain the analogue peptides of this invention have been described above or are known in the art.

The hybrid anti-idiotype polymer may further comprise an effector agent operatively linked to the anti-idiotype polypeptide. Effector agents suitable for

30 use herein were described above for the anti-tumor analogue polypeptide of the invention are also suitable for use with the anti-idiotype polypeptide. Preferred hybrid polymers are polyclonal antibodies raised against the anti-tumor monoclonal antibodies or the analogue peptide of the invention, and a monoclonal antibody obtained by fusion of a B-cell producing an antibody having

35 specificity for the analogue peptide of the invention and an immortalized cell line. Also preferred are fragments of the monoclonal antibody such as Fab, Fab', (Fab')<sub>2</sub> and variable region fragments, analogues and fragments thereof as described above, and CDRs. Also, as described above for the anti-tumor polypeptide analogue, preferred are combinations of the above fragments and

analogues and combinations of the fragments with whole antibodies and analogues thereof. In another preferred embodiment, the anti-idiotype polymer comprises an analogue variable region of a monoclonal antibody linked to a peptide comprising the hexamers or trimers described above or tandem repeats thereof.

5 DNA and RNA segments encoding the anti-idiotype polymer and hybrid polymer, a hybrid vector having the DNA operatively linked thereto and a host cell transfected with the hybrid vector are also contemplated herein.

Also provided herein is an anti-tumor vaccine that comprises the anti-10 idiotype polypeptide of the invention, and a pharmaceutically-acceptable carrier. Typically, the anti-idiotype polypeptide is present in the composition in an amount of about 0.001 to 99.99 wt%, and more preferably about 0.01 to 50 wt% of the composition. However, other amounts are also suitable. Pharmaceutically-acceptable carriers are known in the art and need not be further 15 described herein. The vaccine provided herein may further comprise other ingredients such as adjuvants, and the like. Examples of adjuvants are SAF-1 and Freund's, among others. Suitably, other ingredients typically used for the preparation of vaccines may also be utilized herein. In one embodiment, the vaccine of the invention may be provided in unit form as a powder or in a 20 diluent. In another embodiment, it may be provided in powder form in a sterile container comprising a plurality of doses for preparation prior to utilization. Diluents that are suitable for the preparation of a formulation that may be administered to a patient by injection are known in the art. Examples were provided above.

25 An anti-tumor vaccination kit is also provided by this invention that comprises, the vaccine described above and a diluent in separate sterile containers, and instructions for its use.

An effective amount of the anti-idiotype polypeptide or hybrid polypeptide described above may be administered to vaccinate a human. Typical 30 amounts administered to vaccinate a human are about 0.001 to 5000 mg/kg body weight/dose, more preferably about 0.1 to 5000  $\mu$ g/kg body weight/dose. The anti-idiotype vaccine of the invention may be administered repeatedly in order to boost the active immunization produced by the first dose. An anti-idiotype antibody very likely resembles the epitope on the neoplastic cell to 35 which the anti-tumor antibody binds. Thus, it may be utilized for the production of an immunological response by a subject such as a human or other mammals against its own neoplastic cells.

When an anti-idiotype polypeptide of, e.g., non-human origin is administered to a, e.g., human, it may produce a somewhat detrimental

response. Accordingly, in theory, the smaller the non-human amino acid sequence the anti-idiotype polypeptide contains, the lesser the immunogenic response to its xenogeneic sequences it will elicit in a human. Accordingly, preferred anti-idiotype polypeptides are those containing at least one CDR or 5 variable region of a non-human antibody binding specifically to the anti-tumor polypeptide described herein, optionally as a hybrid polypeptide. Also preferred are human anti-idiotype antibodies, CDR and variable fragments thereof, and fragments thereof that are operatively linked to an effector agent comprising a human polypeptide that may include the constant region of a human antibody 10 and fragments thereof, non-peptide polymers, monomers and atoms that may be radiolabeled as described above. Other types of constructs are also possible, several of which were described above.

Peptides comprising the sequence APDTRPAPG or fragments thereof comprising hexamers with the trimer TRP or TRP by itself or tandem 15 repeats thereof may also be utilized for the preparation of the fusion protein, particularly as part of the antigenic peptide. The peptide comprising the hexapeptide or tripeptide sequences may be utilized as a tandem repeat comprising up to about 10,000 repeats of the basic unit, an in some instances up to about 500,000 repeats. In another embodiment, peptides comprising one 20 or more hexapeptides or tripeptides may be operatively linked to other polypeptide sequences of related or unrelated function, which sequences provide bulk that aids the clearance through the liver and/or kidneys of the immunological complex formed between the circulating unbound or residual antibody or polypeptides utilized for the therapy of neoplasias such as carcinomas and the 25 hexapeptide. The peptides comprising the hexapeptide or tripeptide may also be provided as a hybrid analogue peptide with other analogue peptides described above. In the absence of such treatment, the therapeutic antibody, which may carry a radioisotope, a toxin or other therapeutic molecules, may remain in the circulation for several days and in some instances weeks. This, in the case of a 30 radioactively labeled antibody or analogue of the invention may produce extensive damage, which is highly detrimental to the health of the patient, and in some instances lethal.

Thus, the serum concentration of a circulating antibody or polypeptide that binds to an antigen on the surface or in the cytoplasm of tumor cells such 35 as carcinoma cells or released by the cells may be lowered by administering to a subject the anti-idiotype polypeptide described above, in an amount effective to bind the circulating polypeptide, to thereby accelerate its clearance. Typically, the anti-idiotype polypeptide or hybrid polymer are administered in an amount of about 0.001 to 5000.00 mg/kg body weight/dose, more preferably about 0.01

5000 mg/kg body weight/dose. However, other amounts may also be utilized. The administration of the anti-idiotype polypeptide may be infusion as described above.

The growth or the size of a primary or metastasized tumor may be inhibited or reduced by administering to a subject in need of the treatment an effective amount of an antibody or an anti-tumor hybrid analogue peptide comprising an effector agent selected from the group consisting of radioisotopes, therapeutic drugs and vaccines, and an anti-tumor polypeptide which specifically binds to an antigen on the surface or in the cytoplasm of a tumor cell or released by the cell, allowing the hybrid polypeptide to reach the tumor and the polypeptide to bind thereto, and administering to the subject an amount of the anti-idiotype polypeptide of the invention effective to bind residual or unbound circulating hybrid analogue peptide to thereby accelerate the clearance of the hybrid polypeptide.

Having now generally described this invention, the same will be better understood by reference to certain specific examples, which are included herein for purposes of illustration only and are not intended to be limiting of the invention or any embodiment thereof, unless so specified.

### EXAMPLES

#### Example 1: Methods Utilized

The procedures utilized herein for the reverse-transcription (RT) of RNAs encoding the variable regions and the subsequent amplification of the cDNAs by the polymerase chain reaction (PCR) have been described (Orlandi, R., et al., "Cloning Immunoglobulin Variable Domains for Expression by the Polymerase Chain Reaction", PNAS (USA) 86:3833-3837 (1989); Coloma, M.J., et al., "Primer Design for the Cloning of Immunoglobulin Heavy-Chain Leader-Fvs from Murine Hybridoma Cells Using the PCR", Bio.Techniques 11:152-156 (1991); Gavilondo-Cowley, J.V., et al., "Specific Amplification of Rearranged Immunoglobulin Fv Genes from Murine Hybridoma Cells", Hybridoma 9:407-417 (1990)).

Total RNA is an adequate substrate for RT-PCR. Polyadenylated RNA was utilized herein, however, because it contains only minor levels of contaminating ribosomal RNA and practically no DNA. The polyadenylated RNA was isolated with a Fast Track mRNA isolation kit (Invitrogen Corporation, San Diego, CA).

The oligonucleotides were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City, CA). A PCR murine Ig

plementary DNA (Ns, Norwalk, CT).

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) were cultured in serum (FBS), (1) or RPMI:FBS, bacterial growth. supplement as The freezing

, MuLV<sub>R</sub>5'-C, purchased from r primers were ble 12 below.

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Example 4: Cloning of Chimeric BrE-3 Antibody Polydeoxyribonucleotide

Two expression vectors pAG4622 and pAH4604 were utilized herein (Coloma, M.J., et al., "Novel Vectors for the Expression of Antibody Molecules 5 Using Variable Regions Generated by PCR", *J. Immunol. Methods* 152:89-104 (1992)). These were kindly provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). The construction and expression of chimeric genes was performed as described by Coloma et al. (Coloma, M.J., et al. (1992), *supra*).

10 Oligonucleotides were synthesized and used in a PCR mixture to produce variable heavy ( $V_H$ ) and variable light ( $V_L$ ) fragments with the correct ends for insertion into the pAG4622 and pAH4604 expression vectors. Their sequences are shown in Table 13 below.

**Table 13: Synthesized Oligonucleotides**

15 JO16 (senseV<sub>H</sub> leader) - GGG GATATC CACC ATG TAC TTG GGA CTG AAC TAT GTC TTC A  
 (Seq. ID No: 6);  
 JO17 (sense V<sub>I</sub> leader) - GGG GATATC CACC ATG AAG TTG CCT GTT AGG CTG TTG GT  
 (Seq. ID No: 7);  
 20 JO18 (anti-sense JH3) - GGG GCTAGC TGC AGA GAC AGT GAC CAG AGT CC (Seq. ID No: 8);  
 JO19 (anti-sense Jx1) - GGG GTCGACTTAC G TTT GAT TTC CAG CTT GGT GCC TCC A  
 (Seq. ID No: 9).

The original pCR1000 clones were utilized as the starting templates for  
25 the PCR. The new PCR products were cloned back into pCR1000 and their  
sequence confirmed. Correctly modified and amplified fragments were excised  
with either EcoR V and Sal I (for  $V_L$ ) or with EcoR V and Nhe I (for  $V_H$ ). These  
fragments were then ligated into the respective vectors, which had been cut  
open with the appropriate restriction enzymes. Both the vectors and the inserts  
30 were purified from an agarose gel prior to ligation, using the Bio101 GeneClean  
kit (glass beads) (La Jolla, CA).

**Example 5:** Expression of Murine-Human Chimeric BrE-3 Antibody

The  $V_H$  and  $V_L$  regions in the final murine-human chimeric antibody were sequenced once again to verify that their sequences were correct.

The non-producer myeloma cell line SP2/0-Ag14, (ATCC: CRL 1581, Shulman, M., et al., "A Better Cell Line for Making Hybridomas Secreting Specific Antibodies", Nature 276:269-270, (1978)) was transfected, and a chimeric antibody were isolated as described by Coloma, M.J. et al. (1992), with the following modification. The selection was only undertaken for the uptake of hisD by adding 5mM histidinol to the medium and readjusting the pH to 7.4 with

NaOH.

**Example 6: Production of Transfected Hosts**

After ten days, the stable transfected colonies were clearly established at a frequency of approximately  $10^5$ . The colonies were transferred to a normal medium (without histidinol) and the supernatants from stable transfectants were assayed for the presence of the murine-human chimeric BrE-3 antibody. This was done by capturing the secreted murine-human chimeric BrE-3 antibody with a plate-bound goat anti-human- $\kappa$  antibody and developing with goat anti-human- $\gamma$  antibody as described by Coloma et al. with the following modification. The secondary antibody utilized herein was radiolabeled with  $^{125}\text{I}$ .

**Example 7: Confirmation of Murine-Human Chimeric BrE-3 Antibody Expression**

The supernatants were assayed for binding to human milk fat globule (HMFG) as described by Ceriani et al. (Ceriani R.L., et al., Diagnostic Ability of 15 Different Human Milk Fat Globule Antigens in Breast Cancer", Breast Cancer Res. Treat. 15:161-174 (1990)). HMFG was bound to the microtiter plates as described previously (Ceriani R.L., "Solid Phase Identification and Molecular Weight Determination of Cell Membrane Antigens with Monoclonal Antibodies", in: Monoclonal antibodies and functional cell lines. Progress and application, 20 Bechtol, K.B., McKern, T.J., and Kennett, R., Eds., Plenum Press, New York, pp 398-402 (1984)).

Most colony supernatants were positive by both assays. The colonies that secreted the highest level of chimeric antibody into the supernatants, as determined by these assays, were subcloned and subsequently adapted to 25 serum-free medium for the purification of antibody.

**Example 8: Determination of Affinity Constants**

The antibody-antigen affinity constants for the murine-human chimeric antibody anti-human milk mucin and the whole murine antibody were determined by obtaining the reciprocal value of the concentration of competing unlabeled 30 monoclonal antibody giving 50% binding as described by Sheldon et al. (Sheldon, K., et al., "Characterization of Binding of Four Monoclonal Antibodies to the Human Ovarian Adenocarcinoma Cell Line HEY", Biochem. Cell Biol., 65: 423-428, (1987)). The protocol for the assay was as follows.

Microtiter plates (Dynatech, Chantilly, VA) were prepared using 35 successive layers of methylated BSA, glutaraldehyde, anti- $\beta$ -galactosidase and the bacterial fusion protein 11-2 (a hybrid of  $\beta$ -galactosidase and human mammary mucin) as described in Ceriani et al. (Ceriani, R.L., et al., "A Novel

Serum Assay for Breast Cancer Epithelial Antigen Using a Fusion Protein", Anal. Biochem. 201:178-184 (1992). Each well contained 388ng of the 11-2 fusion protein. To each well were added 25 $\mu$ l  $^{125}$ I-BrE-3 (ATCC No. HB 10028) in RIA buffer (10% bovine calf serum, 0.3% triton X-100, 0.05% sodium azide pH7.4, 5 in phosphate buffered saline), and competed with 25  $\mu$ l of either unlabeled murine antibody or murine-human chimeric antibody in RIA buffer at final concentrations in the nanomolar range.

Iodinations were performed with  $^{125}$ I (17 Ci/mg, Nordion International Inc., Kanata, Ontario, Canada). 50 micrograms of monoclonal antibody BrE-3 10 (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described by Ceriani, R.L. and Blank, E.W., (Ceriani, R.L., and Blank, E.W., "Experimental Therapy of Human Breast Tumors with 131I-Labeled Monoclonal Antibodies Prepared Against the Human Milk Fat Globule", Cancer Res. 48:4664-4672 (1988)). When the counts of bound 15 radiolabeled anti-BrE-3 murine antibody were plotted on the Y axis and the logarithm of the nanomolar (nM) concentration of competing unlabeled anti-BrE-3 murine antibody or murine-human chimeric antibody were plotted in the X axis, both curves overlapped within 5% error (Figure not shown). This proves that the variable region's affinity characteristics have been preserved.

20 **Example 9: Affinity Binding Constants for BrE-3 Murine and Murine-Human Chimeric Antibody**

The purified murine-human chimeric BrE-3 antibody and purified murine BrE-3 antibody gave similar competition curves when tested against  $^{125}$ I-labeled murine BrE-3 binding to its antigen. The affinity binding constants of the murine 25 antibody and the murine-human chimeric antibody were determined in independent competition assays as described in Example 8 above. The values of the constants are  $2.68 \times 10^8 \text{ M}^{-1}$  and  $3.75 \times 10^8 \text{ M}^{-1}$  for the hybrid chimeric BrE-3 polypeptides and for the murine BrE-3 antibody, respectively. These values are not distinguishable at a 95% confidence interval.

30 **Example 10: Amplification of cDNAs Encoding BrE-3 Antibody Variable Regions**

The cDNAs that encode the anti-BrE-3 murine immunoglobulin variable domains ( $V_H$  and  $V_L$ ) were prepared by reverse transcription and PCR amplification (RT-PCR) from polyadenylated RNA isolated from  $10^6$  BrE-3 35 hybridoma cells by the following procedure. The JO2, JO3, JO4, JO14 and  $V_H$ 1BACK primers were synthesized, and their sequences shown in Example 3 above. Other primers were purchased from Novagen. With the exception of  $V_H$ 1BACK, which is a framework-specific primer, all sense primers are specific

for the leader peptide region. All anti-sense primers are specific for the constant regions. The degenerate  $\lambda$  chain of the specific primer *Mulg $\lambda$ V<sub>L</sub>3'-1* (from Novagen), was used to isolate the  $\kappa$  chain cDNA clones because of the similarity of the  $\lambda$  and  $\kappa$ . An identical  $\kappa$  chain clone was isolated with primer JO2 which is specific for the  $\kappa$  chain constant domain. The  $V_H$  region cDNA could not be isolated with the available leader peptide primers. Thus, the  $V_H$ 1BACK primer was used, which yielded the  $V_H$  cDNA  $\gamma$ 72. The leader-peptide primer JO14 was then designed by extrapolating from the framework sequence of  $\gamma$ 72, using cataloged nucleotide sequences (Kabat, E.A., et al., "Sequences of Proteins of 10 Immunological Interest", U.S. Dept. Health and Human Services, NIH publication No. 91-3242, 5<sup>th</sup> Edition (1991). After sequential PCR reactions, this new primer yielded the complete  $V_H$  framework cDNA. This information is summarized in Table 14 below.

Table 14: Primer Combinations for PCR Amplification

15	Clone No.	Sense Primers	Antisense Primers
	$V_L$	152 164	<i>Mulg<math>\kappa</math>V<sub>L</sub>5'-C</i> <i>Mulg<math>\kappa</math>V<sub>L</sub>5'-C</i>
20	$V_H$	$\gamma$ 72 1012 1043	$V_H$ 1BACK JO14(1 <sup>st</sup> PCR) JO14(2 <sup>nd</sup> PCR) JO14(1 <sup>st</sup> PCR) ( <i>Mulg<math>\kappa</math>V<sub>H</sub>5'-C</i> + <i>Mulg<math>\kappa</math>V<sub>H</sub>5'-F</i> ) (2 <sup>nd</sup> PCR)
25			JO2 (JO3 or JO4) JO3 JO4 JO3 <i>Mulg<math>\gamma</math>V<sub>H</sub>3'-2</i>

Example 11: Isolation of Amplified BrE-3  $V_L$  and  $V_H$  cDNA and Sequences

30 The PCR products were cloned without prior purification into pCR1000 (Invitrogen) and sequenced in both directions. Clones 152, 164, 1012, and 1043 were isolated independently during different RT-PCR runs. The sequences of  $V_L$  clones 152 and 164 were found to be identical, as were the sequences of the  $V_H$  clones 1012, 1043. The  $V_H$  and  $V_L$  DNA sequences and their derived 35 protein sequences are shown in Tables 15 and 16 below.

Table 15: BrE-3  $V_L$  Nucleotide and Derived Protein Sequences

DNA Sequence
ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG TTG TTC TGG ATT CCT CCT TCC ATC AGT GAT GTT GTG ATG ACC CAA ACT CCA CTC TCC CTG CCT GTC AGT CTT GGA GAT CAA GCT TCC ATC TCT TGC AGA TCT AGT CAG AAC CTT GTC AAC AAC AAT GGA AAC ACC TAT TTA TAT TGG TTC CTG CAG AAG TCA GGC CAG TCT CCA AAG CTC CTG ATT TAT AGG GCT TCC ATC CGA TTT TCT GGG GTC CCA GAC AGG TTC AGT GGC AGT GGA TCA GAG ACA GAT TTC ACA CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT CTG GGA GTT TAT TTC TGC TTT CAA GGT ACA CAT GTT CGG TGG ACG TTC GGT GGA GGC ACC AAG CTG GAA ATC AAA C (Seq. ID No: 10)

  

Amino Acid Sequence
m k i p v r i i v i l f w i p a s i s d1 v v m t o t p l s l p v s l g d o a s i s c r s s o n l v h n n g n t y l y w f l q k s g q s p k l l i y r a s i r f s g v p d r f s g s g s e t d f t l k i s r v e a e d l g v y f c f o g t h v p w i f g g g t k l e i k (Seq. No: 11)

Table 16: BrE-3  $V_H$  Nucleotide and Derived Protein Sequences

DNA Sequence
ATG TAC TTG GGA CTG AAC TAT GTC TTC ATA GTT TTT CTC TTA AAA GGT GTC CAG AGT GAA GTG AAG CTT GAG GAG TCT GGA GGA GGC TTG GTG CAA CCT GGA GGA TCC ATG AAA CTC TCT TGT GCT GCT TCT GGA TTC ACT TTT AGT GAT GCC TGG ATG GAC TGG GTC CGC CAG TCT CCA GAG AAG GGG CTT GAG TGG GTT GCT GAA ATT AGA AAC AAA GGC AAT ATT CAT GCA ACA TAT TAT GAT GAG TCT GTG AAA GGG AGG TTC ACC ATC TCA AGA GAT GAT TCC AAA AGT AGA AGT TAC CTG CAA ATG ATA AGC TTA AGA GCT GAA GAC ACT GGC CTT TAT TAC TGT ACT GGG GAG TTT GCT AAC TGG GGC CAG GGG ACT CTG GTC ACT GTC TCT GCA G (Seq. ID No: 12)

  

Amino Acid Sequence
m y i g i n y v f i v f l l k g v q s e1 v k l e e s g g g l v q p g g s m k l s c a a s g f t f s d a w m d w v r q s p e k g l e w v a e i r n k a n n h a t y y d e s v k g r f t i s r d d s k s r v y l q m i s l r a e d t g l y y c t g e f a n w g q g t l v t v s a (Seq. ID No: 13)

The sequences were interpreted as described by Kabat et al. (1991). The residues that are shown in lower case correspond to PCR primers. The mature chains begin at D1 ( $V_L$ ) and E1 ( $V_H$ ), respectively. The amino-acids that are underlined are those corresponding to the CDRs. The underlined nucleotides indicate joining segments.

The framework and CDR polypeptide segments were identified according to Kabat et al. (1991), supra.  $V_L$  is a group IIx chain. Part of the CDR 3 and all of framework 4 (FR4) are encoded by  $J_{k1}$ .  $V_H$  belongs to group IIIC. CDR 3 and FR4 are encoded by  $J_{H3}$ . Little or nothing remains from an unidentified D minigene.

Thus, the CDR 3 is only 4 amino-acids long.

Example 12: Comparison of cDNA deduced Amino Acid Sequence with Directly Determined N-Terminal Fragment Sequence

5 Table 17 below shows a comparison between the cDNA-derived polypeptide sequence and the polypeptide sequence determined directly from purified murine BrE-3 monoclonal antibody.

Table 17: Comparison of cDNA-Deduced Protein Sequence with Directly Determined N-terminal Protein Sequence

10	VL	cDNA-deduced Protein sequence	DVVMTQTPLSLPVSLGDQASISCRS
	VL		GVVMTQTPLSLPVVLGDQASIIIXRX
15	VH	cDNA-deduced Protein sequence	EVKLEESGGGLVQPGGSMKLSCAAAS
	VH		EVKLEESGGVLVQPGGSMKLSSAAS

20 The murine BrE-3 antibody was reduced with 5% mercaptoethanol, separated on a 10% SDS polyacrylamide gel, and electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA). Amino acid sequencing was performed directly on the immobilized bands by the Biotechnology Instrumentation Facility, University of California, Riverside. The protein sequence given here is the sequencer's best guess.

25 Once the variable region cDNAs were cloned, it was confirmed that, in fact, they encoded the variable regions of the murine BrE-3 antibody and not those of another antibody by comparing the cDNA-derived amino acid sequences of the cloned murine BrE-3 antibody variable region with the N-terminal sequence of the purified anti-BrE-3 murine antibody directly determined by a single run of protein sequencing. The cDNA sequences were shown to be accurate by comparison with two independently reverse transcribed clones.

30 The general agreement between the predicted and the determined amino-acid sequences shows that the cloned cDNAs encode polypeptides of the same class and subclass as the variable regions of the murine BrE-3 antibody. This indicates that the cDNAs encode authentic variable regions. The authenticity of the variable region polypeptide and, therefore, that of the murine-human chimeric BrE-3 antibody is unquestionable given that the variable regions and the chimeric antibody affinity constant are indistinguishable from those of the murine BrE-3 antibody.

Example 13: Construction of Murine-Human Chimeric Antibody Genes

The vectors used were developed by Coloma et al. (Coloma, M.J., et al. (1992), *supra*) and kindly provided by S.L. Morrison (Dept. of Microbiology and 5 Molecular Genetics, UCLA). Both vectors were derived from pSV2 (Mulligan, R.C., and Berg, P., "Expression of a Bacterial Gene in Mammalian Cells", *Science* 209:1422-1427 (1980)), and contain genomic fragments encoding either the 10 heavy or the light chain constant domains. The vectors accept cDNAs that encode the F<sub>v</sub> regions. To ligate the F<sub>v</sub> cDNAs to the vectors, restriction ends were added to the cDNAs in a set of PCR reactions, using the JO16, JO17, JO18 and JO19 primers.

The pAG4622 light chain vector contains the gene for the human  $\kappa$  chain constant region, including the J-C intron. It encodes xanthine-guanine phosphoribosyltransferase or gpt (Mulligan, R.C., and Berg, P., "Selection for 15 Animal Cells that Express the Escherichia Coli Gene Coding for Xanthine-Guanine Phosphoribosyltransferase", *PNAS (USA)* 78:2072-2076 (1981)) as a dominant selectable marker. It accepts the murine VL cDNA between the ribosome binding site (Kozak, M., "Compilation and Analysis of Sequences Upstream from the Translational Start Site in Eukaryotic mRNAs", *Nucleic Acids Res.* 12:857-872 (1984)), which is preceded by the VH promoter from the anti-dansyl 20 murine monoclonal antibody 27.44 (Coloma, M.J., (1992), *supra*), and the J-C intron. The J-C intron contains the  $\kappa$  chain enhancer (Potter, H., et al., "Enhancer-Dependent Expression of Human  $\kappa$  Immunoglobulin Genes Introduced into Murine Prep-B Lymphocytes by Electroporation", *PNAS (USA)* 81:7161-7165 (1984); Emarine, L., et al., "A Conserved Sequence in the 25 Immunoglobulin J Kappa-C Kappa Intron: Possible Enhancer Element", *Nature* 304: 447-449 (1983)).

The pAH4604 heavy chain vector contains the gene for the heavy chain  $\gamma 1$  constant region, but no J-C intron. It encodes histidinol-dehydrogenase or hisD 30 (Hartman, S.C. and Mulligan, R.C. Two Dominant-Acting Selectable Markers for Gene Transfer Studies in Mammalian Cells", *PNAS (USA)* 85:8047-8051 (1988)) as a dominant selectable marker. It accepts the murine V<sub>H</sub> cDNA between the dansyl promoter-ribosome binding site and the constant  $\gamma 1$  gene. The vector 35 also contains an insert that encodes the heavy chain enhancer (Rabbitts, T.H., et al, "Transcription Enhancer Identified Near the Human C mu Immunoglobulin Heavy Chain Gene is Unavailable to the Translocated c-myc Gene in a Burkitt Lymphoma", *Nature* 306:806-809 (1983)).

The new V<sub>H</sub> and V<sub>L</sub> DNA fragments with appropriate restriction ends were integrated into pAH4604 and pAG4622 as described in Example 4 above. The

vectors were then electroporated (together) into SP2/0-Ag14 myeloma cells as described by Coloma et al. (1992), *supra*.

Example 14: Characterization of Murine-Human Chimeric BrE-3 Antibody

5 The supernatants from stable transfecants were assayed for the presence of the murine-human chimeric antibody as described in Examples 6 and 7 above. High producing transfecants were subcloned and subsequently adapted to grow in serum-free medium. The murine-human chimeric antibody produced by the myeloma cell line was then purified from the culture supernatant using a 10 Sepharose 4B-protein A column (Bio-Rad, Richmond, CA) as described in Ey, P.L., et al. (Ey, P.L., et al., "Isolation of Pure IgG1, IgG2a and IgG2b Immunoglobulins from Murine Serum Using Protein A-Sepharose", *Immunochemistry* 15:429-436 (1978)). Antibody disulfide bonds were reduced to separate the light and heavy chains by heating for 10 min at 65° in Laemmli 15 loading buffer containing 5% beta-mercaptoethanol. The separated chains were then chromatographed on a SDS polyacrylamide gel (10%). The reduced murine-human chimeric BrE-3 antibody and murine antibodies were electrophoresed in separate lanes next to 97.4, 66.2, 45.0, 31.0 and 2.5 Kdalton protein markers. 20 Table 18 below shows the apparent molecular weights of the two bands obtained for both.

**Table 18: Apparent Molecular Weights of Light and Heavy Chains of Murine and Chimeric BrE-3 Antibodies**

	Chimeric Antibody		Murine Antibody	
	Heavy Chain (Kd)	Light Chain (Kd)	Heavy Chain (Kd)	Light Chain (Kd)
25	50	30	49	29

The heavy and light chains of the anti-BrE-3 chimeric antibody separate as expected when electrophoresed on a polyacrylamide gel.

Example 15: Tissue Binding Studies with Chimeric BrE-3 Antibody

30 Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was conducted with the murine-human chimeric BrE-3 antibody. A control was stained with the anti-human secondary antibody only. Positive staining resulted from the use of the murine-human chimeric BrE-3 antibody, followed by the anti-human antibody 35 specific binding. (Pictures not shown).

The breast carcinoma tissue sections were stained with the supernatant

of the transfected cells using the Vectastain ABC method (Vector Labs, Burlingame, CA). The tissue stained with the goat anti-human Ig secondary antibody only shows background or non-specific staining of necrotic areas of the tissue section.

5 The tissue stained with murine-human chimeric BrE-3 antibody, followed by the secondary antibody, shows specific staining of the breast carcinoma cells in the breast tissue sections.

**Example 16: Chimeric BrE-3 Antibody Imaging Studies**

10 The anti-BrE-3 murine monoclonal antibody has been shown to be highly effective for imaging and for the radioimmunotherapy of breast cancers. For example, in a pharmacokinetic study of 15 breast cancer patients conducted with an  $^{111}\text{In}$ -MXDTPA-BrE-3 radio-immunoconjugate (anti-BrE-3 antibody), the serum levels were low in most patients, the blood clearance correlated with the circulating antigen and the imaging results showed that about 86% of all sites 15 could be imaged (Liebes, L., et al., "Pharmacokinetics of  $^{111}\text{In}$ -BrE-3 Monoclonal Antibody in Patients with Breast Carcinoma", Proc. Am. Assoc. Cancer Res. 33:216(Abs No. 1292) (1992)). A  $^{90}\text{Y}$ -BrE-3 radioimmunoconjugate having similar pharmacokinetic characteristics and extrapolating the  $^{111}\text{In}$ -BrE-3 dosimetry results provide a superior therapeutic agent, as well.

20 As with many other monoclonal antibodies, however, the clinical applications of the anti-BrE-3 murine antibody, a whole murine antibody, are limited by the HAMA response. A chimeric monoclonal antibody should give a more restricted HAMA response.

**Example 17: Chimeric BrE-3 Antibody Immunogenicity**

25 The BrE-3 variable region murine polypeptides have been cloned without the constant regions to produce less immunogenic polypeptides than the parent murine antibody. It has, moreover, been shown herein that the murine-human chimeric BrE-3 antibody lacking its original murine constant region preserves its antigen binding characteristics.

30 An BrE-3 antibody variable region alone or as a murine-human chimeric antibody also containing a constant human region or fragment thereof would be significantly less immunogenic to humans than the parent murine antibody. The hybrid polypeptide comprising the variable region of the BrE-3 antibody and the constant region of a human antibody was shown to preserve the original binding 35 affinity of the murine antibody. In this hybrid polypeptide, approximately 2/3 of its contiguous non-human immunogenic targets ( $C_L$  and  $C_H$  regions) were entirely replaced by human constant domains.

Example 18: PCR Primers used in First Isolation of Anti-KC-4 cDNAs

The PCR primers were purchased from Novagen (Madison, WI). Their sequences, reproduced from the booklet provided by Novagen, are shown in 5 Table 19 below.

Table 19: PCR Primer Sequences

	Mulg $\kappa$ V $_L$ 5'-C: sense primer mix for kappa leader.
	ACTAGTCGACATGAAGTTGCCTGTTAGGCTGTTGGTGCTG (Seq. ID No: 14)
10	ACTAGTCGACATGGAGWCAGACACACTCCTGYTATGGGT(Seq. ID No: 15)
	ACTAGTCGACATGGATTWCAGGTGCAGATTWTCAGCTTC (Seq. ID No: 16)
	Mulg $\kappa$ V $_L$ 3'-1: antisense kappa constant region.
	CCCAAGCTTACTGGATGGTGGAAAGATGGA (Seq. ID No: 17)
15	Mulg $V_H$ 5'-F: sense primer mix for heavy chain leader.
	ACTAGTCGACATGRACCTTGGGYTCAGCTTGRTTT (Seq. ID No: 18)
	ACTAGTCGACATGAGAGTGCTGATTCTTTGTG (Seq. ID No: 19)
	ACTAGTCGACATGGATTGGCTGATTTTTTATTG (Seq. ID No: 20)
	Mulg $\gamma$ V $_H$ 3'-2: antisense gamma constant region.
20	CCCAAGCTTCCAGGGRCCARKGGATARACIGRTGG (Seq. ID No: 21)

Example 19: Cloning of Murine-Human Chimeric anti-KC-4 Antibody Ribonucleotide

The two expression vectors pAG4622 and pAH4604 described in 25 Example 4 were utilized. Oligonucleotides synthesized and used in a PCR to produce V $_H$  and V $_L$  fragments with the correct ends for insertion into the pAG4622 and pAH4604 expression vectors are shown in Table 20 below.

Table 20: PCR Primers Sequences

30	JO20 - sense kappa leader
	GGG GATATC CACC ATG AAG TTG CCT GTT AGG CTG TTG (Seq. ID No: 22)
	JO21 - antisense JK2
	CCC GTCGACTTAC G TTT TAT TTC CAG CTT GGT CCC CCC T (Seq. ID No: 23)
	JO22 - sense V $_H$ leader
35	GGG GATATC CACC ATG GAC TTT GGG CTC AGC TTG GTT TT (Seq.ID No: 24)
	JO24 - antisense JH3
	CCC GCTAGC TGC AGA GAC AGA GAC CAG AGT cc (Seq. ID No: 25)

The original pCR1000 clones were the starting templates for the PCR and the 40 rest of the procedures as described in Example 4 above.

Example 20: Expression of the anti-KC-4 Chimeric Antibody Gene

15        The  $V_H$  and  $V_L$  regions in the anti-KC-4 murine-human chimeric antibody were sequenced once again to verify that their sequences were correct. The transfection of the non-producer myeloma cell line SP2/0-Ag14, (ATCC: CRL 1581) and isolation of polypeptide was conducted as described in Example 5 above.

20        Example 21: Production of Transfected Hosts

After ten days, stable transfected colonies were clearly established at a frequency of approximately 1/10,000. The colonies were transferred to normal medium and the assays conducted as described in Example 6 above.

25        Example 22: Confirmation of anti-KC-4 Murine-Human Chimeric Antibody Expression

30        The supernatants were assayed for binding to human milk fat globule (HMFG) and the breast epithelial mucin (BEM) as described previously in Example 7 above. HMFG and BEM were bound to the microtiter plates as described previously by Ceriani, R.L. (1990), *supra*. In this radioassay the bound anti-KC-4 chimeric antibody (HMFG and BEM) was detected by anti-human gamma chain conjugated to  $^{125}\text{I}$ . Most colony supernatants were positive by both assays. The colonies that secreted the highest level of chimeric antibody in the supernatants, as determined by these assays, were subcloned.

Example 23: Western Blot

35        75  $\mu\text{l}$  of the culture supernatant was added to 20  $\mu\text{l}$  of 4x Laemmli buffer and 5  $\mu\text{l}$   $\beta$ -mercaptoethanol and the mixture was heated at 65 °C for 15 min., in order to reduce antibody disulfide bonds and, thus, separate heavy from light chains. 20  $\mu\text{l}$  of the treated sample was chromatographed in duplicate lanes on a 10% SDS polyacrylamide gel together with other antibodies that were treated similarly and that were loaded for comparison. Pre-stained size markers (BioRad, Richmond, CA) were also loaded.

40        5        The chromatographed proteins were electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA) in 90% 30 mM CAPS pH11, 10% methanol, for 1 hour at 25 V and at 4°C. The membrane was cut into 2 parts containing identical antibody samples. The 2 membranes were immersed in 20% bovine calf serum in PBS and shaken slowly at room temperature for 10 1 hour 35 min.  $^{125}\text{I}$ -labeled goat anti-human  $\kappa$  chain antibody was added to one membrane and  $^{125}\text{I}$  labeled goat anti-human  $\gamma$  chain antibody to the other

membrane. Antibodies were labeled at a specific activity of approximately 10 mCi/mg using the chloramine T method as described by Ceriani, R.L. and Blank, E.W. (1988), the labeled antibodies were diluted to 4,000 cpm/ $\mu$ l in RIA buffer.

After incubating 3 hours at room temperature the blots were washed 5 twice in TBS for 10 min each time, once in TBST (50 mM TRIS pH7.5, 3 mM EDTA 25 mM NaCl) 10 min and once more in TBS (TBS with 0.05% Tween 20) for 10 min. The membranes were dried and exposed to Kodak XAR film.

Western blot analysis of culture supernatants revealed that three antibody chains were expressed that corresponded to the three antibody chains 10 seen in the original anti-KC-4 murine antibody. These were a heavy chain that stained with goat anti-human  $\gamma$  chain  $^{125}$ I-labeled antibody, and two light chains that stained with goat anti-human  $\alpha$  chain  $^{125}$ I-labeled antibody (Figure not shown).

The treatment of the original anti-KC-4 murine antibody with 15 N-glycosidase F (Boehringer Mannheim GmbH Germany) following the recommendations of the manufacturer, produced a noticeable decrease in the intensity of the "top" light chain and a concomitant increase in the intensity of the bottom light chain (Figure not shown).

The explanation for the existence of an extra light chain is that this chain 20 is glycosylated. Three lines of evidence substantiate this. First, the detection of an asparagine-linked glycosylation site in the amino acid sequence of the light chain. That is the triad NIS (Asn-Ile-Ser) in framework 3. Second, the decrease of the intensity in the putative glycosylated band after treatment with N-glycosidase F, while concomitantly the intensity of the non-glycosylated band 25 was increased. Finally, 2 corresponding light chain bands are seen in the chimeric antibody version. The extra light chain in the chimeric version cannot be a contaminant since it was specifically stained by goat anti-human  $\alpha$  chain antibody. It can only be a product expressed by pAG4622. Thus both light chains must have the same  $V_L$  amino acid sequence and the same human 30 constant region. These observations show that approximately half of the light chains of both the anti-KC-4 murine and chimeric antibodies are glycosylated at the asparagine-linked glycosylation site.

**Example 24: Amplification of cDNAs Encoding anti-KC-4 Antibody F<sub>v</sub> Regions**

35 The cDNAs that encode the anti-KC-4 murine immunoglobulin  $V_H$  and  $V_L$  were prepared as described in Example 9 above from polyadenylated RNA isolated from 100 million KC-4 hybridoma cells. All clones were obtained from independent PCRs. The sequences of the primers are given in Example 19 and

20 above. All primers are specific for either the leader peptide region or for the constant regions. The primer combinations utilized herein are shown in Table 21 below.

Table 21: Primer Combination for PCR Amplifications

	Clone No.	Primer combinations
5	96	Mulg <sub>κ</sub> V <sub>L</sub> 5'-C + Mulg <sub>κ</sub> V <sub>L</sub> 3'-1
	107	Mulg <sub>κ</sub> V <sub>L</sub> 5'-C + Mulg <sub>κ</sub> V <sub>L</sub> 3'-1
	K1	JO20 + JO21
10	66	Mulg <sub>κ</sub> V <sub>H</sub> 5'-F + Mulg <sub>κ</sub> V <sub>H</sub> 3'-2
	209	Mulg <sub>κ</sub> V <sub>H</sub> 5'-F + Mulg <sub>κ</sub> V <sub>H</sub> 3'-2
	H3	JO22 + JO24
	H7	JO22 + JO24

Example 25: Isolation of Amplified anti-KC-4 F<sub>V<sub>L</sub></sub> (V<sub>L</sub>) and F<sub>V<sub>H</sub></sub> (V<sub>H</sub>) cDNA and Sequences

15 The PCR products were cloned, without prior purification, into pCR1000 (Invitrogen) and sequenced in both directions. The V<sub>H</sub> and V<sub>L</sub> DNA sequences and their derived protein sequences are shown in Tables 22 and 23 below.

Table 22: V<sub>L</sub> Nucleotide Sequences anti-KC-4 V<sub>L</sub> (kII-Jk2)

20

ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG ATG TTC TGG ATT CCT  
GCT TCC AGC AGT GAT GTT TTG ATG ACC CAA ACT CCT CTC TCC CTG  
CCT GTC AGT CTT GGA GAT CAA GCC TCC ATC TCT TGC AGA TCT AGT  
CAG AGC ATT GTA CAT AGT ATT GGA AAC ACC TAT TTA GAA TGG TAC  
CTG CAG AAA CCA GGC CAG TCT CCA AAG CTC CTG ATC TAC AAA GTT  
TCC ATC CGA TTT TCT GGG GTC CCA GAC AGG TTC AGT GGC AGT GGA  
TCA GGG ACA GAT TTC ACA CTC ATT ATC AGC AGA GTG GAG GCT GAG  
GAT CTG GGA ATT TAT TAC TGC TTT CAA GGT TCA CAT GTT CCG TAC  
30 ACG TTC GGA GGG GGG ACC AAG CTG GAA ATA AAA C (Seq. ID No: 26)

Table 23: V<sub>H</sub> Nucleotide Sequences anti-KC-4 V<sub>H</sub> (IIIID-9-JH)

35

ATG GAC TTT GGG CTC AGC TTG GTT TTC CTT GTC CTT ATT TTA AAA  
GGT GTC CAG TGT GAA GTG CAG ATG GTG GAG TCT GGG GGA GTG AAG  
CCT GGA CGG TCC CTG AAA CTC TCC TGT GCA GCC TCT GGA TTC GCT  
TTC AGT AGC ATT GCC ATG TCT TGG GTT CGC CAG GAG AAG AGG CTG  
GAG TGG GTC GCA GAA ATT AGT AGT GGT ATT TAC GCC TAC TAT  
40 CAA GAC ACT GTG AGC GGC CGA TTC ACC AGA GAC ATT GCC AAG AAC  
ACC CTG TAC CTG GAA ATG AGC ACT CTG AGG TCT GAG GAC AGC GCC  
ATG TAT TAC TGT GCA AGG GAG GGT ATC CGG GCC TGG TTT GCT TAC  
TGG GGC CAA GGG ACT CTG GTC TCT GTC TCT GCA G (Seq. ID No: 27)

Example 26: Amino Acid Sequences of anti-KC-4  
Chimeric Antibody Fv Regions

5 After the anti-KC-4 F<sub>v</sub> region cDNAs were cloned, and sequenced, and their cDNA-derived amino acid sequence was compared with the N-terminus sequence directly determined by a single run of amino acid sequencing on purified anti-KC-4 antibody. The cDNA sequences were shown to be accurate since in both cases they were identical for clones that were prepared from  
10 independent reverse transcription reactions. This confirms that the cloned cDNAs are authentic anti-KC-4 F<sub>v</sub> regions. The sequences are shown in Tables 24 and 25 below.

Table 24: V<sub>L</sub> Amino Acid Sequences  
anti-KC-4V<sub>L</sub>(kII-Jk2)

15

MKLPVRLLVLMFWIPASSS (Seq. ID No: 28)  
FR1 DVLMTQTPLSLPVSLGQASISC (Seq. ID No: 29)  
CDR1 RSSQSIVHSNGNTYLE (Seq. ID No: 30)  
FR2 WYLQKPGQSPKLLIY (Seq. ID No: 31)  
20 CDR2 KVSIRFS (Seq. ID No: 32)  
FR3 GVPDRFSGSGSGTDFTLNISRVEAEDLGYYC (Seq. ID No: 33)  
CDR3 FQGSHVPYT (Seq. ID No: 34)  
FR4 FGGGTKLEIK (Seq. ID No: 35)

Table 25: V<sub>H</sub> Amino Acid Sequences anti-KC-4V<sub>H</sub> (IIID-D9-JH3)

25

MDFGLSLVFLVLILKGVQC (Seq. ID No: 36)  
FR1 EVQMVESGGGLVKPGGSLKLSCAASGFAFS (Seq. ID No: 37)  
CDR1 SYAMS (Seq. ID No: 38)  
FR2 WVRQSPEKRLEWVA (Seq. ID No: 39)  
CDR2 EISSGGNYAYYQDTVTG (Seq. ID No: 40)  
30 FR3 RFTISRDNAKNTLYLEMSSLRSEDTAMYCAR (Seq. ID No: 41)  
CDR3 EGIPAWFAY (Seq. ID No: 42)  
FR4 WGQGTLVSVSA (Seq. ID No: 43)

The sequences were interpreted as described by Kabat et al. (1991), supra. The residues that are underlined correspond to PCR primers. The mature V<sub>L</sub> and  
35 V<sub>H</sub> chains begin at amino-acids D and E of framework 1 (FR1), respectively. Framework and CDR protein segments were identified according to Kabat et al. (1991), supra. V<sub>L</sub> is a group II  $\kappa$  chain. Part of the CDR 3 and all of the

framework 4 (FR4) are encoded by Jk2.  $V_H$  belongs to group IIId. CDR 3 and FR4 resulted from a genomic recombination involving minigenes D9 and JH3. There is an asparagine glycosylation site in the light chain in FR3. The site reads NIS (Asn Ile Ser).

5    **Example 27:**    Comparison of cDNA-deduced Amino  
Acid Sequence with Directly Determined  
N-Terminal Fragment Sequence

A comparison between the cDNA-derived polypeptide sequence and the amino acid sequence determined directly on the purified anti-KC-4 monoclonal 10 antibody was undertaken. The results are shown in Table 26 below.

**Table 26: Comparison of cDNA-deduced with Directly Determined N-Terminal Amino Acid Sequences**

$V_H$ , cDNA-deduced	FIRST BAND TOP
$V_H$ , Protein sequence	EVQMVESGGGLVKPGGSLKLS (Seq. ID No: 44) EVQMVESGGGLVKPGGXLKLS (Seq. ID No: 45)
$V_L$ , cDNA-deduced	SECOND BAND
$V_L$ , Protein sequence	DVLMTQTPLSLPVSLGDQASI (Seq. ID No: 46) DVLMTQTPLSLPVXXGDQASI (Seq. ID No: 47)
$V_L$ , cDNA-deduced	THIRD BAND
$V_L$ , Protein sequence	DVLMTQTPLSLPVSLGDQASI (Seq. ID No: 48) DVLMTQTPLSLPVSLGDQASI (Seq. ID No: 49)

X    uncertain or alternative calls.

A sample of anti-KC-4 chimeric antibody (approximately 190  $\mu$ g) was reduced with 5% beta-mercaptoethanol (65°C for 15 min.), separated on three lanes of a 10% SDS polyacrylamide gel, and electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA) in 90% 30 mM CAPS pH11. 5    10% methanol, for 1 hour at 25 V and at 4°C. The transferred protein species were stained with Commassie Brilliant Blue. 3 bands were seen in each lane, of which 2 migrated as expected for a heavy and light chain. The third band migrated above the light chain. Amino acid sequencing was performed directly on the immobilized bands by the Biotechnology Instrumentation Facility, 10    University of California, Riverside. The amino acid sequence given here is the sequencer's best guess.

Example 28: Construction of anti-KC-4 Murine-Human Chimeric Antibody Genes

The vector used were described in Example 1 above. Restriction ends were added to the cDNAs in a set of PCR reactions, using primers J020, 21, 5 22, and 24.

The pAG4622 light chain vector and the pAH4604 heavy chain vector were described in Example 12 above.

The new  $V_H$  and  $V_L$  DNA fragments with appropriate restriction ends were integrated into pAH4604 and pAG4622 as described in Example 12 above.

10 The vectors were then electroporated (together) also as described in Example 12 above.

Example 29: Tissue Binding Studies

The supernatants from stable transfecants were assayed for the 15 presence of the anti-KC-4 murine-human chimeric antibody as described in Example 13 above. The chimeric antibody secreted in the supernatant bound both HMFG and BEM very strongly. In addition, the supernatants containing 20 anti-KC-4 murine-human chimeric antibody were used to stain human breast carcinoma tissue sections by using the immunoperoxidase immunohistochemical staining technique. The intensity of the staining was comparable to that obtained with the original murine monoclonal antibody. The anti-KC-4 monoclonal antibody is known to bind the human milk fat globule and the breast 25 epithelial mucin. This binding specificity of the anti-KC-4 murine monoclonal antibody was maintained even after the recombinant procedure. The anti-KC-4 chimeric antibody bound very strongly to HMFG and BEM as determined by a 30 radioassay (Ceriani, et al., Breast Cancer Res. Trent. 15:161 (1990)). In addition, the anti-KC-4 chimeric antibody bound several human breast tumors in histopathological sections in a manner comparable to the anti-KC-4 murine monoclonal antibody, as detected by immunostaining described in Example 15 above. This specificity of binding demonstrated the retained binding reactivity of the variable regions of anti-KC-4 murine antibody by the polypeptide of the invention when attached to the human  $F_c$  fragment.

Example 30: Materials and Assays for Epitope Mapping

The specific details of the preparation of materials, cell lines, and 35 techniques employed were disclosed by Peterson et al. (Peterson, J.A., et al., "Molecular Analysis of Epitope Heterogeneity of the Breast Mucin", Breast Epithelial Antigens, Ed. Ceriani, R.L., Plenum Press, NY (1991)), the relevant text of which is incorporated herein by reference.

Overlapping peptide hexamers were synthesized onto the ends of polyethylene pins using an Epitope Scanning Kit (Cambridge Research Biochemicals, Cambridge, UK), which is based on a method originally described by Geysen et al. (Geysen, H.L., et al., "Use of Peptide Synthesis to Probe Vital

5 Antigens for Epitopes to a Resolution of a Single Amino Acid", PNAS (USA) 81:3998-4002 (1984)). The polyethylene pins were arranged in a 8x12 configuration that fits into a 96 well microtiter dish. The pins are supplied with an alanine attached to the ends to which the amino acids are added consecutively using pentafluorophenyl active esters of fluorophenyl-  
10 methyloxycarbonyl (Fmoc)-L-amino acids. Each consecutive overlapping hexamer differs from the previous one by a single amino acid and enough were synthesized to span the entire sequence of the peptide to be tested so that every combination of hexamer was present. Each monoclonal antibody was tested for binding to the synthetic peptides using an ELISA method with horseradish  
15 peroxidase-conjugated goat anti-murine IgG (Promega, Madison, WI) and color development with 2,2-azino-bis (3-ethylbenzothiazoline- 6-sulfonic acid (Sigma, St. Louis, MO).

20 The hexapeptides starting with A, P, D, and T bind well to the antibodies (Hexamers 1 to 3 and 20), whereas the hexamers starting between these positions did not. The hexamers that were prepared are shown in Table 27 below. The linear amino acid sequence essential for its binding to the antigen may be deduced from the hexamer that each monoclonal antibody binds. For  
25 example, the anti-BrE-3 antibody required the sequence TRP within the hexamer. Other monoclonal antibodies required other amino acid sequences (e.g., anti-Mc5, TRPAP; anti-Mc1, DTR; anti-BrE-1, DTRPI). BrE-2 also required TRP but its different specificity for normal and tumor tissue indicates that its epitope on the native antigen is different from BrE-3.

Table 27: Epitope Mapping of Repeat Peptide Breast Mucin

Hexamer	G V T S A	P D T R P A P G S T A P P A H G V T S	A P D T R P
1		P D T R P A	
2		D T R P A P	
3		T R P A P G	
4		R P A P G S	
5		P A P G S T	
6		A P G S T A	
7		P G S T A P	
8		G S T A P P	
9		S T A P P A	
10		T A P P A H	
11		A P P A H G	
12		P P A H G V	
13		P A H G V T	
14		A H G V T S	
15		H G V T S A	
16		G V T S A P	
17		V T S A P D	
18		T S A P D T	
19		S A P D T R	
20		A P D T R P	

Example 31: Epitope Mapping

5        Five different monoclonal antibodies (Mc1, Mc5, BrE1, BrE2 and BrE3) were prepared using HMFG for immunization. All identified epitopes on the highly glycosylated large molecular weight breast mucin. By immunohistochemistry they appeared to recognize different epitopes since each had different tissue and tumor specificities (Peterson, J.A., et al., "Biochemical and Histological 10      Characterization of Antigens Preferentially Expressed on the Surface and Cytoplasm of Breast Carcinomas Cells Identified by Monoclonal Antibodies Against the Human Milk Fat Globule", Hybridoma 9:221-235 (1990)). Each monoclonal antibody bound to a different spectrum of normal tissues and their specificities for different carcinomas were different. Anti-BrE2 and anti-BrE3, 15      however, were quite similar. In addition, by screening breast Agt11 cDNA expression libraries with some of these monoclonal antibodies, cDNA clones were isolated that produced fusion proteins that bound all of them, while other cDNA clones bound just some (Larroca, D., et al., "High Level Expression in E. Coli of an Alternate Reading Frame of pS2 mRNA that Encodes a Mimotope of 20      Human Breast Epithelial Mucin Tandem Repeat" Hybridoma 11(2):191-201 (1992)). This binding to the fusion proteins indicated that the epitopes for these five monoclonal antibodies included the polypeptide portion of this glycoprotein. To confirm this the binding of these monoclonal antibodies to two synthetic 25      polypeptide 20-mers (P D T R P A P G S T A P P A H G V T S A and A P P A H G V T S A P D T R P A P G S T) that spanned the tandem repeat consensus sequence was tested (Gendler, S.J., et al., "Cloning of Partial cDNA Encoding

Differentiation and Tumor-Associated Mucin Glycoproteins Expressed by Human Mammary Epithelium", PNAS (USA) 84:6060-6064 (1987); Siddiqui, J., et al., "Isolation and Sequencing of a cDNA Coding for the Human DF3 Breast Carcinoma-Associated Antigen", PNAS (USA) 85:2320-2323 (1988)).

5 One was started at the beginning of the published 20 amino acid repeat (Gendler, S.J., et al. (1987), *supra*) unit, and the other was started in the middle. All five monoclonal antibodies bound to both synthetic peptides, as did DF3, a 10 monoclonal antibody against breast carcinoma cells produced by others (Hull, S.R., et al., "Oligosaccharide Differences in the DF3 Sialomucin Antigen from Normal Human Milk and the BT-20 Human Breast Carcinoma Cell Line", Cancer Comm. 1:261-267 (1989)). Three other monoclonal antibodies against other 15 components of the HMFG that do not cross-react with the breast mucin, Mc13, against a 70 KDa glycoprotein, and Mc3 and Mc8, against a 46 KDa glycoprotein do not bind to these synthetic peptides (data not shown) (Ceriani, R.L., et al., "Characterization of Cell Surface Antigens of Human Mammary Epithelial Cells with Monoclonal Antibodies Prepared Against Human Milk Fat Globule", Somat. Cell Genet. 9:415-427 (1982); Peterson, J.A., et al., "Biochemical and Histological Characterization of Antigens Preferentially Expressed on the Surface and Cytoplasm of Breast Carcinoma Cells Identified by 20 Monoclonal Antibodies Against the Human Milk Fat Globule", Hybridoma 9:221-235 (1990)).

Example 32: Approach for Humanization of Antibodies

25 The present humanization approach is based on Padlan, E.A., "Choosing the Best Framework to Use in the Humanization of an Antibody by CDR-Grafting: Suggestions from 3-D Structural Data", Antibody Engineering 2nd. Annual Conf. San Diego, CA (Dec 16-17, 1991).

30 The fine specificity may be preserved in a "humanized" antibody only if the CDR structures, their interaction with each other, and their interaction with the rest of the variable domains can be maintained. (Padlan, E.A.(1991), *supra*). This requires the preservation of residues of the FR amino acids which contact the CDRs, those which are involved in the  $V_L$ - $V_H$  contact, and those which are buried and could influence the overall domain structure and the structure of the combining site.

35 By examination of murine Fab structures, for which atomic coordinates are available, the FR amino acids that are probably "important" in maintaining the structure of the combining site may be determined (Padlan, E.A., 8th International Congress of Immunol., Budapest, Hungary, Abstracts p. 19 (August 2-28, 1992)).

The specificity of an antibody depends on the CDR structures and sometimes, on some of its neighboring residues as well. These CDR structures, in turn, depend on contacts with framework amino acids and on the interaction of the  $V_L$  and  $V_H$  domains. Thus, to ensure the retention of binding affinity, not only the CDR residues must be preserved, but also those FR residues that contact either the CDRs or their opposite domains, as well as all buried residues, which give shape to the variable domains. The buried amino acids are placed in exactly the same positions in human and in murine frameworks (Padlan, E.A., "A Possible Procedure for Reducing the Immunogenicity of Antibody Variable Domains While Preserving Their Ligand-Binding Properties", *Molecular Immunology* 28:489-498 (1991)).

This approach was applied to design humanized analogues of the variable regions of the murine antibodies of the invention. The humanization or design of the exemplary analogue peptide provided herein was undertaken as follows. The identification of the residues, which are most probably "important" in preserving the combining site structure, permits the selection of the best human FR sequences to use in the "humanization" of each chimeric antibody of known structure or analogues peptides of the invention. The results of the analysis can be used also to predict which FR amino acids should probably be retained in those cases where no three-dimensional structural data are available.

The present procedure was designed to reduce the immunogenicity of the xenogeneic antibodies by preparation of their chimeric derivatives or fragments thereof while preserving their antigen-binding properties. In general, the antigen binding properties of an antibody are primarily determined by its CDRs. The CDRs of the murine antibody were "grafted" herein onto a human framework. In addition, the FR amino acids in the antibody, that are judged as probably important in maintaining the combining-site structure, may also be retained in the humanized molecule.

**30      Example 33:      Choice of a Murine Model of Known Structure**  
**for the Humanization of the BrE-3 Antibody**

The classification of the  $V_H$  and  $V_L$  domains of an antibody such as the anti-BrE-3 antibody was done according to Kabat, E.A., et al. (Kabat, E.A., et al., "Sequences of Proteins of Immunological Interest" NIH (1991)). The anti-BrE-3 kappa chain  $V_L$  domain belongs to group II and the  $V_H$  domain belongs to group IIIc. A murine antibody was then found, whose structure had been determined, and whose variable regions belong to the same classes. The anti-fluorescyl murine antibody 4-4-20 shown in Table 1 above (Herron, J.N., et al., "Three-Dimensional Structure of a Fluorescein-Fab Complex Crystallized in 2-Methyl-2,4-

Pentanediol", Proteins, 5:271-280 (1989) fits these requirements since, like BrE-3, it has  $V_L$  and  $V_H$  domains belonging to groups II and IIIc. Thus, the three-dimensional structures of antibodies BrE-3 and 4-4-20 should be similar, and BrE-3 may be modeled after 4-4-20.

5      **Example 34:      Choice of Target Human Framework**  
          **for Humanization of BrE-3 Antibody**

The choice of the target human framework was not based on the similarity of the amino acid sequence of the entire framework, but strictly on the similarity at the residues that were judged to be structurally important according 10 to the 4-4-20 model. That is, only amino acids that could be involved in contacts with CDR of the opposite chain, or amino acids whose side-chains were predicted to be inwardly pointed. The positions of these amino acids are shown in Tables 8 and 9 and also in Tables 2, 3, 4, 5 and 6 above. These positions are as follows.

15      For the light chain variable region framework: 1, 2, 3, 4, 5, 6, 11, 13, 19, 21, 23, 35, 36, 37, 38, 44, 45, 46, 47, 48, 49, 58, 60, 61, 62, 69, 71, 73, 75, 78, 82, 86, 88, 98, 102, and 104.  
          For the heavy chain variable region framework: 4, 6, 12, 18, 20, 22, 24, 27, 28, 29, 30, 36, 37, 38, 39, 40, 45, 46, 47, 48, 49, 66, 67, 68, 69, 71, 73, 76, 78, 80, 82c, 86, 88, 90, 91, 92, 93, 94, 103, 107, 109 and 111.

The numbering system is conventionally accepted (Kabat, et al. (1991), supra) and shown in Tables 10 and 11 above. In this case, the consensus sequences of all human F<sub>v</sub> regions were selected as the target human framework 25 to minimize the immunogenicity of the product.

First, the sequences of the murine variable chains were aligned with consensus sequences from all known human variable region classes (Herron, J.N., (1989), supra) and the number of differences in the amino-acids that must be retained from the murine were scored. The positions of these amino acids 30 were obtained from those of murine monoclonal antibody 4-4-20, which was chosen to model the anti-BrE-3 antibody as shown in Tables 28 and 29 below.

Table 28: Choice of BrE-3 V<sub>L</sub> Target Human Framework

		BrE-3 V <sub>L</sub>					
		CDR1		CDR2		CDR3	
BRE3K		.....	....	....	....	....	....
HuKi-n		DVVMTQTPLSLPVSLGDQASISC	RSSQNLVHN-NGNTLYL	WFLQKSGQSPKLLIY	RASIRFS		
		.IQ....S.SA.V..RVT.T.	.A..S..XXS-ISN.A.	.YQ..P.KA.....	.A..SLE		
HuKi-nt		.I....S.....TP.EP.....	....S.L.SXD.....N	.Y..P....Q....	LV.N.A.		
HuKi-ii		E.L..S.GT.SL.P.ER.TL..	.A..SVSSS.---A	.YQ..P..A.R....	G..S..AT		
HuKi-v		.I....S.D..A....ER.T.N.K..	SVLYSS.NKN.A	.YQ..P.P.....	W..T.E		
identity with the murine sequence							
* the murine residues that are structurally important.							

Table 29: Choice of BrE-3 V<sub>H</sub> Target Human Framework

		BrE-3 V <sub>H</sub>					
		CDR1		CDR2		CDR3	
BRE3VH		.....	....	....	....	....	....
HuHI		EVKLEESGGGLVQPGGSMKLS	CAASGFTFS	DAWMD-	WVROSPEKGLEWVA		
		Q.Q.VQ..AEVKK..A.V.V..K..Y..T	SYAIS-....A.GQ....MG				
HuHII		Q.Q.Q..P..K.SQTLS.T.TV..GSV.	SYXWSWN	I..P.G....IG			
HuHIII		..Q.V.....LR.....	SYA.S-....A.G.....S				
identity with the murine sequence							
* the murine residues that are structurally important.							

Based on these scores, the human frameworks belonging to groups V<sub>H</sub>II and V<sub>H</sub>III were chosen to receive the anti-BrE-3 CDRs plus other important amino acids.

#### Example 35: Identification of BrE-3 Murine-Human Differences

5 The original murine sequences (BrE-3 V<sub>L</sub> or V<sub>H</sub>) were aligned with their closest human (Human KII or HIII) relatives that were chosen after comparing their sequences in Table 28 above. The alignment of these two sequences is shown in Table 30 below. The information in this table is also contained in Table

28 above, but is reproduced here for clarity. The CDRs are not shown, since their sequences were not changed during the humanization process. Thus, Table 30 shows the maximum number of amino acids that can be changed toward the humanization of the anti-BrE-3 antibody, based on the consensus human sequences obtained from the current databases (Kabat, E.A., et al. (1991), supra). If all these positions were to be replaced with the corresponding human amino acids, the corresponding CDR grafted antibody variable regions would be attained.

**Table 30: Corresponding Amino Acid Sequences of VK BrE-3 and Human KII**

	CDR1	CDR2
BrE-3 VK	DVVMQTPLSLPVLGDQASISC	WFLQKSGQSPKLLIY
Human KII	.I....S.....TP.EP.....	.Y...P....Q....
	CDR3	
BrE-3 VK	GVPDRFSGSGSETDFTLKRVEAEDLGVYFC	FGGGTKLEIK
Human KII	.....G.....V...Y.	..Q...V...
	CDR1	
BrE-3 VII	EVKLEESGGGLVQPGGSMKLSCAASGFTS	WVRQSPEKGLEWVA
Human III	..Q.V.....LR.....	....A.G.....S
	CDR2	
BrE-3 VII	RFTISRDDSKSRVYLQMLRAEDTGLYYCTG	
Human III	.....N..NTL....N.....AV...AR	
	CDR3	
BrE-3 VII	WGQGTLLTVSA	
Human III	.....S	

Table 31 and 32 below contain the same information as Table 30 above in a different format. They show the numbers of the residues that would have to be changed in order to completely convert the original murine framework completely into a human consensus framework.

**Table 31: BrE-3 V<sub>l</sub> Amino Acid Candidates for Change into Human Consensus Sequences**

V <sub>l</sub>	FY36	EG68	GQ100
VI2			
TS7	SP40	LV83	LV104
ST14	KQ45	FY87	LP15
DE17			
QP18			

Table 32: BrE-3 V<sub>H</sub> Amino Acid Candidates for Change into Human Consensus Sequences

V <sub>H</sub>			
	KQ3	SA40	DN73
5	AS113	EV5	EG42
	SN76	ML18	AS49
	RT77	KR19	VL78
	IN82a	GA88	LV89
10			TA93
			GR94

Example 36: Identification of Important Murine BrE-3 Amino Acids

The "important" murine amino acids that should be preserved were chosen based on the contacts of a particular amino acid with the CDRs, and with 15 the opposite chains and/or whether their side chains are pointing inwardly or outwardly. The positions of these "important" amino acids were determined based on the examination of the known structures of other antibodies. This information is provided in Tables 33 and 34 below.

Table 33: Important V<sub>L</sub> Amino Acid Positions to be Preserved

V <sub>L</sub>		
	V2	CDR contact
	F36	CDR contact
	K45	CDR contact
25	F87	Possible contact with VH
	G100	Possible contact with VH
	L104	Buried

Table 34: Important V<sub>H</sub> Amino Acid Positions to be Preserved

V <sub>H</sub>		
	M18	Buried
	S40	Buried
	A49	CDR contact
35	D73	CDR contact
	S76	Buried
	V78	CDR contact
	L89	Might affect interaction with V <sub>L</sub>
40	T93	CDR contact
	G94	CDR contact

Most of the "important" amino acids were selected on the basis of the structure of antibody 4-4-20 and according to Tables 2, 3, 4, 5, 6, 8 and 9 above. Two important amino acids out of each chain, however, were selected based on more general structural analysis, using other antibody structures. This 5 was done to maximize the chances of conserving ligand binding properties. In particular the preservation of the Leu at 89-H was suggested in order to ensure the maintenance of, the  $V_L$ : $V_H$  contact. Although the residue at 89-H is usually not in contact with  $V_L$  and is only partly buried, it nonetheless contributes to the interface. A Val for Leu replacement at this position could very well create a 10 "cavity" which could affect the contact. An Ile for Leu replacement would probably be fine since these amino acids have essentially the same side chain volume.

Finally, by comparing the position of all amino acids that are candidates for mutation, shown in Tables 31 and 32 above, with those that are "important" 15 and should be preserved, shown in Tables 33 and 34, the final selection of amino acid positions for actual mutation was attained. Any "important" amino acid position was eliminated from the list of candidates. Table 35 below shows the amino acids that were selected for change from murine to human identities to obtain the present humanized analogue.

20

Table 35: Selected Amino Acids for Mutation

	$V_L$	$V_H$
	TS7	KQ3
25	ST14	EV5
	LP15	KR19
	DE17	EG42
	QP18	RT77
	SP40	IN82a
30	EG68	GA88
	LV83	AS113

Example 37: Introduction of Changes in BrE-3 Amino Acid Sequence

35 The changes were done at the DNA level in sequential manner. All but one of the codon mutations were performed using enzymatic inverse PCR (EIPCR), a mutagenesis technique developed by Stemmer and Morris (Stemmer, W.P.C and Morris, S.K., "Enzymatic Inverse Polymerase Chain Reaction: a Restriction Site Independent, Single Fragment Method for High Efficiency 40 Site-Directed Mutagenesis", BioTechniques 13:146-220 (1992)).

First, the entire plasmid, containing the target cDNA was amplified by inverse PCR using terminal mutagenic oligonucleotides. Second, Bsal was used to cut the ends of the incorporated primers. This enzyme cuts at a site that is displaced from its recognition site. Thus, after digestion of the 5 open amplified plasmid with Bsal, the Bsal recognition sequence was removed from the ends of the DNA. The DNA was left with complementary sticky ends and could be closed into a functional plasmid that contains the mutagenized region. The amino acid and DNA sequences of the non-mutated (wild-type) 10 variable light and heavy chains of the anti-BrE-3 antibody are shown in Tables 15 and 16 above. The amino acid sequences of the anti-BrE-3 antibody frameworks and the mutations that were performed for the humanization, the oligonucleotide that was used for the mutagenesis, and the method of mutagenesis are shown in Tables 36 to 44 below.

Table 36: F<sub>VL</sub> FR1 Mutation Sites

Position	FR1	Analogue 1 (8 Changes)	DNA Codon (FR1) (Analogue)	Method for Mutagenesis	Primers
<b>Leader Peptide Not Shown</b>					
	D				
	V				
	V				
	M				
	T				
	Q				
7	T	S	ACT TCT	EIPCR*	(J037,J038)
	P				
	L				
	S				
	L				
	P				
	V				
14	S	T	AGT ACT	EIPCR	(J037,J038)
15	L	P	CTT CCT	EIPCR	(J037,J038)
	G				
17	D	E	GAT GAG	EIPCR	(J037,J038)
18	Q	P	CAA CCA	EIPCR	(J037,J038)
	A				
	S				
	I				
	S				
	C				

\* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 37: F<sub>VL</sub> FR2 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
<b>CDR1 Not Shown</b>					
	W				
	F				
	L				
	Q				
	K				
40	S	P	TCA CCA	EIPCR*	(J039,J040)
	G				
	Q				
	S				
	P				
	K				
	L				
	L				
	I				
	Y				

\* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 38: F<sub>VL</sub> FR3 Mutation Sites

Position	FR3	Analogue 1 (8 Changes)	DNA Codon (FR3) (Analogue)	Method for Mutagenesis	Primers
<b>CDR2 Not Shown</b>					
	G				
	V				
	P				
	D				
	R				
	F				
	S				
	G				
	S				
	G				
	S				
68	E	G	GAG GGG	EIPCR*	(J041,J042)
	T				
	D				
	F				
	T				
	L				
	K				
	I				
	S				
	R				
	V				
	E				
	A				
	E				
83	D				
	L	V	CTG GTG	EIPCR	(J041,J042)
	G				
	V				
	Y				
	F				
	C				

\* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 39: F<sub>VL</sub> FR4 Mutation Sites

Position	FR4	Analogue 1 (10 Changes)	Analogue 2 (6 Changes)	DNA Codon (FR4) (Analogue)	Method for Mutagenesis	Primers
<b>CDR3 Not Shown</b>						
	F		F			
	G		G			
	G		G			
	G		G			
	T		T			
	K		K			
	L		L			
	E		E			
	I		I			
	K		K			

Table 40: F<sub>VH</sub> FR1 Mutation Sites

Position	FR1	Analogue 1 (8 Changes)	DNA Codon (FR1) (Analogue)	Method for Mutagenesis	Primers
<b>Leader Peptide Not Shown</b>					
3	E				
	V				
3	K	Q	AAG CAG	EIPCR <sup>†</sup>	(J057,J058)
5	L				
	E	V	GAG GTG	EIPCR	(J057,J058)
	S				
	G				
	G				
	L				
	V				
	Q				
	P				
	G				
	G				
	S				
	S				
19	M				
	K	R	AAA AGA	EIPCR	(J057,J058)
	L				
	S				
	C				
	A				
	A				
	S				
	G				
	F				
	T				
	F				
	S				

<sup>†</sup>EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 41: F<sub>VH</sub> FR2 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
<b>CDR1 Not Shown</b>					
	W				
	V				
	R	Q			
	S				
	P				
42	E	G	GAG GGG	EIPCR*	(J055,J056)
	K				
	G				
	L				
	E				
	W				
	V				
	A				

\* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 42: F<sub>VH</sub> FR3 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
<b>CDR2 Not Shown</b>					
	R				
	F				
	T				
	I				
	S				
	R				
	D				
	D				
	S				
	K				
	S				
77	R	T	AGA ACT	EIPCR*	(J053,J054)
	V				
	Y				
	L				
	Q				
	M				
82	I	N	ATA AAT	EIPCR	(J053,J054)
	S				
	L				
	R				
	A				
	E				
	D				
88	T			EIPCR	(J053,J054)
	G	A	GGC GCC		
	L				
	Y				
	Y				
	C				
	T				
	G				

\* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 43: F<sub>vh</sub> FR4 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
CDR3 Not Shown					
	W				
	G				
	Q				
	G				
	T				
	L				
	V				
	T				
	V				
	S				
113	A	S	GCA TCT	PCR*	(J051,J052)*

\*PCR: Polymerase Chain Reaction

Table 44: Sequences of Mutagenic Oligonucleotides

J037 TCC CTG GGT CTC ACT CCT GGA GAG CCA GCT TCC ATC TCT TGC AGA TCT AGT (Seq. ID No. 50)  
 J038 AGC TTG GGT CTC AGG AGT GAC AGG CAG GGA GAG TGG AGA TTG GGT CAT CAC AAC (Seq. ID No. 51)  
 J039 G TTC CTG GTC TCG CCA CGC CAG TCT CCA AAG CTC CTG (Seq. ID No. 52)  
 J040 T TGG AGG TCT CCC TGG CTT CTG CAG GAA CCA ATA TAA AT (Seq. ID No. 53)  
 J041 TTC ACA GGT CTC ATC AGC AGA GTG GAG GCT GAG GAT GTG GGA GTT TAT TT (Seq. ID No. 54)  
 J042 AGC CTC GGT CTC GCT GAT CTT GAG TGT GAA ATC TGT CCC TGA TCC ACT GC (Seq. ID No. 55)  
 J051 CCT GGA GGA TCC ATG AGA CTC TCT TGT GCT (Seq. ID No. 56)  
 J052 GTT GGG GCT AGC AGA AGA GAC AGT GAC CAG AGT (Seq. ID No. 57)  
 J053 TAC CTG GGT CTC AAT AGC TTA AGA GCT GAA GAC ACT GCC CTT TAT TAC TGT (Seq. ID No. 58)  
 J054 TTC AGC GGT CTC GCT ATT CAT TTG CAG GTA CAC AGT ACT TTT GGA ATC ATC (Seq. ID No. 59)  
 J055 GTC CGC GGT CTC CCA CGG AAG CGG CTT GAG TGG GTT GCT GAA ATT AGA AA (Seq. ID No. 60)  
 J056 CTC AAG GGT CTC CCC TGG AGA CTG GCG GAC CCA GTC CAT CCA GGC ATC A (Seq. ID No. 61)  
 J057 T GAG GAG GTC TCA GGA GGC TTG GTG CAA CCT GGA GGA TCC ATG AGA CTC TCT (Seq. ID No. 62)  
 J058 GG TTG CGG TCT CCC TCC AGA CTC CAC AAG CTG CAC TTC ACT CTG GAC A (Seq. ID No. 63)

\* note Primer set J051 and J052 was originally intended for mutagenizing K19 to R (by J051) and A113 to S (by J052), as described below. Primer J051, however, was somehow defective (it did not run as a single band on a polyacrylamide gel) and thus only the A113 to S mutagenesis was successful. Mutation K19 to R was accomplished at a latter time with EIPCR, using primers J057 and J058.

Example 38: Synthesis of Primers

All primers except J051 and J052 were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City CA). Primers J051 and J052 were purchased from Keystone Laboratories, Inc, Menlo Park, CA.

5 Both, the PCR method used with J051 and J052 and the EIPCR method used with all other primer sets, are described below.

The plasmid DNA template was extracted with a kit purchased from QIAGEN (Tchapsworth, CA) and diluted to 1ng/ $\mu$ l in 10mM TRIS 1mM EDTA pH 7.5 - 8. This plasmid is composed of vector pCR1000 (Invitrogen Corporation, San Diego, CA) into which the cDNA encoding the variable region to be 5 humanized was inserted.

A mixture of PCR primers was made where each primer was present at a concentration of 10 pmole/ $\mu$ l in water.

The PCR amplification conditions were as follows. All reagents as well as the GeneAmp PCR system 9600 were purchased from Perkin Elmer Cetus. 10 Optimal PCR conditions were determined empirically for each pair of mutagenic primers. A matrix of conditions varying the concentration of MgCl<sub>2</sub>, mutagenic primers, and template plasmid DNA was set up. The annealing and extension temperatures during PCR may be varied.

Plasmid template (500 pg/ $\mu$ l), 0.5  $\mu$ M each mutagenic oligonucleotide, 15 1 mM MgCl<sub>2</sub>, 10 mM TRIS pH 8.3, 50 mM KCl, 0.2 mM each nucleotide triphosphate (dGTP, dATP, TTP, dCTP), and Taq polymerase 1 unit/ 20 $\mu$ l reaction mixture.

Example 39: Hot Start PCR

All the components of the PCR mixture, with the exception of Taq 20 polymerase, were mixed in a 95  $\mu$ l volume. The mixture was then dispensed in 19  $\mu$ l aliquots into 5 PCR tubes. The reason for performing five independent reactions was to decrease the odds that unwanted mutations be isolated as a result of nucleotide misincorporation during PCR. The tubes were heated to 95°C for 5 minutes and then cooled to 72°C. While at that temperature 1  $\mu$ l 25 of an appropriate Taq polymerase dilution in 10 mM TRIS, pH 8.3, 50 mM KCl was added to the reaction tubes. The temperature cycling then proceeded as follows.

94°C, 3 min  
[(94°C, 1 min) (50°C, 1 min) (72°C, 3.5 min)] 3 cycles  
30 [(94°C, 1 min) (55°C, 1 min) (72°C, 3.5 min)] 25 cycles  
72°C, 10 min

Example 40: Extra Final Extension

After cycling, the contents of the five tubes were mixed and an extra 35 final extension reaction was carried out. Extra nucleotide triphosphates were added (to 125  $\mu$ M), 5 units of Taq polymerase were also added and the mixture was heated to 72°C for 10 minutes.

Example 41: Purification of PCR Products

The PCR products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5  $\mu$ g/ml Ethidium Bromide. The correct DNA band was visualized with UV light (366 nm), excised from the gel and extracted with the GeneClean kit purchased from Bio 101, La Jolla, CA.

Example 42: Restriction Digestion

The DNA was then digested with Bsal for two hours at 60 degrees celsius in 25  $\mu$ l (20.5  $\mu$ l of DNA, 2.5  $\mu$ l 10 X buffer 4 (NEB) and 2  $\mu$ l BsaI (NEB). BsaI sites were designed near the 5' end of the PCR primers. The primers included 6 extra nucleotides 5' of the BsaI sites to facilitate digestion by BsaI. There were no BsaI sites elsewhere in the plasmid. Other restriction enzymes may be used as advised by Stemmer and Morris (Stemmer and Morris (1992), *supra*). This special class of restriction enzyme cuts at a site that is different from its recognition site but, nevertheless, at a precise distance from it. Using this method, there was no need for having restriction sites in the sequence in order to perform the mutagenesis.

Example 43: Second Purification

The restricted products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5  $\mu$ g/ml Ethidium Bromide. The correct DNA band was visualized with UV light (366 nm), excised from the gel and extracted with the GeneClean kit purchased from Bio 101, La Jolla, CA.

Example 44: Ligation (Reclosure of Plasmid)

The ligation mixtures consisted of 5  $\mu$ l extracted DNA, 2  $\mu$ l 10x ligation buffer (NEB), 1  $\mu$ l T4 DNA polymerase (NEB), 12  $\mu$ l water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation was carried out at room temperature for 2 hrs., or alternatively at 14°C overnight.

Example 45: Transformation and Sequencing

The reclosed plasmids were then transformed into *E. coli*. We used Inv 30 alpha F' competent cells purchased from Invitrogen Corporation, San Diego CA.

Plasmid DNA was then prepared from a few transformants and sequenced to verify that mutagenesis was successful.

Example 46: Mutagenesis of BrE-3 Antibody using J051 and J052

These oligonucleotides were designed to mutagenize K19 to R (by J051)

and A113 to S (by JO52), not by using EIPCR but using the normal PCR (with primers pointing to each other). Primer JO51 carried a BamHI site and primer JO52 carried an Nhel site. After mutagenic PCR amplification the resulting amplified DNA cassette was inserted into the plasmid in lieu of the corresponding 5 wild-type DNA fragment. There was no compelling reason to use this method over the EIPCR method except that conveniently placed restriction sites (BamHI and Nhel) were available. This method, however, yielded only the A113 to S mutation. A subsequent analysis showed that the JO51 primer, which carried the K19 to R mutation ran aberrantly on a polyacrylamide gel.

10 The protocol for the mutagenic amplification step was as follows. Plasmid template (500 pg/ $\mu$ l), 0.75  $\mu$ M each mutagenic oligonucleotide, 2 mM MgCl<sub>2</sub>, 10 mM TRIS pH 8.3, 50 mM KCl, 0.2 mM each nucleotide triphosphate (dGTP, dATP, TTP, dCTP), and Taq polymerase 1 unit/20  $\mu$ l reaction mixture..

15 The PCR was hot started as described for EIPCR above. The temperature cycling conditions used were as follows.

94°C, 3 min

[(94°C, 1 min) (44°C, 1 min) (72°C, 1 min)] 3 cycles

[(94°C, 5 sec) (65°C, 1 min) (72°C, 1 min)] 32 cycles

72°C, 10 min

20 The extra final extension and purification of PCR products were conducted as described for EIPCR above.

25 The restriction digestion of the vector and the insert were conducted as follows. The PCR product was digested with BamHI and Nhel for 1 hr. 50 min. at 37°C [19  $\mu$ l of DNA, 2.5  $\mu$ l 10Xbuffer 3 (NEB), 1.5  $\mu$ l BamHI (NEB), 1.5  $\mu$ l Nhel (NEB). The vector, which is the starting plasmid described above, was digested under similar conditions [1  $\mu$ l plasmid (1  $\mu$ g), 2  $\mu$ l 10Xbuffer 3 (NEB), 1.5  $\mu$ l BamHI (NEB), 1.5  $\mu$ l Nhel (NEB) 14  $\mu$ l water, 2 1/2 hrs. at 37°C].

30 The restricted products, vector and insert, were then purified once more as described above for EIPCR.

35 The ligation of the fragments was conducted as follows. The ligation mixtures consisted of 5  $\mu$ l vector, 5  $\mu$ l insert, 2  $\mu$ l 10x ligation buffer (NEB) 1  $\mu$ l T4 DNA polymerase (NEB), 7  $\mu$ l water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation is carried out at 14°C overnight. A control ligation with vector only was carried out in parallel. The transformation of the host cells was conducted as described for EIPCR.

Example 47: Plasmid Preparation and Sequencing of Humanized BrE-3 V<sub>H</sub>

Plasmid DNA was then prepared from several independent transformants. A few of the plasmids that were shown by restriction analysis to contain the insert were then sequenced to verify that the mutagenesis was successful. When one of the sequenced DNAs contained the desired mutation 5. it was utilized for the next mutation cycle. The fully mutated humanized analogue BrE-3 DNA sequences for the  $V_h$  and  $V_l$  segments are shown in Tables 45 and 46 below.

**Table 45: BrE-3 Antibody  $V_l$  Humanized Analogues DNA Sequences  
BrE-3  $V_l$  FR-HZ**

10

ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG TTG TCC ATT CCT GCT TCC ATC AGT GAT GTT GTG  
ATG ACC CAA TCT CCA CTC TCC CTG CCT GTC ACT CCT GGA GAG CCA GCT TCC ATC TCT TGC AGA TCT  
AGT CAG AAC CTT GTC CAC AAC AAT GGA AAC ACC TAT TTA TAT TGG TTC CTG CAG AAG CCA GCC CAG  
TCT CCA AAG CTC CTG ATT TAT AGG GCT TCC ATC CGA TTT TCT GGG GTC CCA GAC AGG TTC AGT GGC  
15 AGT GGA TCA GGG ACA GAT TTC ACA CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT GTG GGA GTT TAT  
TTC TGC TTT CAA GGT ACA CAT GTT CGG TGG AGG TTC GGT GGA GGC ACC AAG CTG GAA ATC AAA C  
(Seq. ID No. 64)

**Table 46: BrE-3 Antibody VH Humanized Analogue DNA Sequences**

**BrE-3 VH FR-HZ**

20 ATG TAC TTG GGA CTG AAC TAT GTC TTC ATA GTT TTT CTC TTA AAA GGT GTC CAG AGT GAA GTG CAG  
CTT GTG GAG TCT GGA GGA GGC TTG GTG CAA CCT GGA GGA TCC ATG AGA CTC TCT TGT GCT GCT TCT  
GGA TTC ACT TTT AGT GAT GCC TGG ATG GAC TGG GTC CGC CAG TCT CCA GGG AAG GGG CTT GAG TGG  
GTT GCT GAA ATT AGA AAC AAA GCC AAT AAT CAT GCA ACA TAT TAT GAT GAG GCT TGT AAA GGG AGG  
TTC ACC ATC TCA AGA GAT GAT TCC AAA AGT ACT GTG TAC CTG CAA ATG AAT AGC TTA AGA GCT GAA  
25 GAC ACT GCC CTT TAT TAC TGT ACT GGG GAG TTT GCT AAC TGG GGC CAG GGG ACT CTG GTC ACT GTC  
TCT TCT G (Seq. ID No.: 65)

**Example 48: Humanized BrE-3 Antibody Expression**

Two expression vectors pAG4622 and pAH4604 (Coloma, M.J., et al. (1992), *supra*) were used that were developed and provided by S.L. Morrison 30 (Dept. of Microbiology and Molecular Genetics, UCLA). Any cDNA encoding a signal peptide and either the variable heavy chain or the variable light chain can, in principle, be inserted into these vectors resulting in a construction that encodes an IgG1, K, antibody with human constant regions. Correctly modified cDNAs were excised from pCR1000 with EcoRV and Sal I and inserted into pAG4622. These encode the modified light chain. The wild-type heavy chain 35

was similarly excised from pCR1000 by digestion with EcoRV and *Nhe*I and inserted into pAH4604. The restriction and ligation reactions necessary to accomplish these operations were performed under the conditions stipulated by the enzyme manufacturers (New England Biolabs, Beverly, MA). Both the 5 vectors and the inserts were purified from an agarose gel prior to ligation, using the Bio101 (La Jolla, CA) GeneClean kit (glass beads). The  $V_H$  and  $V_L$  regions in the final constructions were sequenced once again to verify that they are correct. The non-producer myeloma cell line SP2/0-Ag14, ATCC: CRL 1581, (Shulman M., et al. (1978), *supra*) was transfected with both plasmid 10 constructions, and antibody producers were isolated following the recommendations outlined in (Coloma, M.J. et al. (1992), *supra*) except that selection was done only for the uptake of *hisD* (by adding 5 mM histidinol to the medium and readjusting the pH to 7.4 with NaOH). Usually after ten days, stable transfecant colonies were established at a frequency of approximately 15  $10^{-5}$  to  $10^{-4}$ . Colonies were then transferred to normal medium (without histidinol). The culture media were either Dulbecco's modified Eagle's medium (DME): fetal bovine serum (FBS), 90:10, v/v, or a mixture of DME:RPMI:FBS, 45:45:10, v/v/v. Penicillin and streptomycin were added to prevent bacterial growth.

20 The supernatants from stable transfectants were assayed for the presence of the antibodies. This was done by capturing the secreted chimeric antibody with a plate-bound goat anti-human-kappa chain antibody and developing with goat anti-human-gamma chain antibody, essentially as described previously (Coloma, M.J. (1992), *supra*) except that the secondary antibody was 25 radiolabeled with  $^{125}\text{I}$ . The supernatants were also assayed for binding to human milk fat globule (HMFG) as described previously (Ceriani R.L., et al., "Diagnostic Ability of Different Human Milk Fat Globule Antigens in Breast Cancer", Breast Cancer Res. Treat., 15:161-174 (1990)). HMFG is bound to the microtiter plates as described previously (Ceriani, R.L. (1984), *supra*). Usually most colony 30 supernatants were positive by both assays. Colonies that secrete the highest level of antibody in the supernatants, as determined by these assays, were subcloned and subsequently adapted to serum-free medium for the purification of antibody. Serum free medium contains HL-1 supplement as directed by the manufacturer (Ventrex Labs., Portland, ME).

35 Example 49: Half Humanized-Half Chimeric BrE-3 Antibody

A BrE-3 humanized light chain was paired with an anti-BrE-3 non-humanized chimeric heavy chain by co-transfection of SP2/0 myeloma cells with hybrid plasmids carrying the respective DNA sequences and those of a human

F<sub>c</sub>

Example 50: Determination of Affinity Constants for Half Humanized and Fully Humanized BrE-3 Antibodies

The secreted half humanized-half chimeric and fully humanized antibodies were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), *supra*). Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), *supra*) using successive layers of methylated BSA, glutaraldehyde, anti- $\beta$ -galactosidase and the bacterial fusion protein 11-2 (a hybrid of  $\beta$ -galactosidase and human mammary mucin). Each well contained 388 ng of the 11-2 fusion protein. To each well were added 25  $\mu$ l  $^{125}$ I-BrE-3 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % sodium azide pH7.4, in phosphate buffer saline) and compete with 25  $\mu$ l of either unlabeled murine or chimeric antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM. Iodinations were performed with  $^{125}$ I (17Ci/mg, Nordion International). Fifty micrograms of monoclonal antibody BrE-3 (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), *supra*)

Antibody-antigen affinity constants were determined by taking the reciprocal of the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. (1987), *supra*). The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to antigen against the same radiolabeled antibody. Both, the half humanized-half chimeric antibody and the fully humanized antibody competed about as well or better with the anti-BrE-3 murine antibody for the antigen.

Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

Example 51: Histochemical Specificity of Half and Fully Humanized BrE-3 Antibodies

Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain useful affinity for the carcinoma antigens. Breast carcinoma tissue sections were stained with the supernatant of the half humanized/half chimeric and fully humanized transfected cells using

the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies showed strong staining patterns.

**Example 52: Gel Chromatography of Half Humanized-Half Chimeric and Fully Humanized BrE-3 Antibodies**

5       Antibody disulfide bonds were reduced by heating for 10 min at 65°C in Laemmli loading buffer containing 5% beta-mercaptoethanol. The separated chains were then chromatographed on a SDS polyacrylamide gel (10%). Two bands were observed for both antibodies of similar migration pattern as the murine antibody. These data were also confirmed by Western blotting.

10      **Example 53: Deded Amino Acid Sequences of Humanized BrE-3 Variable Light and Heavy Chains**

The amino acid sequences of the light and heavy chains of the analogue humanized antibody are shown in Tables 47 and 48 below. These amino acid sequences may be improved either to increase affinity for the antigen or to 15 decrease immunogenicity in humans. Numerous variants of this sequence may be engineered in accordance with the invention.

**Table 47: Humanized BrE-3 V<sub>L</sub> Analogue Amino Acid Sequence BrE-3 V<sub>L</sub> FR-HZ**

20	Leader	mkipvrllvILFWIPASIS (Seq. ID No: 66)
	FR1	DVVMTOQSPLSLPVTPGEPASISC (Seq. ID No: 67)
	CDR1	RSSQNLVHNNGNTYLY (Seq. ID No: 68)
	FR2	WFLOKPGQSPKLLIY (Seq. ID No: 69)
	CDR2	RASIRFS (Seq. ID No: 70)
25	FR3	GPVDRFSGSGSGTDFTLKISRVEAEDVGVYFC (Seq. ID No: 71)
	CDR3	FQGTHHPWT (Seq. ID No: 72)
	FR4	FGGGTKLEIK (Seq. ID No: 73)

30      **Table 48: BrE-3 V<sub>H</sub> Analogue Amino Acid Sequence BrE-3V<sub>H</sub> FR-HZ**

35	Leader	mylglnyvflVFLLKGVQS (Seq. ID No: 74)
	FR1	EVQLVESGGGLVQPGGSMRLSCAASGFTFS (Seq.ID No: 75)
	CDR1	DAWMD (Seq. ID No: 76)
	FR2	WVRQSPGKGLEWVA (Seq. ID No: 77)
	CDR2	EIRNKANNHATYYDESVKG (Seq. ID No: 78)
	FR3	RFTISRDDSKSTVYLQMNSLRAEDTALYYCTG (Seq. ID No: 79)
	CDR3	EFAN (Seq. ID No: 80)
40	FR4	WGQGTLVTVSS (Seq. ID No: 81)

Example 54: Choice of Murine Model of Known Structure for Humanization of anti-KC-4 Antibody

The classification of the  $V_H$  and  $V_L$  domains of an antibody such as the anti-KC-4 antibody was done according to Kabat et al. (Kabat, E.A., et al., 5 "Sequences of Proteins of Immunological Interest" NIH (1991). The KC-4G3 kappa chain  $V_L$  domain belongs to group II and the  $V_H$  domain belongs to group IIId. A murine antibody was then found, whose structure had been determined, and whose variable regions belong to the same classes. The anti-myohemerythrin peptide antibody B13I2 fits these requirements since, like the 10 anti-KC-4 murine antibody, it has  $V_L$  and  $V_H$  domains belonging to groups II and IIId (Stanfield, R.L., et al., "Crystal Structures of an Antibody to a Peptide and its complex with Peptide Antigen at 2.8 Å", Science 248:712-719 (1990)). Thus, the three-dimensional structures of antibodies the anti-KC-4 and B13I2 15 antibodies should be similar, and the humanization of the anti-KC-4 antibody may be modeled after B13I2.

Example 55: Choice of Target Human Framework for Humanization of Chimeric anti-KC-4 Antibody

The choice of the target human framework was based strictly on the similarity at the residues that were judged to be structurally important according 20 to the B13I2 model. That is, only amino acids that could be involved in contacts with CDRs of the opposite chain, or amino acids whose side-chains were predicted to be inwardly pointed. The positions of these amino acids are shown in Tables 49 below.

Table 49: Important Amino Acid Positions for anti-KC-4 Antibody

25

Light Chain Variable Region Framework

2, 3, 4, 6, 7, 11, 13, 19, 21, 22, 23, 35, 36, 37, 38, 43, 44, 46, 47, 48, 49, 58, 60, 61, 62, 69, 71, 73, 75, 78, 82, 85, 86, 87, 88, 98, 102, 104 and 106.

30

Heavy Chain Variable Region Framework

2, 4, 6, 12, 18, 20, 22, 24, 27, 28, 29, 36, 37, 38, 39, 43, 45, 46, 47, 48, 49, 66, 67, 69, 71, 78, 80, 82, 82c, 86, 88, 90, 91, 92, 93, 94, 103, 107, 109 and 111.

35

The numbering system is conventionally accepted (Kabat, et al. (1991), supra) and is shown in Tables 10 and 11 above. In this case, the consensus sequences of all human  $F_v$  regions were selected as the target human framework to minimize the immunogenicity of the product.

First, the sequences of the murine variable chains were aligned with consensus sequences from all known human variable region classes (Herron, J.N., (1989), *supra*) and the number of differences in the amino-acids that must be retained from the murine were scored. The positions of these amino acids 5 were obtained from those of the B13I2 murine monoclonal antibody, which was chosen to model the humanization of the anti-KC-4 antibody.

Based on these scores, the consensus sequences human frameworks belonging to groups  $V_k$ II and  $V_h$ III were chosen to receive the anti-KC-4 murine antibody CDRs plus other important amino acids.

10 **Example 56:** Identification of Murine-Human  
anti-KC-4 Antibody Differences

The original murine sequences (anti-KC-4  $V_k$  or  $V_h$ ) were aligned with their closest human (Human KII or HIII) relatives that were chosen after comparing their sequences as described in Example 34 above. In the present 15 example, it was intended to be substituted as many amino acids as possible in going from the murine to the humanized variable consensus sequences, leaving the important amino acids intact as described in Examples 55 and 57. The amino acids chosen to be preserved were a subset of those listed above. These 20 were selected by analogy to the B13I2 sequence. The single exception was the glycine (100) residue of the original framework of the variable region of the murine kappa chain, which was retained despite not being encompassed in Table 49 above since it was thought that it might contact the variable domain of the heavy chain. Such contacts were observed in at least three Fab that lack a gly at this position.

25 **Example 57:** Identification of Important Murine  
anti-K-4 Antibody Amino Acids

The "important" murine amino acids were chosen for preservation based on the contacts of a particular amino acid with the CDRs, and with the opposite 30 chains and/or whether their side chains are pointing inwardly or outwardly. The positions of these "important" amino acids were determined based on the examination of the known structures of other antibodies.

Most of the "important" amino acids were selected on the basis of the structure of antibody B13I2 and according to Tables 2, 3, 4, 5, 6, 8 and 9 above. The final selection of amino acid positions for actual mutation was 35 attained by comparing the position of all amino acids that are candidates for mutation with those that are "important" and should be preserved. Any "important" amino acid position was eliminated from the list of candidates. Table 50 below shows the amino acids that were selected for change in the

murine sequence to attain the humanized sequence in the present exemplary analogue.

**Table 50: Anti-KC-4 Murine Antibody Variable Region  
Amino Acids Selected for Mutation**

5	Position	KC-4G3 Murine Identity ---> Consensus Human Identity
Light Chain Variable Region		
10		
	14	S
	17	D
	25	K
	74	N
	83	L
Heavy Chain Variable Region		
15		
	13	K
	19	K
	29	R
	44	R
	74	A
	81	F
	82*	M
	84	M
	89	M
	119	A
	119	S

The change N----> K at position 74 in the variable light chain knowingly eliminated an N-linked glycosylation site, which was present in the original murine monoclonal antibody.

**Example 58: Introduction of Changes in Amino Acid Sequence  
for Humanization of anti-KC-4 Antibody**

The introduction of the changes in the amino acid sequence was not done as described in Example 37 above. Instead the DNA encoding each humanized variable region was synthesized in a single polymerase chain reaction (PCR) using overlapping oligonucleotides in accordance with the method described by Ye et al. (Ye, Q-Z, Jonhson, L.L., and Baragi, V., "Gene Synthesis and Expression in *E. coli* for PUMP, a Human Matrix Metalloproteinase", BBRC 186(1):143-149 (1992)). The sequences of the mutagenic are shown in Table 51 below.

**Table 51: Primers for Humanization of anti-KC-4 Murine Antibody Variable Regions**

J059	CCCCGGATCC	TITAAAAGGT	GTCCAGTGTG	AAAGTCAGAT	GGTGGAG	TCTG	(SEQ. ID No: 82)
5	GAATTCGGGGC	TAGCACTAGA	GACAGTGACC	AGAGTCCCTT	GGCCCCAG	(SEQ. ID No: 83)	
J061	AGTGCAGATG	GTGGAGTCIG	GGGGAGGCCT	AGTGCACCT	GGAGGGTCCC	TGAGACTCTC	
	CTGTGAGGCC	TCTGGATTCG	CTTCAGTAG	CTATGCCATG	T	(SEQ. ID No: 84)	
J062	CTTGATGATG	GGCGTAATTA	CCACCACTAC	TAATTTCTGC	GACCCACTCC	AGCCCCTTCC	
	CTGGAGCTG	CCGAACCCAA	GACATGGCAT	AGCTACTGAA	A	(SEQ. ID. No.: 85)	
10	J063	TAATTACGCC	TACTATCAAG	ACACIGTGAC	GGGGCGATT	ACCATCTCCA	GAGACAATTC
	CAAGAACACC	CTGTACCTGC	AAATGAACAG	TCTGAGGGCT	G	(SEQ. ID. No.: 86)	
J064	CCAGAGTCCC	TTGGCCCCAG	TAAGCAAACC	AGGCCGGAT	ACCGTAGTCC	TCCCTTGCAC	
	AGTAATACAC	GGCCGTGTC	TCAGCCCTCA	GACTGTTCAT	T	(SEQ. ID. No.: 87)	
J073	GGGAAGCTTG	ATATCCACCA	TGAAGTTGCC	TCTTAGGCTG	TTGGTGTGA	TGTTCTGGAT	
15	TCTTGC	(SEQ. ID No.: 88)					
J074	AAAGATTCG	TCGACTTACG	TTTATTTC	AGCTTGGTCC	CCCTCC	GAA CGTGTACGGA	
	ACATGT	(SEQ. ID. No.: 89)					
J075	CTGATGTTCT	GGATTCCTGC	TTCCAGCAGT	GATGTTTGA	TGACCC	AAAC	TCCTCTCTCC
	CTGCCCTGCA	CTCCAGGAGA	CCCAAGCTCC	ATCTCTTGC	(SEQ. ID. No.: 90)		
20	J076	CTGTGGAGAC	TGGCTGGTT	TCTGCAGGTA	CCATTCTAAA	TAGGTG	TTTC
	TACAATGCTC	TGACTAGATC	TGCAAGAGAT	GGAGGGTGGC	(SEQ. ID. No.: 91)		
J078	CGAACCTGTA	CGGAACATGT	GAACCTGAA	ACCAAGTAATA	AATTCC	CACA	TCCTCAGGCT
	CCACTCTGCT	GATCTTGAGT	GTGAAATCTG	TCCCTGATCC	(SEQ. ID. No.: 92)		

25 **Example 59: Synthesis of Primers for Humanization of anti-KC-4 Antibody**

All primers were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City CA) using 40nmole columns, cycle 1:63, with Trityl off. None were purified before use. The EIPCR method was used 30 for preparing all primer sets. Their sequences are shown in Table 51 above.

The plasmid DNA template was extracted with a kit purchased from QIAGEN (Tchapsworth, CA) and diluted to 1ng/ $\mu$ l in 10mM TRIS 1mM EDTA pH 7.5 - 8. This plasmid is composed of vector pCR1000 (Invitrogen Corporation, San Diego, CA) into which the cDNA encoding the variable region to be 35 humanized was inserted.

**Example 60: Synthesis of anti-KC-4 Humanized Heavy Chain Variable Regions**

A mixture of PCR primers was made, where each primer was present at a concentration of 10 pmole/ $\mu$ l in water.

40 Four 101 'mer oligonucleotides (J061, J062, J063 and J064), one 50'mer (J059), and one 49'mer (J060), were used for the synthesis of the humanized variable heavy chain. The oligonucleotides concentrations were

estimated using the formula

$$c = [(A_{260})/30]\mu\text{g}/\mu\text{l}$$

The PCR amplification conditions were as follows. All reagents as well as the GeneAmp PCR system 9600 were purchased from Perkin Elmer Cetus.

5 Optimal PCR conditions were determined empirically for each pair of mutagenic primers. A matrix of conditions varying the concentration of  $\text{MgCl}_2$ , mutagenic primers, and template plasmid DNA were set up as follows. However, the annealing and extension temperatures during PCR may be varied.

10 2 $\mu\text{M}$  primer JO59 150nM each of primers JO61, 62, 63 and 64.  
2 $\mu\text{M}$  primer JO60 200 $\mu\text{M}$  each of dGTP, dATP, TTP, and dCTP.  
10mM KCl 20mM Tris-HCl pH 8.8  
10mM  $(\text{NH}_4)_2\text{SO}_4$  2 units per 100 $\mu\text{l}$  reaction Vent DNA polymerase (New England Biolabs)  
0.1% Triton X-100 polymerase (New England Biolabs)  
6mM  $\text{MgSO}_4$

15 **Example 61: Hot Start PCR for Humanization of anti-KC-4 Antibody**

All the components of the PCR mixture, with the exception of Vent DNA polymerase, were mixed. The mixture was then dispensed in 19  $\mu\text{l}$  aliquots into 5 PCR tubes. The reason for performing five independent reactions was to decrease the odds that unwanted mutations be isolated as a result of nucleotide 20 misincorporation during PCR. The tubes were heated to 95°C for 5 minutes and then cooled to 72°C. While at that temperature 1  $\mu\text{l}$  of an appropriate Vent DNA polymerase dilution in 1 x buffer was added to the reaction mixture (hot start). The temperature cycling then proceeds as follows.

25 [(96°C, 6 sec) (55°C, 10 sec) (72°C, 30 sec)] 3 cycles  
[(96°C, 5 sec) (60°C, 10 sec) (72°C, 30 sec)] 29 cycles  
72°C, 10 min

**Example 62: Extra Final Extension for Humanized anti-KC-4 Antibody DNA**

After cycling, one extra final extension reaction was carried out. Extra 30 deoxyribonucleotide triphosphates (to 125  $\mu\text{M}$ ) and 1 unit of Vent DNA polymerase were added, and the mixture was heated to 72°C for 10 minutes.

The resulting synthetic DNA fragment was digested with Drai and NheI and inserted into the same restriction sites a plasmid construct encoding the corresponding murine heavy chain variable region.

35 **Example 63: Synthesis of anti-KC-4 Humanized Light Chain Variable Regions**

The light chain variable region ( $V_L$ ) genes were synthesized in a similar way as described in Examples 60 to 62 above for the heavy chain variable regions.

Example 64: Purification of Humanized anti-KC-4 PCR Products

The PCR products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5  $\mu$ g/ml ethidium bromide. The correct DNA band were visualized with UV light (366 nm), excised from the gels and extracted with the GeneClean kit (Bio 101, La Jolla, CA).

5

Example 65: Litigation of Humanized anti-KC-4 DNA to Plasmid (Reclosure of Plasmid)

The ligation mixtures consisted of 5  $\mu$ l extracted DNA, 2  $\mu$ l 10x ligation buffer (NEB) 1  $\mu$ l T4 DNA polymerase (NEB), 12  $\mu$ l water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation into a pBluescript KS<sup>+</sup> plasmid (Stratagene) was carried out at room temperature for 2 hrs., or alternatively at 14°C overnight.

10

15

Example 66: Transformation and Sequencing of Humanized anti-KC-4 DNA

The reclosed plasmids were then transformed into E. coli utilizing Inv alpha F' competent cells purchased from Invitrogen Corporation, San Diego CA. Plasmid DNA was then prepared from a few transformants and sequenced to verify that mutagenesis was successful.

20

Example 67: Hybrid Plasmid Preparation and Sequencing

Plasmid DNA was then prepared and sequenced to verify that the mutagenesis was successful. The mutated anti-KC-4 humanized analogue DNA sequences for the V<sub>H</sub> and V<sub>L</sub> segments are shown in Tables 52 and 53 below.

25

Table 52: Humanized anti-KC-4 Antibody V<sub>L</sub> Analogue DNA Sequences

30

35

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anti-KC-4 V<sub>L</sub> FR-HZ

ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG ATG TTC TGG ATT CCT GCT  
TCC AGC AGT GAT GTT TTG ATG ACC CAA ACT CCT CTC TCC CTG CCT GTC  
ACT CCA GGA GAG CCA GCC TCC ATC TCT TGC AGA TCT AGT CAG AGC ATT  
GTA CAT AGT AAT GGA AAC ACC TAT TTA GAA TGG TAC CTG CAG AAA CCA  
GGC CAG TCT CCA CAG CTC CTG ATC TAC AAA GTT TCC ATC CGA TTT TCT  
GGG GTC CCA GAC AGG TTC AGT GGC AGT GGA TCA GGG ACA GAT TTC ACA  
CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT GTG GGA ATT TAT TAC TGC  
TTT CAA GGT TCA CAT GTT CCG TAC ACG TTC GGA GGG GGG ACC AAG CTG  
GAA ATA AAA C (SEQ. ID. No.: 93)

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**Table 53: Humanized anti-KC-4 Antibody  $V_h$  Analogue DNA Sequences**

		anti-KC-4 $V_h$ FR-HZ
5		ATG GAC TTT GGG CTC AGC TTG GTT TTC CTT GTC CTT ATT TTA AAA GGT
		GTC CAG TGT GAA GTG CAG ATG GTG GAG TCT GGG GGA GGC TTA GTG CAG
		CCT GGA GGG TCC CTG AGA CTC TCC TGT GCA GCC TCT GGA TTC GCT TCC
		AGT AGC TAT GCC ATG TCT TGG GTT CGC CAG GCT CCA GGG AAG GGG CTG
10		GAG TGG GTC GCA GAA ATT AGT AGT GGT GGT AAT TAC GCC TAC TAT CAA
		GAC ACT GTG ACG GGC CGA TTC ACC ATC TCC AGA GAC AAT TCC AAG AAC
		ACC CTG TAC CTG CAA ATG AAC AGT CTG AGG GCT GAG GAC ACG GCC GTG
		TAT TAC TGT GCA AGG GAG GAC TAC GGT ATC CCG GCC TGG TTT GCT TAC
		TGG GGC CAA GGG ACT CTG GTC ACT GTC TCT AGT (SEQ. ID. No.: 94)

**Example 68: Expression of Anti-KC-4 Humanized Antibody**

Two expression vectors pAG4622 and pAH4604 (Coloma, M.J., et al. 1992, *supra*) were used that were developed and provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). Any cDNA encoding a signal peptide and either the variable heavy chain or the variable light chain can, in principle, be inserted into these vectors resulting in a construction that encodes an IgG1, K, antibody with human constant regions. Correctly modified cDNAs were excised from pCR1000 with EcoRV and Sal I and inserted into pAG4622. These encode the modified light chain. The wild-type heavy chain was similarly excised from pCR1000 by digestion with EcoRV and NhEl and inserted into pAH4604. The restriction and ligation reactions necessary to accomplish these operations were performed under the conditions stipulated by the enzyme manufacturers (New England Biolabs, Beverly, MA). Both the vectors and the inserts were purified from an agarose gel prior to ligation, using the Bio101 (La Jolla, CA) GeneClean kit (glass beads). The  $V_h$  and  $V_l$  regions in the final constructions were sequenced once again to verify that they were correct. The non-producer myeloma cell line SP2/0-Ag14, ATCC: CRL.1581, (Shulman M., et al. (1978), *supra*) was transfected with both plasmid constructions, and antibody producers were isolated following the recommendations outlined in Coloma et al. (Coloma, M.J. et al. (1992), *supra*) except that selection was done only for the uptake of hisD (by adding 5 mM histidinol to the medium and readjusting the pH to 7.4 with NaOH). Usually after ten days, stable transfectant colonies were established at a frequency of approximately  $10^5$  to  $10^4$ . Colonies were then transferred to normal medium (without histidinol). The culture media were either Dulbecco's modified Eagle's medium (DME): fetal bovine serum (FBS), 90:10, v/v, or a mixture of DME:RPMI:FBS, 45:45:10, v/v/v. Penicillin and streptomycin were added to prevent bacterial growth.

The supernatants from stable transfectants were assayed for the presence of the antibodies. This was done by capturing the secreted chimeric

antibody with a plate-bound goat anti-human-kappa chain antibody and developing with goat anti-human-gamma chain antibody, essentially as described previously (Coloma, M.J. (1992), *supra*) except that the secondary antibody was radiolabeled with  $^{125}\text{I}$ . The supernatants were also assayed for binding to human  
5 milk fat globule (HMFG) as described previously (Ceriani R.L., et al., "Diagnostic Ability of Different Human Milk Fat Globule Antigens in Breast Cancer", *Breast Cancer Res. Treat.*, 15:161-174 (1990)). HMFG is bound to the microtiter plates as described previously (Ceriani, R.I. (1984), *supra*). Usually most colony  
10 supernatants were positive by both assays. Colonies that secrete the highest level of antibody in the supernatants, as determined by these assays, were subcloned and subsequently adapted to serum-free medium for the purification of antibody. Serum free medium contains HL-1 supplement as directed by the manufacturer (Ventrex Labs., Portland, ME).

Example 69: Half Humanized-Half Chimeric anti-KC-4 Antibody

15 An anti-KC-4 humanized light chain was paired with an anti-KC-4 non-humanized chimeric heavy chain by co-transfection of SP2/0-Ag14 myeloma cells with hybrid plasmids carrying the respective DNA sequences and those of a human  $F_c$ . The cell line obtained, HuKC-4V1 was deposited with the ATCC on September 23, 1993 and awarded an accession number HB 11454.  
20 In addition, an anti-KC-4 humanized heavy chain was paired with an anti-KC-4 non-humanized chimeric light chain as described in Example 69 above. The thus obtained cell, HuKC-4V3 was deposited with the ATCC on September 23, 1993 and awarded an accession number HB 11456.

Example 70: Fully Humanized anti-KC-4 Antibody

25 An anti-KC-4 fully humanized antibody was prepared by pairing fully humanized anti-KC-4 light and heavy chains by co-transfection as described in Example 69 above. The cell line, HuKC-4V2, was deposited with the ATCC on September 23, 1993 and awarded the accession number HB 11455.

30 Example 71: Determination of Affinity Constants for Half and Fully Humanized anti-KC-4 Antibodies

The secreted half humanized antibodies and the fully humanized antibody were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), *supra*). Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), *supra*) using successive layers of  
35

methylated BSA, glutaraldehyde, anti- $\beta$ -galactosidase and the bacterial fusion protein 11-2 (a hybrid of  $\beta$ -galactosidase and human mammary mucin). Each well contained 388 ng of the 11-2 fusion protein. To each well were added 25  $\mu$ l  $^{125}$ I-KC-4 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % 5 sodium azide pH 7.4, in phosphate buffer saline) and compete with 25  $\mu$ l of either unlabeled murine or chimeric antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM). Iodinations 10 were performed with  $^{125}$ I (17 Ci/mg, Nordion International). 50  $\mu$ g anti-KC-4 monoclonal antibody (Coulter, Hialeah, FL) were labeled at a specific activity of 10. 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), *supra*).

The antibody-antigen affinity constants were determined by taking the reciprocal of the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. 15 (1987), *supra*). The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to the antigen against the same radiolabeled antibody. The fully humanized antibody was shown to compete about as well or better with the anti-KC-4 murine antibody for the KC-4G3 antigen.

20 Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

Example 72: Histochemical Specificity of Half and Fully Humanized anti-KC-4 Antibodies  
25 Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain the affinity for the KC-4G3 carcinoma antigen of the murine antibody. Breast carcinoma tissue sections were stained with the 30 supernatant of the KC-4 murine and fully humanized transfected cells using the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies showed strong staining patterns.

The following Table 54 shows the results of the immunoperoxidase staining of five human breast carcinomas with either the standard anti-KC-4G3 35 murine or the fully humanized antibodies. Both stained the same tissues at a comparable level.

**Table 54:** Immunoperoxidase Staining of Human Breast Carcinoma Tissue Sections with Murine and Fully Humanized anti-KC-4 Antibodies

	Breast Tumor	Murine Antibody	Fully Humanized Antibody
5	1	++	++
	2	+++	+++
	3	-	-
	4	++	++
	5	+++	+++

10 **Example 73:** Binding to HMFG of Half Humanized and Fully Humanized anti-KC-4 Antibodies  
 Tissue culture supernatants from transfectants of all three anti-KC-4 variants of the humanized antibody were shown to bind the human milk fat globule (HMFG) as determined by radio-immunodetections.

15 **Example 74:** Half Humanized and Fully Humanized anti-KC-4 Antibodies Bind to Goat anti-Human  $\kappa$  or  $\gamma$  Antibodies  
 Tissue culture supernatants from transfectants of all three variants of the anti-KC-4 humanized antibody were shown to bind in sandwich 20 radioimmunodetections to both goat anti-human kappa chain antibody bound to microtiterplate wells (750ng/well), and to radio-iodinated  $^{125}\text{I}$ -labeled goat anti-human gamma chain antibodies.

The results of these sandwich assays demonstrate that both chains of the humanized antibodies indeed possess human kappa and gamma constant 25 regions.

**Example 75:** Deduced Amino Acid Sequences of Humanized anti-KC-4 Variable Light and Heavy Chains

The amino acid sequences of the light and heavy chains of the analogue 30 humanized antibody are shown in Tables 55 and 56 below. The actual amino acid sequences may be varied either to increase affinity for the antigen or to decrease immunogenicity in humans. Numerous variants of this sequence may be engineered in accordance with the invention.

Table 55: Humanized anti-KC-4 Antibody V<sub>L</sub> Analogue Sequence


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anti-KC-4 V<sub>L</sub> FR-HZ

MKLPVRLVL MFWIPASSSD VLMTQTPLSL PVTGEPASI SCRSSQSIVH  
 SNGNTYLEWY LQKPGQSPQL LIYKVSIRFS GVPDRSGSG SGTDFTLKIS  
 5 RVEAEDVGIY YCFQGSHVPY TFGGGTKLEI K (Seq. ID No: 95)

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Table 56: Humanized anti-KC-4 Antibody V<sub>H</sub> Analogue Sequence


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anti-KC-4 V<sub>H</sub> FR-HZ

MDFGLSLVFL VLILKGVQCE VQMVESGGGL VQPGGSLRLS CAASGFAFSS  
 10 YAMSWVRQAP GKGLEWVAEI SSGGNYAYQQ DTVTGRFTIS RDNSKNTLYL  
 QMNSLRAEDT AVYYCAREDY GIPAWFAYWG QGTLTVSS (Seq. ID No: 96)

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Example 76: Half Humanized-Half Chimeric anti-BrE-3 Antibodies

A humanized BrE-3 light chain was paired with a non-humanized chimeric  
 15 BrE-3 heavy chain by co-transfection of SP2/0-Ag14 myeloma cells with hybrid  
 plasmids carrying the respective DNA sequences and those of a human F<sub>c</sub>. The  
 cell line obtained, HuBrE3V1, was deposited with the ATCC on November 10,  
 1993 and awarded an accession number HB \_\_\_\_.

In addition, a humanized BrE-3 heavy chain was paired with a non-  
 20 humanized chimeric BrE-3 light chain as described in Example 69 above. The  
 thus obtained cell, HuBrE3V3 was deposited with the ATCC on November 10,  
 1993 and awarded an accession number HB \_\_\_\_.

Example 77: Fully Humanized BrE-3 Antibody

The fully humanized BrE-3 antibody was prepared by pairing fully humanized  
 25 BrE-3 light and heavy chains by co-transfection as described in Example 69  
 above. The thus obtained cell line, HuBrE3V2, was deposited with the ATCC on  
 November 13, 1992 and awarded the accession number HB 1111200.

Example 78: Half Humanized and Fully Humanized  
 30 BrE-3 Antibodies Bind to Goat  
 anti-Human  $\kappa$  or  $\gamma$  Antibodies

Tissue culture supernatants from transfectants of all three variants of the  
 humanized BrE-3 antibody were shown to bind in sandwich  
 radioimmunodetections to both goat anti-human kappa chain antibody bound to

microtiterplate wells (750ng/well), and to radio-iodinated  $^{125}\text{I}$ -labeled goat anti-human gamma chain antibodies.

The results of these sandwich assays demonstrate that both chains of the humanized antibodies indeed possess human kappa and gamma constant regions.

**Example 79: Binding to HMFG of Half Humanized and Fully Humanized BrE-3 Antibodies**

Tissue culture supernatants from transfectants of all three BrE-3 variants of the humanized antibody were shown to bind the human milk fat globule (HMFG) as determined by radio-immunodetections.

**Example 80: Competition Assays and Determination of Affinity Constants for Half Humanized BrE-3 Antibodies**

The secreted half humanized antibodies were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), *supra*). The fully humanized antibody (HuBrE3 V2) was not purified. Its concentration in the culture supernatant was determined by radioimmunodetection using a plate-bound goat anti-human kappa chain antibody as a capturing antibody and using radiolabeled goat anti-human gamma chain antibody as a detecting antibody. A parallel standard curve of human IgG $\kappa$  was used to determine the unknown concentration of HuBrE3 V2. Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), *supra*) using successive layers of methylated BSA, glutaraldehyde, anti- $\beta$ -galactosidase and the bacterial fusion protein 11-2 (a hybrid of  $\beta$ -galactosidase and human mammary mucin). Each well contained 388 nanograms of the 11-2 fusion protein. To each well were added 25  $\mu\text{l}$   $^{125}\text{I}$ -BrE-3 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % sodium azide pH 7.4, in phosphate buffer saline) and competed with 25  $\mu\text{l}$  of either unlabeled murine or humanized antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM). Iodinations were performed with  $^{125}\text{I}$  (17 Ci/mg, Nordion International). 50  $\mu\text{g}$  BrE-3 monoclonal antibody (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), *supra*).

All humanized versions of the BrE-3 antibody were shown to compete about as well or better with the labeled murine BrE-3 antibody for the antigen as did the murine BrE3 antibody. The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to the antigen against the same radiolabeled antibody. The antibody-antigen affinity constants were determined by taking the reciprocal of

the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. (1987), *supra*).

Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the 5 murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

**Example 81: Histochemical Specificity of Half Humanized BrE-3 Antibodies**

Immunohistochemical staining using the immunoperoxidase technique of 10 consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain the affinity for the carcinoma antigen of the murine BrE-3 antibody. Breast carcinoma tissue sections were stained with the supernatant of cells transfected with the HuBrE3 V1 humanized antibody using the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies 15 showed strong staining patterns.

The following Table 57 shows the results of the immunoperoxidase staining of five human breast carcinomas with either the standard BrE-3 murine or the half humanized HuBrE3 V1 antibody. Both stained the same tissues at a comparable level.

**Table 57: Immunoperoxidase Staining of Human Breast Carcinoma Tissue Sections with Murine and Half Humanized BrE-3 Antibodies**

Breast Tumor	Murine Antibody	Half Humanized Antibodies
1	++	++
2	+++	+++
3	-	-
4	++	++
5	+++	+++

**Example 82: Hybridoma Cell Deposits**

The following cell lines were deposited as present examples of the best mode of the invention. The hybridoma cell lines expressing the murine-human fully chimeric BrE-3 and anti-KC4 antibodies and the fully humanized BrE-3 antibody were deposited with the ATCC on November 13, 1992 under the Budapest Treaty, and have been assigned Accession Nos. HB 11199 (Chimeric BrE-3 A1C10), HB 11201 (Chimeric anti-KC-4 1E8) and HB 11200 (Humanized BrE-3 A1C10). The hybridoma cell line expressing the anti-KC-4 humanized

antibody was deposited with the ATCC on September 23, 1993 and has been assigned Accession No. HB 11455 (Humanized HuKC-4 V2). The half chimeric/half humanized antibody cell lines deposited with the ATCC are as follows: ATCC No. HB 11454 (light humanized/ heavy chimeric chains, HuKC-4V1) and ATCC No. HB 11456 (Heavy humanized/ light chimeric chains, HuKC-4V3) were deposited on September 23, 1993, and ATCC No. HB \_\_\_\_\_ (light humanized/ heavy chimeric chains, BrE3V1) and ATCC No. HB \_\_\_\_\_ (heavy humanized/ light chimeric chains, BrE3V3) were deposited on November 10, 1993 under the Budapest Treaty as examples of the best mode of the invention known to the inventors.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

**WHAT IS CLAIMED AS NOVEL AND UNOBIUS AND  
DESIRED TO BE PATENTED IN LETTERS PATENT IS:**

1. A pure, isolated analogue peptide which selectively binds to an antigen that is present on the surface or in the cytoplasm of a neoplastic cell or that is released by the cell, the peptide being selected from the group consisting essentially of at least one CDR of the light or heavy chain of an antibody of a first species having affinity and specificity for the human mammary fat globule (HMFG) antigen and an antigen found on the surface or the cytoplasm of a neoplastic cell or that is released by the cell; at least one variable region of the light or heavy chains of an antibody of the first species having affinity and specificity for the HMFG antigen and an antigen found on the surface or the cytoplasm of a neoplastic cell or that is released by the cell, wherein about 1 to at least 46 amino acids in the FRs are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids; combinations thereof, wherein each analogue peptide is operatively linked to at least one other variable region peptide or analogue thereof; and mixtures thereof.
2. The anti-neoplastic analogue peptide of claim 1, wherein the antibody of the first species is selected from the group consisting of murine, rat, goat, rabbit, canine, primate, guinea pig, bovine, equine, feline and porcine antibodies.
3. The anti-neoplastic analogue peptide of claim 1, being able to compete with the antibody secreted by the hybridoma cell ATCC No. HB 10028 for the human mammary fat globule antigen.
4. The antineoplastic analogue peptide of claim 1, having an amino acid sequence selected from the group consisting of amino acid Sequence ID Nos: 67 through 73; amino acid Sequence ID Nos: 75 through 81; fragments thereof comprising at least one CDR or one CDR and two flanking regions thereof per chain; combinations thereof, wherein

each analogue peptide is operatively linked to at least one other analogue peptide; and mixtures thereof.

5. A glycosylated analogue peptide selectively binding to an antigen present on the surface or in the cytoplasm of a neoplastic cell or that is released by the cell, comprising the analogue peptide of claim 1; and at least one glycosyl residue operatively linked to the peptide.

6. A composition of matter, comprising the analogue peptide of claim 1; and a carrier.

7. A diagnostic kit for the in vivo therapy of neoplasms, comprising the composition of claim 6 in pharmaceutically-acceptable form; and instructions for its use.

8. The kit of claim 7, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

9. The kit of claim 7, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, DNA monomers, RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers or copolymers.

10. An in vitro carcinoma diagnostic kit, comprising the composition of claim 6; a solid support having operatively linked thereto an antigen which selectively binds to the analogue peptide; and instructions for its use.

11. The kit of claim 10, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

12. The diagnostic kit of claim 10, further comprising anti-constant region immunoglobulins, protein G, protein A, other antibody binding molecules or binding fragments thereof.

13. A hybrid analogue polymer, comprising at least one analogue peptide of claim 1 and at least one effector agent operatively linked to the

peptide; combinations thereof; and mixtures thereof.

14. The hybrid analogue polymer of claim 13, wherein the effector agent is selected from the group consisting of monomers, and non-peptide and peptide polymers other than the constant region of an antibody of the same species.

15. The hybrid analogue polymer of claim 13, wherein the effector agent comprises a radioisotope, an enzyme, or a phosphorescent or a fluorescent label.

16. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, phosphorescent and fluorescent agents, enzymes, DNA and RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers or copolymers.

17. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a non-peptide polymer selected from the group consisting of ester, ether, vinyl, amido, imido, alkylene, arylalkylene, cyanate, urethane, and isoprene polymers, halogenated polymers, DNA and RNA polymers, copolymers thereof, and copolymers thereof with peptide polymers or monomers.

18. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a peptide polymer selected from the group consisting of the constant region of antibodies or fragments thereof, the CDRs or variable regions of antibodies, whole antibodies, Fab and Fab', (Fab')<sub>2</sub>, or antibody fragments of a species other than the first species, analogues thereof, hormones, enzymes, peptide transmitters, combinations thereof, and combination thereof with non-peptide polymers, copolymers or monomers.

19. The hybrid analogue polymer of claim 18, wherein the analogue peptide comprises amino acid sequences of non-human origin; and the peptide polymer comprises at least one constant region of the light or heavy chains of a human antibody or fragments thereof capable

of binding to anti-constant region immunoglobulins, protein G or protein A or other antibody binding molecules.

20. The hybrid analogue polymer of claim 19, being selected from the group consisting of an analogue peptide comprising two heavy and two light chains, each light and heavy chain comprising at least one non-human CDR or analogue variable region peptide and at least one human constant region or fragment thereof; at least one human-non-human analogue Fab, Fab', or (Fab')<sub>2</sub> fragment thereof; fragments thereof; combinations thereof; and mixtures thereof.

21. The hybrid analogue polymer of claim 20, wherein each pair of the analogue heavy and light chains has a different predetermined specificity.

22. The hybrid analogue polymer of claim 13, wherein the analogue peptide and the effector agent are operatively linked by a polymer.

23. The hybrid analogue polymer of claim 13, being selected from the group consisting of at least one CDR or analogue variable region of the heavy chain of a first antibody of the first species or fragment thereof comprising at least one CDR and two flanking regions thereof and a first effector agent; at least one CDR or analogue variable region of the light chain of a second antibody of the first species or fragments thereof comprising at least one CDR and two flanking regions thereof, and a second effector agent, wherein each chain has a predetermined specificity; combinations thereof; and mixtures thereof.

24. The hybrid analogue polymer of claim 13, wherein at least one first pair of light and heavy chains comprising at least one CDR or analogue variable region or fragments thereof is linked to at least one second pair of the analogue light and heavy chains comprising at least one CDR or one analogue variable region or fragments thereof.

25. An anti-carcinoma composition, comprising the hybrid analogue polymer of claim 13; and a carrier.

26. An in vivo carcinoma therapy and diagnosis kit, comprising the anti-carcinoma composition of claim 25 in pharmaceutically-

acceptable form; and instructions for its use.

27. The kit of claim 26, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

28. The kit of claim 27, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, DNA monomers, RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers and copolymers.

29. A diagnostic kit for carcinomas, comprising the anti-carcinoma composition of claim 25; a solid support having operatively linked thereto an antigen which specifically binds to the hybrid analogue peptide; and instructions for its use.

30. The kit of claim 29, further comprising a molecule capable of selectively binding to the hybrid analogue polymer, the molecule having an effector agent operatively linked thereto.

31. The diagnostic kit of claim 29, further comprising anti-constant region immunoglobulins, protein G, protein A, other antibody binding molecules or binding fragments thereof.

32. A method of determining the presence of neoplastic cells in a tissue, comprising contacting a tissue suspected of comprising neoplastic cells with the anti-neoplastic analogue peptide of claim 1 and allowing the analogue peptide to bind to antigen associated with any neoplastic cells present in the tissue to form an analogue peptide-cell antigen complex; and  
detecting the presence of any complexes formed.

33. An in vitro method of determining the presence of neoplastic cells in a tissue comprising the method of claim 32, wherein the tissue is excised from a subject prior to being contacted with the analogue peptide in vitro.

34. An in vitro method of diagnosing a neoplasm comprising the method of claim 33, and comparing the results with a standard cut-off

value determined by comparing results obtained for normal and neoplastic samples, wherein a result above the cut-off value is diagnostic of a neoplasm.

35. The in vitro method of diagnosing a neoplasm of claim 34, conducted by contacting a biological sample obtained from a subject suspected of having a neoplasm with a known amount of the anti-tumor analogue peptide of claim 1 in the presence of a solid supported antigen molecule that is selectively bound by the analogue peptide and allowing the formation of solid supported analogue peptide-antigen molecule complexes and analogue peptide-sample antigen complexes with any neoplastic cell antigen present in the sample; detecting any complex formed between the analogue peptide and the solid supported neoplastic antigen; and comparing the result obtained with the standard cut-off value.

36. An in vivo method of imaging a neoplasm present in a subject, comprising the method of claim 32, wherein a pharmaceutically-acceptable composition comprising an effective amount of the analogue peptide in radiolabeled form to reach the neoplasm and bind thereto is administered to a subject suspected of having a primary or metastasized neoplasm; and the detection is of any binding of the labeled analogue peptide to the antigen on the surface or in the cytoplasm of the neoplastic cells.

37. An in vivo method of diagnosing the presence of a neoplasm in a subject comprising the method of claim 36, and comparing the results with a standard cut-off value determined by comparing results obtained for normal and neoplastic samples, wherein a result above the cut-off value is diagnostic of a neoplasm.

38. A method of inhibiting the growth or reducing the size of a primary or metastasized neoplasm in a subject comprising administering to a subject in need of the treatment an effective amount of the anti-tumor composition of claim 25 in pharmaceutically-acceptable form, wherein the effector agent comprises a therapeutic agent.

39. The method of claim 38, further comprising administering to

the subject a molecule capable of selectively binding to the analogue peptide and having an effector agent operatively linked thereto.

40. A method of purging neoplastic cells from a biological fluid, comprising the method of claim 32: and separating any analogue peptide-cell complexes present from the remainder of the fluid.

41. An ex vivo method of purging neoplastic cells from a biological sample obtained from a subject, comprising obtaining a biological sample from a subject suspected of having a neoplasm; conducting the method of claim 40 with the biological sample; and replenishing the purged biological sample to the subject.

42. An in vitro histochemical method of assessing the presence of neoplastic cells in a tissue, comprising obtaining from a patient suspected of having a neoplasm a tissue sample and preparing therefrom a tissue substrate; contacting the tissue substrate with the analogue peptide of claim 1 and allowing the analogue peptide to bind to any neoplastic cells present therein; and detecting the presence of any complexes formed.

43. A hybridoma cell expressing the analogue peptide of claim 1.

44. The hybridoma cell of claim 43, having the ATCC No. HB 11200.

45. The hybridoma cell of claim 43, having the ATCC No. HB 11455.

46. The hybridoma cell of claim 43, having the ATCC No. HB 11454.

47. The hybridoma cell of claim 43, having the ATCC No. HB 11456.

48. The hybridoma cell of claim 43, having the ATCC No. HB (HuBrE3V1).

49. The hybridoma cell of claim 43, having the ATCC No. HB (HuBrE3V3).

50. A composition, comprising the hybridoma cell of claim 43, and a diluent or carrier.

51. A pure, isolated analogue polydeoxyribonucleotide, comprising an analogue oligodeoxyribonucleotide encoding the analogue peptide of

claim 1.

52. The analogue polydeoxyribonucleotide of claim 51, wherein the analogue oligodeoxyribonucleotide comprises a DNA sequence selected from the group consisting of DNA Sequence ID No: 64; DNA Sequence ID No: 65; fragments thereof encoding at least one CDR or one CDR and two flanking regions per chain; redundant DNA sequences thereof; combinations thereof; and mixtures thereof.

53. A hybrid vector, comprising a vector having the analogue polydeoxyribonucleotide of claim 51 operatively linked thereto.

54. The hybrid vector of claim 53, further comprising a polydeoxyribonucleotide comprising an oligodeoxyribonucleotide encoding an effector peptide, the effector peptide-encoding polydeoxyribonucleotide being operatively linked to the vector.

55. A transfected host cell, carrying the hybrid analogue vector of claim 53.

56. A pure, isolated analogue polyribonucleotide, comprising an analogue oligoribonucleotide encoding the analogue peptide of claim 1.

57. A method of producing an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of neoplastic cell, or is released by the cell, comprising a) cloning the analogue polydeoxyribonucleotide of claim 40 into a vector to form a hybrid vector;

b) transfecting a host cell with the hybrid vector and allowing the expression of the analogue peptide; and c) isolating the analogue peptide or mixtures thereof.

58. The method of claim 57, further comprising site-specifically modifying the codon of at least one amino acid in the framework region of a non-human antibody prior to step a) to obtain the analogue polydeoxyribonucleotide.

59. The method of claim 57, wherein steps a) and b) are conducted by cloning analogue polydeoxyribonucleotides encoding analogue peptides selected from the group consisting of at least one CDR or analogue variable region of the first species or fragments thereof comprising at least one CDR and two flanking regions of the heavy or

light chains of the antibody of the first species; and the method further comprises d) allowing the expressed analogue peptides to interact with one another to form double chains.

60. A method of producing a hybrid analogue peptide comprising an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or to an antigen released by the cell and an effector peptide, the method comprising a) transfecting a host cell with the hybrid analogue vector of claim 54 and allowing the expression of the anti-carcinoma hybrid analogue peptide; and b) isolating the anti-carcinoma hybrid analogue peptide or mixtures thereof.

61. An anti-idiotype polymer, comprising polyclonal antibodies raised against anti-tumor antibodies or the analogue peptide of claim 1; monoclonal antibodies thereof capable of specifically binding the anti-neoplasm antibody; fragments thereof selected from the group consisting of CDRs, Fab, Fab', (Fab')<sub>2</sub>, variable regions, and fragments thereof; analogues thereof selected from the group consisting of antibodies, Fab, Fab', (Fab')<sub>2</sub>, and variable regions wherein about 1 to at least 46 amino acids in the FR are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids; combinations thereof, and combinations thereof with an oligopeptide comprising the amino acid sequence APDTRPAPG or fragments thereof comprising the TRP trimer hexamers thereof comprising the trimer, or tandem repeats thereof, wherein each analogue peptide is operatively linked to at least one other analogue peptide; and mixtures thereof.

62. A hybrid anti-idiotype polymer, comprising the anti-idiotype polypeptide of claim 61; and an effector agent operatively linked to the anti-analogue idiotype polypeptide.

63. An anti-carcinoma vaccine, comprising the anti-idiotype polypeptide of claim 61; and a pharmaceutically-acceptable carrier.

64. The anti-carcinoma vaccine of claim 63, in unit dose form.

65. A anti-tumor vaccination kit, comprising the vaccine of claim 63 and a diluent in separate sterile containers; and instructions for its use.

66. A method of vaccinating against neoplasms comprising administering to a subject an effective amount of the composition of claim 63.

67. The vaccination method of claim 66, wherein the anti-idiotype polypeptide is administered in an amount of about 0.1 to 5000.00  $\mu\text{g}/\text{kg}$  body weight/dose.

68. A method of lowering the serum concentration of a circulating antibody or polypeptide that binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or that is released by the cell, comprising administering to a subject in need of the treatment a pharmaceutically-acceptable composition comprising an amount of the anti-idiotype polypeptide of claim 61 effective to bind the circulating antibody or polypeptide and accelerate its clearance.

69. The method of claim 68, wherein the anti-ideotype peptide is administered in an amount of about 0.01 to 5.000.00 mg/kg body weight/dose.

70. A method of inhibiting the growth or reducing the size of a primary or metastasized carcinoma tumor in a subject, comprising administering to the subject an effective amount of a pharmaceutically-acceptable composition comprising an anti-carcinoma hybrid polymer comprising an effector agent selected from the group consist of radioisotopes and therapeutic drugs, and an anti-carcinoma polypeptide which specifically binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or that is released by the cell; allowing the hybrid polymer to reach the tumor and the anti-carcinoma polypeptide to bind thereto; and administering to the subject a pharmaceutically-acceptable composition comprising an amount of the anti-idiotype polypeptide of claim 61 effective to bind any residual or unbound circulating anti-carcinoma hybrid polypeptide and accelerate the clearance of the hybrid polymer.



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<p>(21) International Application Number: PCT/US93/11445</p> <p>(22) International Filing Date: 16 November 1993 (16.11.93)</p> <p>(30) Priority data: 07/977,696 16 November 1992 (16.11.92) US 08/129,930 30 September 1993 (30.09.93) US 08/134,346 8 October 1993 (08.10.93) US</p> <p>(71) Applicant (for all designated States except US): CANCER RESEARCH FUND OF CONTRA COSTA [US/US]; 2055 North Broadway, Walnut Creek, CA 94596 (US).</p> <p>(72) Inventors; and</p> <p>(75) Inventors/Applicants (for US only) : DO COUTO, Fernando, J., R. [US/US]; 7710 Oak Creek Drive, Pleasanton, CA 94588 (US). CERIANI, Roberto, L. [US/US]; PETERSON, Jerry, A. [US/US]; 1089 Via Roble, Lafayette, CA 94549 (US). PADLAN, Eduardo, A. [US/US]; 4006 Simms Drive, Kensington, MD 20895 (US).</p>		<p>(74) Agent: AMZEL, Viviana; V. Amzel &amp; Assoc., 2055 North Broadway, Walnut Creek, CA 94596 (US).</p> <p>(81) Designated States: AT, AU, BB, BG, BR, CA, CH, DE, DK, ES, FI, GB, HU, JP, KP, KR, LK, LU, MG, MN, MW, NL, NO, PL, RO, RU, SD, SE, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</p> <p>(88) Date of publication of the international search report: 7 July 1994 (07.07.94)</p>	

(54) Title: PEPTIDES AND ANTI-SENSE PEPTIDES WITH BROAD NEOPLASTIC SPECIFICITY

## (57) Abstract

An analogue peptide comprises the variable regions of the light or heavy chains of an antibody of a first species selectively binding to a tumor antigen, such as a carcinoma antigen, has 1 to 46 amino acids of the framework regions per chain substituted with amino acids such as those present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 variable region CDRs per chain and optionally flanking regions thereof of 1 to 10 or more amino acids, alone or with an N-terminal fragment of 1 to 10 or more amino acids, combinations or mixtures thereof. The polypeptide may also comprise an effector agent and/or be glycosylated, and is presented as a composition with a carrier. The analogue peptides are used in diagnostic kits for neoplasms such as carcinomas and methods for in vivo imaging and treating a primary or metastasized neoplasm such as a carcinoma, and in vitro diagnosing a neoplasm, ex vivo purging neoplastic cells from a biological fluid. RNAs and DNAs encode the analogue peptide, and a hybrid vector carrying the nucleotides and transfected cells express the peptides and a method produces the analogue peptide. An anti-idiotype polypeptide comprises polyclonal antibodies raised against an antineoplastic antibody or the analogue peptide of this invention, monoclonal antibodies thereof, Fab, Fab', (Fab')<sub>2</sub>, CDR, variable region, or analogues or fragments thereof, combinations thereof with an oligopeptide comprising a TRP trimer, tandem repeats thereof, or combination or mixtures thereof. An anti-idiotype hybrid polypeptide with an effector agent and the anti-idiotype polypeptide, an anti-neoplastic vaccine, an anti-neoplastic vaccination kit, a method of vaccinating against neoplasms such as carcinomas and a method of lowering the serum concentration of a circulating antibody or polypeptide are provided.

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A. CLASSIFICATION OF SUBJECT MATTER  
 IPC 5 C12N15/13 A61K47/48 C07K15/28 G01N33/574 C12N5/20

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B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
 IPC 5 C12N A61K C07K G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO,A,92 04380 (UNILEVER PLC) 19 March 1992  see the whole document -----	1-3,5,6, 50,51, 53,55-61
X	WO,A,90 12319 (JOHN MUIR CANCER & AGING INSTITUTE) 18 October 1990 see the whole document -----	4
X	WO,A,86 02945 (COULTER CORPORATION) 22 May 1986 see the whole document -----	4
P,X	EP,A,0 534 742 (CELLTECH LIMITED) 31 March 1993 see the whole document -----	1-6, 50-65

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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**INTERNATIONAL SEARCH REPORT**

Int'l application No.  
PCT/US 93/11445

**Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)**

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:  
Remark : Although claims 32, 36-39 and 66-70 are directed to a method of treatment of (diagnostic method practised on) the human body, the search has been carried out and based on the alleged effects of the composition.
2.  Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.  Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

**Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)**

This International Searching Authority found multiple inventions in this international application, as follows:

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

**Remark on Protest**

The additional search fees were accompanied by the applicant's protest.  
 No protest accompanied the payment of additional search fees.